Tuberculosis



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Tuberculosis results in an estimated 1.7 million deaths each year and the worldwide number of new cases (more than 9 million) is higher than at any other time in history. 22 low-income and middle-income countries account for more than 80% of the active cases in the world. Due to the devastating effect of HIV on susceptibility to tuberculosis, sub-Saharan Africa has been disproportionately affected and accounts for four of every five cases of HIV-associated tuberculosis. In many regions highly endemic for tuberculosis, diagnosis continues to rely on century-old sputum microscopy; there is no vaccine with adequate effectiveness and tuberculosis treatment regimens are protracted and have a risk of toxic effects. Increasing rates of drug-resistant tuberculosis in eastern Europe, Asia, and sub-Saharan Africa now threaten to undermine the gains made by worldwide tuberculosis control programmes. Moreover, our fundamental understanding of the pathogenesis of this disease is inadequate. However, increased investment has allowed basic science and translational and applied research to produce new data, leading to promising progress in the development of improved tuberculosis diagnostics, biomarkers of disease activity, drugs, and vaccines. The growing scientific momentum must be accompanied by much greater investment and political commitment to meet this huge persisting challenge to public health. Our Seminar presents current perspectives on the scale of the epidemic, the pathogen and the host response, present and emerging methods for disease control (including diagnostics, drugs, biomarkers, and vaccines), and the ongoing challenge of tuberculosis control in adults in the 21st century.

Introduction

Tuberculosis has plagued humankind worldwide for thousands of years. John Bunyan (Nov 28, 1628-Aug 31, 1688), an English Christian writer and preacher, described tuberculosis as "The Captain among these men of death" at a time when tuberculosis case rates in London had reached 1000 per 100 000 population per year.1 Tuberculosis continued to cause many deaths in London during the 19th century and accounted for up to 25% of deaths in Europe. The death toll from tuberculosis began to fall as living standards (housing, nutrition, and income) improved early in the 20th century, well before the advent of antituberculosis drugs. Despite the first antituberculosis drugs being discovered more than 60 years ago, tuberculosis today still kills an estimated 1.7 million people each year.2 Progress in the scaling up of tuberculosis diagnostic, treatment, and control efforts worldwide over the past decade has been associated with improvements in tuberculosis control in many parts of the world, but progress has been substantially undermined by the HIV-1 epidemic, the growing challenge of drug resistance, and other increasingly important epidemiological factors that continue to fuel the tuberculosis epidemic.3 Greater investment in new technologies, basic science, and translational and applied research has led to progress in the development of improved tuberculosis diagnostics, drugs, treatment regimens, biomarkers of disease activity, and vaccines; new perspectives in the pathogenesis of tuberculosis are also emerging. Our Seminar focuses on tuberculosis in adults and presents current perspectives on the scale of the epidemic, the pathogen and host response, current and emerging methods for disease control (including diagnostics, drugs, biomarkers, and vaccines), and the ongoing challenge of tuberculosis control in the 21st century.

Epidemiology

The estimated total number of incident cases of tuberculosis worldwide rose to 9 · 4 million in 2009—more than at any other time in history. The worldwide tuberculosis incidence rates are estimated to have peaked in 2004 and to have decreased at a rate of less than 1% per year since that time. However, the overall worldwide burden continues to rise as a result of the rapid growth of the world population. Most cases are in Asia and Africa, with smaller proportions of cases in the eastern Mediterranean region, European region, and the Americas (figure 1). 22 countries account for 80% of the worldwide burden and the five countries that rank first to fifth in the world in terms of total numbers of incident cases in 2009, were India, China, South Africa, Nigeria, and Indonesia.

About 12% (1·1 million cases) of the worldwide tuberculosis caseload was HIV-associated and most of

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Search strategy and selection criteria

Our Seminar is focused on tuberculosis in adults. Readers are referred to recent reviews on tuberculosis in children. Our search strategy included a 7-year review of PubMed (2004–11), the Cochrane library (2004–10), WHO and WHO-STOP TB publications (2000–10), and Embase (2004–10), and three recent comprehensive tuberculosis textbooks (Tuberculosis: a comprehensive clinical reference [Philadelphia, PA: Saunders, 2009]; Tuberculosis: the essentials, 4th edn [London: Informa, 2009]; Handbook of tuberculosis—vols 1, 2, and 3 [Hoboken, NJ: John Wiley and Sons Inc, 2008]). The search terms used were "tuberculosis", "Mycobacterium tuberculosis", "tuberculosis" and "HIV", "immunity", "pathogenesis", "clinical features", "diagnosis", "diagnostic tests", "biomarkers", "imaging", "radiology", "treatment", "prevention", "latent infection", "vaccines", "control", "drug-resistant", "extensively drug-resistant", "pregnancy", "vulnerable groups", "prevention", "research priorities", "packages of care". We also included commonly referenced older published work on tuberculosis, and cite so-called state of the art tuberculosis review articles, to provide more details and references than cited in our Seminar.

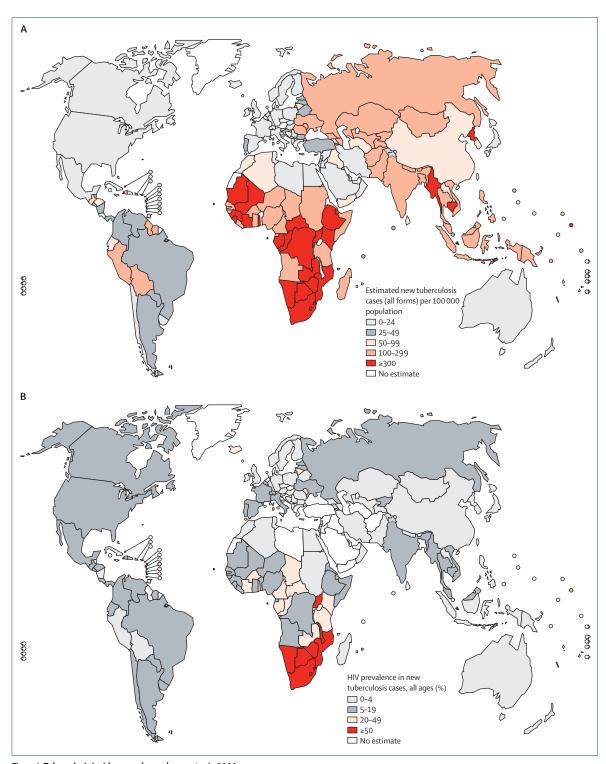


Figure 1: Tuberculosis incidence and prevalence rates in 2009
Estimated tuberculosis incidence rates (A). Estimated HIV prevalence in new tuberculosis cases (B). Reproduced from Global tuberculosis control.

these cases were in sub-Saharan Africa (about four of every five cases) and southeast Asia (figure 1).⁴ HIV has fuelled a three to five times increase in tuberculosis incidence rates in many high HIV prevalence countries

in sub-Saharan Africa (figure 2), especially in the south and east of the continent.²⁵ In the worst affected countries of South Africa and Swaziland, about 1% of the population develops tuberculosis each year, much of it due to HIV.

The collapse of the Soviet Union and the multidrugresistant (MDR) tuberculosis epidemics in marginalised populations such as prisoners and drug abusers have been key factors underlying the increases in tuberculosis incidence rates noted in eastern Europe (figure 2).

Tuberculosis remains a disease of poverty that is inextricably associated with overcrowding and undernutrition.⁶⁷ The table shows a range of other risk factors for tuberculosis. Infection with HIV is the most potent of these risk factors, with the risk of people infected with HIV developing tuberculosis being more than 20-times greater than that of people not infected with HIV.8 Other risk factors include heavy alcohol consumption and smoking; the latter roughly doubles risk of tuberculosis²⁷ and might account for up to half of all deaths in men with tuberculosis in India.14 Diabetes is associated with an about three-times increase in tuberculosis risk (table) and accounted for about 20% of smear-positive tuberculosis cases in India in 2000.28 Immunosuppressive drugs such as corticosteroids have long been associated with the risk of tuberculosis, but tuberculosis associated with tumour necrosis factor (TNF) antagonists for the treatment of rheumatological disorders is now an increasing problem in industrialised countries.26 The evidence for a human genetic contribution to susceptibility to tuberculosis is now growing (table).24 Genetic variants known to affect susceptibility include the natural resistance-associated macrophage protein (NRAMP), the vitamin D receptor (VDR), and the nitric oxide synthase (NOS2A) and interferon-y pathways. Although the effect size for most of these associations is only moderate, the cumulative effect of these polymorphisms to the burden of tuberculosis in different populations could be substantial but remains to be defined.3

Over the past two decades there has been the worldwide emergence of MDR tuberculosis, then extensively resistant (XDR) tuberculosis, and, most recently, strains that are resistant to all antituberculosis drugs. $^{29-32}$ MDR tuberculosis is caused by *Mycobacterium tuberculosis* that is resistant to at least isoniazid and rifampicin, and XDR tuberculosis is caused by MDR tuberculosis strains that are also resistant to any fluoroquinolone and one of three injectable aminoglycosides (capreomycin, kanamycin, and amikacin). It is estimated that there were about $0\cdot 5$ million incident cases of MDR tuberculosis and $50\,000$ cases of XDR tuberculosis in 2007^2 Of the cases of MDR tuberculosis, about $0\cdot 3$ million were new cases (primary drug resistance) and $0\cdot 2$ million were patients previously treated for tuberculosis (acquired drug resistance).

The countries ranked first to fifth in terms of total numbers of drug-resistant cases were India, China, the Russian Federation, South Africa, and Bangladesh. MDR tuberculosis accounts for 5.7% of new cases and 25.6% of previously treated cases in China, and together China and India account for about 50% of the total worldwide burden of MDR tuberculosis.³² In parts of the Russian Federation however. MDR tuberculosis accounts for as

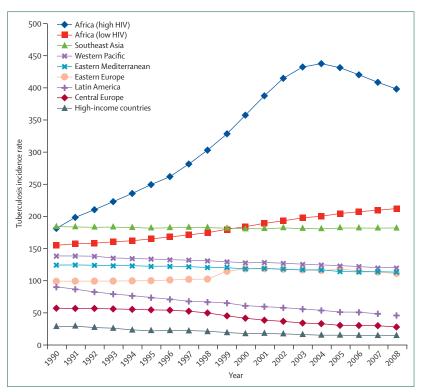


Figure 2: Trends in estimated tuberculosis incidence rates in nine subregions, 1990–2008

Data from Global tuberculosis control—a short update to the 2009 report.²

much as a quarter of new cases of tuberculosis³² and is a crucial problem in prisons in eastern Europe³³ and sub-Saharan Africa.³⁴ By March, 2010, 58 countries had reported at least one case of XDR tuberculosis to WHO.³² The intersection of the drug-resistant tuberculosis and HIV epidemics further threatens to undermine tuberculosis control, creating a so-called perfect storm in regions where the prevalence of these is high, such as in KwaZulu Natal province, South Africa.³⁵

Microbiology of Mycobacterium tuberculosis

M tuberculosis was first identified by the German scientist Robert Koch (figure 3), who announced the discovery on March 24, 1882. The M tuberculosis complex of organisms, which can cause human disease, consists of M tuberculosis, Mycobacterium africanum, Mycobacterium bovis, Mycobacterium microti, and Mycobacterium canetti. M bovis was responsible for about 6% of all human tuberculosis deaths in Europe before the introduction of milk pasteurisation; subsequent attenuation of a laboratory strain of M bovis led to the development of the BCG vaccine in 1921.

M tuberculosis is an obligate intracellular pathogen that can infect several animal species, although human beings are the principal hosts.³⁶ It is an aerobic, acid-fast, nonmotile, non-encapsulated, non-spore forming bacillus. It grows most successfully in tissues with high oxygen content, such as the lungs. Compared with the cell walls of other bacteria, the lipid-rich cell wall is relatively

	Effect
HIV ^{5,8}	Greatly increased susceptibility to infection, primary progressive disease, reactivation, and recurrence; disease incidence rate ratio of between 20 and 37 for people infected with HIV depending on country HIV prevalence
Diabetes ^{9,10}	About three-times increased risk of tuberculosis (especially in insulin-dependent disease); higher mortality
Undernutrition and vitamin deficiencies ^{11,12}	Undernutrition, low body-mass index, and vitamin D deficiency are each associated with increased risk of tuberculosis disease
Overcrowded living conditions ¹³	Increased exposure to infectious cases
Smoking ¹⁴⁻¹⁶	About two-times increased risk of infection, progression to tuberculosis disease and death
Indoor air pollution15	About two-times increased risk of disease (weak evidence)
Silicosis ¹⁷	About three-times greater risk in South African gold miners with silicosis
Alcohol ¹⁸	About three-times increased risk of disease associated with consumption >40 g per day
Sex ¹⁹	The ratio of incident tuberculosis disease in men:women is about 2:1 in adults but not children
Age ²⁰	Major effect on risks of acquisition, disease progression, form of disease, and mortality risk
End-stage renal failure ²¹	More than ten-times increased risk
Malignancy ^{22,23}	Both solid organ and haematological malignancies associated with increased risk
Genetic susceptibility ²⁴	There is a growing list of genes associated with risk of tuberculosis, including genes for natural resistance-associated macrophage protein 1, interferon γ, nitric oxide synthase 2A, mannan binding lectin, vitamin D receptor, and some Toll-like receptors
TNF antagonist therapy ^{25,26}	Risk of tuberculosis disease increased about one and a half times in rheumatology patients in North America; risk greater with TNF antibodies than with soluble TNF receptor
Corticosteroid therapy ²⁵	Risk of tuberculosis disease increased about two times in rheumatology patients in North America
TNF=tumour necrosis factor.	

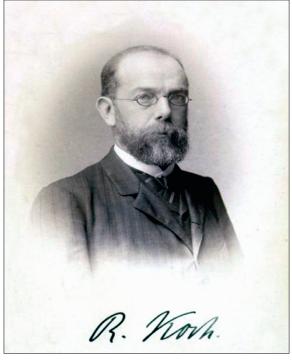


Figure 3: Robert Koch (Dec 11, 1843-May 27, 1910)
Photograph provided by Stefan Kauffman, Max Planck Institute, Berlin.

impermeable to basic dyes unless combined with phenol. Thus, *M tuberculosis* is neither gram positive nor gram negative but is instead described as acid-fast, since once stained it resists decolourisation with acidified organic solvents (figure 4). Since other bacteria, such as non-tuberculous mycobacteria and *Nocardia* spp, also contain

mycolic acids, they are also acid-fast and cannot be distinguished from *M tuberculosis* on microscopic sputum smear examination. *M tuberculosis* divides every 15–20 h, which is extremely slow compared with other bacteria (*Escherichia coli* divides every 20 minutes). This slow replication rate and ability to persist in a latent state result in the need for long durations of both drug therapy of tuberculosis and for preventive therapy in people with *M tuberculosis* infection.

Although many questions remain unanswered about the origins of M tuberculosis, advances in mycobacterial genomics are now providing evidence that the amount of sequence variation in the M tuberculosis genome might have been underestimated and that some genetic diversity does have important phenotypic consequences. 37,38 Although there are many M tuberculosis strains, studies of the phylogeny and biogeography of M tuberculosis have revealed six main strain lineages that are associated with particular geographical regions.³⁸ Much research is focused on elucidating possible differences between strains with regard to transmission and pathogenesis. It is speculated that the Beijing family of strains originated in Asia and the strain W and strain W-like families are responsible for many cases of drug resistance. This family of strains is distributed worldwide and is able to spread in large clonal clusters. However, it has not been defined if this family has a genetic advantage that enables it to spread, cause disease, and develop drug resistance.39 The sequencing of the M tuberculosis genome was a major step forward towards increasing our overall understanding of the bacterium40 and this has subsequently led to identification of specific antigens for the development of new diagnostics tests, vaccines, and biomarkers for tuberculosis.

Host-pathogen interactions

The yearly probability of developing active clinical tuberculosis after inhalation of an M tuberculosis aerosol from an infectious patient with active tuberculosis is very small, with an estimated lifetime risk of about 10%.27 The risk of transmission is highest within the first few years after infection, but decreases substantially thereafter. Most immunocompetent individuals (over 90% of those infected) either eliminate M tuberculosis or contain it in a latent state. So-called latent tuberculosis is a clinical disorder in individuals infected with M tuberculosis in whom the host immune system retains sufficient control over replication of the bacterium such that the individual remains free of tissue damage and symptoms. The presence of viable *M tuberculosis* bacilli in such individuals has been shown by culturing the organism from tissues obtained from healthy individuals who died from traffic accidents but who had no macroscopic or histological evidence of tuberculosis.41

An estimated 2 billion people worldwide have latent M tuberculosis infection. 42 Despite the great importance of this enormous reservoir of potential disease, the interactions of M tuberculosis with the human host that mediate clinical latency are largely unknown. With advances in technology, our understanding of pathogenesis and protective immune responses to infection with M tuberculosis is constantly growing.43 M tuberculosis has evolved elaborate survival mechanisms in human beings that allow it to remain in a clinically latent state, although the mechanisms of persistence remain incompletely defined. The high rates of clinical tuberculosis in people infected with HIV and in those with various inherited defects of the interferon-γ signalling pathway indirectly suggest a key role for the adaptive immune responses after antigen recognition by specific T cells.44,45

However, the first interaction between M tuberculosis and the host is with the innate immune system and seems to be mediated by pattern recognition receptors. Recognition by macrophages and dendritic cells of the biochemical products of M tuberculosis such as mannosylated lipoarabinomannan,46 trehalose dimycolate,47 and N-glycolyl muramyl dipeptide48 trigger innate responses and might be important in establishing ensuing host-pathogen interactions. However, the final effector pathway remains unclear. In addition to macrophage activation, a potential role for neutrophils in the innate immune response is also emerging. Studies suggest that neutrophils are not simply scavenging phagocytic cells but are infected with *M tuberculosis* in the sputum and within cavities of patients with active tuberculosis.49 Several antimicrobial peptides, such as cathelicidin LL37, produced by neutrophils, have activity against M tuberculosis. 50 Uptake of M tuberculosisinduced apoptotic neutrophils by macrophages also triggers macrophage activation, which provides a subsequent link between the innate and acquired immune response.⁵¹ Furthermore, there is growing interest in the

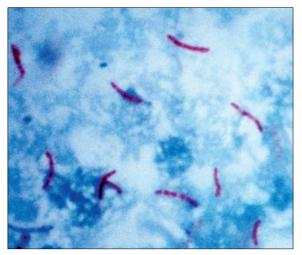


Figure 4: High-power micrograph of acid-fast bacilli in the sputum of a patient with tuberculosis, shown by Ziehl-Neelsen staining (×1000)

link between susceptibility to tuberculosis and vitamin D deficiency; this has pleiotropic effects on the immune system,⁵² including macrophage activation and induction of the antimycobacterial peptide LL37.⁵³ Vitamin D supplementation might enhance antimycobacterial immune function in vitamin D deficient populations. A randomised clinical trial⁵⁴ showed that vitamin D supplementation was associated with more rapid sputumculture conversion in a subset of individuals with specific vitamin D receptor polymorphisms and further clinical trials of vitamin D supplementation in the treatment and prevention of tuberculosis are warranted.

Conventional dogma, supported by an extensive body of research, has focused on cell-mediated immunity, with T lymphocytes and macrophages regarded as having the dominant role in protective immunity to M tuberculosis. 43,55 In immunocompetent individuals, this culminates in the formation of granulomas, which are highly effective in containing, but not eliminating, the infection. 55-57 These granulomas have a dominant role in the pathogenesis of M tuberculosis and other intracellular pathogens (figure 5) and are defined as focal, compact collections of inflammatory cells in which mononuclear cells dominate and are usually formed as a result of an undegradable product, microorganisms, or hypersensitivity reaction.⁵⁶ Recent molecular studies suggest that mycobacteria might promote cellular recruitment to the granuloma, which further suggests that granuloma formation is part of a pathogen-directed virulence programme. Therefore, although granuloma formation seems to function as a host defence mechanism, there are also apparent advantages of this response conferred to M tuberculosis. 55,58-60

Within granulomas, *M tuberculosis* might shield itself from immune-based killing mechanisms and escape therapeutic concentrations of antituberculosis drugs, potentially promoting the emergence of drug-resistant

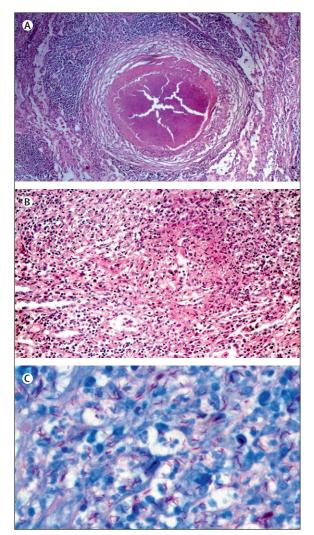


Figure 5: Micrographs of tissue specimens from patients with tuberculosis Low-power micrograph (×100) of a haematoxylin and eosin stained tissue section from an immunocompetent patient with tuberculosis that shows a well formed tuberculous granuloma with a central area of caseous necrosis surrounded by epithelioid macrophages, giant cells, and Tlymphocytes, and surrounding outer fibrosis (A). Medium-power (×250) micrograph of tissue from a patient infected with HIV with advanced immunodeficiency that shows mononuclear infiltrate but absence of granuloma formation (B). High-power (×400) tissue section from the same specimen from the patient with advanced immunodeficiency with Ziehl-Neelsen staining showing numerous acid-fast bacilli with little evidence of a cellular immune host response (C). Micrographs provided by Colleen Wright, Stellenbosch University, South Africa.

strains. At a cellular level, the granuloma macrophage might also have two mutually contradictory roles: activated macrophages are capable of killing or controlling the growth of *M tuberculosis*, and yet they also provide the primary growth niche for this intracellular organism. ^{55,58-60} The fate of infected macrophages has an essential role in protection against *M tuberculosis* by regulating innate and adaptive immunity. Virulent strains of *M tuberculosis* inhibit apoptosis and trigger macrophage necrosis, thereby evading innate immunity and delaying the start of adaptive immune responses. ⁶¹

Granulomas are absent or poorly formed in people with poor immune responses, particularly those infected with HIV (figure 5). 45,62 Through several mechanisms, HIV-1 coinfection leads to functional and numeric depletion of M tuberculosis-specific CD4 T lymphocytes and type-1 cytokine production. The resulting dysfunction of the CD4 T lymphocytemacrophage immune axis impairs the host's ability to orchestrate cell-mediated immune responses and form immunologically competent granulomas.63 As a result, histological examination of tissue specimens might reveal uncontrolled *M tuberculosis* replication with little evidence of a host cellular response (figure 5). Sites of tuberculosis disease and granulomas themselves provide the ideal microenvironment for the propagation of HIV-1, thereby maximising the adverse consequences of HIV-1 at the crucial interface between M tuberculosis and the host.45

Our understanding of the host–pathogen dynamics of infection with M tuberculosis has fundamentally changed in recent years. The traditional theory that distinguishes latent infection from active disease as distinct binary states is overly simplistic. Granulomas are not fixed inert structures, as previously described; they are very active with constantly changing dynamic structures of metabolically active tissues.64 It is thought likely that a continuous spectrum of states exists both in the same individual and between different individuals, with varying degrees of immune control and mycobacterial bacillary load,64-66 and that HIV substantially shifts this spectrum in favour of bacillary replication.67 In view of this new theory, new biomarkers are needed that can more precisely define these disease states and assess the probability of progression of infection with *M tuberculosis* to active tuberculosis disease.

Biomarkers

High on the tuberculosis research agenda is the discovery of host and pathogen biomarkers of active tuberculosis for diagnosis, monitoring treatment, and assessing outcomes (including cure and relapse). A biomarker is defined as a characteristic that is objectively measured and assessed as an indicator of normal biological processes, pathogenic processes, or pharmacological responses to a therapeutic intervention. Biomarkers thus provide information about current health status, future health status, and advance knowledge of pathogenesis. Therefore, they might potentially be used to predict reactivation risk, tuberculosis cure, eradication of latent tuberculosis, and vaccine efficiency, and also provide endpoints for clinical trials. By contrast, a diagnostic test classifies patients at a single timepoint as having active tuberculosis, latent infection with M tuberculosis, or neither. A biomarker of set of biomarkers might serve as a diagnostic test.

Biomarkers can be either host or pathogen specific sand tuberculosis-specific ones are needed to serve as

surrogate endpoints, assisting candidate selection during drug discovery, accelerating dose selection in early clinical studies, shortening the time to licensing of new drugs, and development and assessment of new vaccines for tuberculosis. Although progress has been slow, newer technologies are being used to study large cohorts of patients and have yielded new data.⁷⁰⁻⁷⁴

In an attempt to identify new biomarkers, multiplexed assays are now being used to compare gene expression between patients with tuberculosis, healthy people with latent infection, and healthy people with no exposure to *M tuberculosis* (controls).^{70,75,76} Several biomarkers, when combined, might be substantially better than any single marker and a small number of studies suggest that specificity and higher predictive values can be achieved by measuring several variables with proteomics, transcriptomics, and metabolomics.⁷⁵ Recent transcriptomic studies from South Africa,^{76,77} a high tuberculosis endemic area, have identified signatures involving expression profiles of genes in blood cells that could distinguish active tuberculosis, latent infection, cure, and tuberculosis recurrence.

Another study of South African patients with active and latent infection with tuberculosis" identified a whole-blood 393-transcript signature for active tuberculosis in intermediate-burden and high-burden settings, correlating with the radiological extent of disease and reverting to that of healthy controls after treatment. The investigators also identified a specific 86-transcript signature that discriminates active tuberculosis from other inflammatory and infectious diseases. Modular and pathway analysis revealed that the tuberculosis signature was dominated by a neutrophil-driven interferon-inducible gene profile. The study provides a broad range of transcriptional biomarkers with potential as diagnostic and prognostic techniques.

Separate studies might be needed to assess biomarkers in people with HIV infection. With accumulation of data from multiparameter assay studies, it is becoming clear that biomarkers, and the associations they generate, are probably specific and restricted to the population under study and might not necessarily be broadly applicable.

Diagnostics

The estimated worldwide detection rate for new sputum smear-positive cases of tuberculosis of 62% in 2008 fell substantially short of the 2005 target detection rate of 70%,² and the lack of accurate and rapid diagnostics remains a major obstacle to progress in this regard. Over 90% of the worldwide burden of tuberculosis is in low-income and middle-income countries where the diagnosis of tuberculosis still relies heavily on sputum smear microscopy and chest radiology. These techniques are often unsatisfactory and unavailable at patients' first point of contact with the health system. There is a great need for rapid point-of-care tests that can be readily used at all levels of the health system and in the community.

Childhood tuberculosis and sputum smear-negative pulmonary and extrapulmonary tuberculosis in adults remain the greatest diagnostic challenge.

Progress has been made over the past decade to improve existing tuberculosis diagnostics and develop new technologies, some of which have been endorsed by WHO. 69.78 In a meta-analysis, 79 sputum processing with bleach or sodium hydroxide and centrifugation was associated with an average 13% increase in the sensitivity of smear microscopy. Fluorescence microscopy also increases sensitivity by 10%, while retaining similar specificity compared with conventional Ziehl-Neelsen staining. 80 This technology permits much more efficient reading of slides, which is crucial to improving laboratory performance. Although traditional fluorescence microscopes are expensive, much cheaper fluorescence microscopes with light-emitting diodes (LEDs) are equally sensitive and were endorsed by WHO in 2009.81

Automated liquid culture systems are now the gold standard for the diagnosis of tuberculosis; they are substantially faster and have a 10% greater yield than solid media. In 2007, these systems were recommended by WHO to be used in combination with antigen-based species confirmation for diagnosis and drug susceptibility testing (DST) in low-income and middle-income countries.⁸² However, such systems are expensive and prone to contamination. Alternative inexpensive noncommercial culture and DST methods were endorsed by WHO in 2009 for use as an interim solution in resource-constrained settings.⁸¹ These alternatives include microscopically observed drug susceptibility (MODS) and the nitrate reductase assav.⁷⁸

To enhance capacity for rapid diagnosis of MDR tuberculosis, WHO in 2008 approved the use of line probe assays (LPAs) for the rapid molecular detection of drug resistance in smear-positive specimens or culture isolates.82 Two commercial LPAs have shown high accuracy when applied to culture isolates and one of these, the GenoType MTBDRplus assay (Hain Lifescience GmbH, Nehren, Germany), also shows very good performance characteristics when applied directly to smear-positive sputum specimens.83,84 In 2009, the GenoType MTBDRsl assay became available which is also able to detect resistance to fluoroquinolones, aminoglycosides, and ethambutol in culture isolates or smear-positive sputum specimens.85 When used in combination with the GenoType MTBDRplus assay, this potentially provides a means of rapid detection of XDR tuberculosis. These and similar molecular assays reduce the time to diagnosis of MDR and XDR tuberculosis from weeks or months to a matter of days. However, it has yet to be shown whether the use of such assays improves patient outcomes.

Although commercially available, serological tests for tuberculosis are of little diagnostic value,⁷⁸ mycobacterial antigen detection is theoretically more attractive, overcoming many of the limitations inherent to

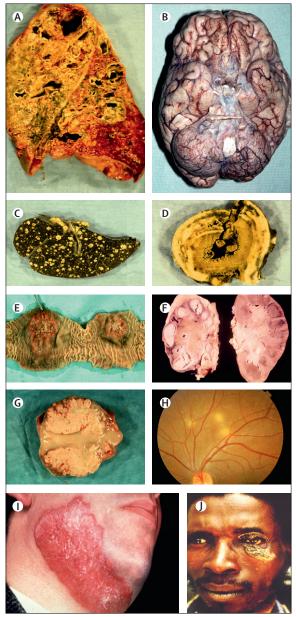


Figure 6: Tuberculosis in various organs

Lung specimen showing several cavities, caseous necrosis, and extensive lung pathology due to tuberculosis (A). Ventral view of a brain specimen showing basal tuberculous meningitis and a fibrinous meningeal exudate (B). Multiple caseating lesions of miliary tuberculosis in the spleen (C). Cross-section of the heart showing chronic tuberculous pericarditis with a dense 2 cm thick band of fibrosis (D). Caseating tuberculous ulcers of the ileum (E). Kidney specimens showing caseating lesions due to tuberculosis (F). Section of lymph node showing florid caseating tuberculous lesions (G). Eye fundus with miliary choroid tubercles in an HIV-infected patient with miliary tuberculosis (H); photograph provided by Miles Stanford, King's College London, London, UK). Chronic granulomatous lesions of the face due to Mycobacterium tuberculosis (lupus vulgaris; I and J). Photographs (except that of the eye) provided by Sebastian Lucas, St Thomas's Hospital, London, UK.

immune-based assays. A simple commercially available assay is able to detect lipoarabinomannan excreted in the urine of patients with tuberculosis. Although the

sensitivity has been disappointing in patients not infected with HIV, moderate sensitivity and high specificity has been noted in patients infected with HIV with advanced immunodeficiency^{86,87} and point-of-care versions of this assay are being assessed.⁸⁸

Nucleic acid amplification tests (NAATs) are the most promising development in tuberculosis diagnostics. ⁷⁸ In the USA and Europe, these tests have been shown to have high specificity, but slight and variable sensitivity, especially for sputum smear-negative disease. ^{89,90} Simplified versions of these assays with higher sensitivity are being developed. A simplified manual NAAT that uses loop-mediated isothermal amplification with a simple visual colorimetric readout, ⁹¹ is being assessed in peripheral laboratory facilities in resource-constrained settings.

A sensitive and specific fully automated and commercially available NAAT assay has now been developed for use outside reference laboratory centres.92 This Xpert MTB/ RIF assay (Cepheid, Sunnyvale, CA, USA) uses a series of molecular probes and real-time PCR technology to detect *M tuberculosis* and the *rpoB* rifampicin resistance mutation. The cartridge-based system dispenses with the need for the sputum to be processed in advance, needs minimum laboratory expertise, and results are available in less than 2 h, which permits a specific tuberculosis diagnosis and rapid detection of rifampicin resistance. A large multicountry assessment93 found excellent performance characteristics. In culture-positive patients, a single direct Xpert MTB/RIF assay identified 551 (98%) of 561 patients with sputum smear-positive tuberculosis and sensitivities were 72.5% when processing one sputum specimen, 85.1% when processing two, and 90.2% when processing three for sputum smear-negative disease.⁹³ Specificity was 99.2%. If these results are replicated under field conditions at points of care, and the price of introducing the Xpert MTB/RIF assay to points of care in resource-poor countries is brought down, it will represent a major breakthrough in rapid tuberculosis diagnostics and for rifampicin resistance screening.

For the past century, the tuberculin skin test has been the only screen available for the diagnosis of latent infection with tuberculosis. Its major failing is its inability to reliably distinguish individuals infected with *M tuberculosis* from individuals sensitised to other mycobacteria, including BCG. A decade ago the interferon-γ release assays (IGRAs) were developed whereby interferon-γ titres were measured after in-vitro stimulation of peripheral blood mononuclear cells with antigens such as ESAT-6 and CFP-10 (immunodominant antigens expressed by members of the *M tuberculosis* complex). These have now become the gold standard for identifying individuals whose immune system has previously encountered *M tuberculosis*.

Two commercial methods were introduced—the T-SPOT. TB test (Oxford Immunotech, Abingdon, UK) and the QuantiFERON-TB Gold in tube (Cellestis Ltd, Carnegie, Australia)—and have been extensively tested in many clinical situations and in individuals infected with HIV.⁹⁴

The assessment of these tests' results for detection of latent tuberculosis have been difficult because of the absence of a gold standard for tuberculosis latency. A meta-analysis of these studies showed that IGRAs are at least as sensitive and more specific than the tuberculin skin test. Longitudinal studies have shown that the predictive value of IGRAs for reactivation of tuberculosis in immunosuppressed individuals is better than that provided by the tuberculin skin test in individuals vaccinated with BCG.

High levels of interferon- γ release are detected by these assays in about 70–90% of individuals with active disease and these levels decrease after treatment is completed, although such reductions are not consistently recorded. 95–97 The high sensitivities but low specificities of the IGRAs noted in several phase 2 tuberculosis diagnostic studies suggest that IGRAs can be used as rule out but not rule in tests for diagnosis of active tuberculosis. 94,98,99 Further studies of unselected patients need to be done in controlled trials. More recent phase 2 studies of other markers of T-cell responses have shown that interferon γ -inducible protein 10, interleukin 10, and monocyte chemotactic protein 1, show potential for improved detection of active tuberculosis in patients. 100

Clinical presentation

Although tuberculosis predominantly affects the lung, it can cause disease in any organ (figure 6) and must be included within the differential diagnosis of a vast range of clinical presentations. Symptoms and signs include those associated with the specific disease site as well as nonspecific constitutional symptoms such as fever, weight loss, and night sweats. However, in the early stages of disease, symptoms might be absent as shown by community-based active case finding studies in Asia¹⁰¹ in which about one in four culture-confirmed cases of pulmonary tuberculosis were reported to be asymptomatic.

A high index of suspicion for tuberculosis must especially be maintained when caring for patients living with HIV infection, since risk of tuberculosis is high and diagnosis is difficult. Although patients with high CD4 cell counts present with typical features of tuberculosis, progressive immunodeficiency substantially affects the spectrum of disease with increasing risk of extrapulmonary and disseminated disease. 102,103 Conventional screening for cough lasting 2 or 3 weeks typically has a sensitivity of less than 50% for active tuberculosis in this patient group and about 20% of patients detected on active screening report no symptoms at all.104 Active microbiological screening for tuberculosis irrespective of symptoms is therefore an important approach and might detect a substantial burden of disease. Up to 25% of patients screened before starting antiretroviral drugs in South Africa have sputum-culture positive tuberculosis.86,105 Clinical samples from extrapulmonary sites such as fine needle aspiration biopsy of lymph nodes might also provide incremental diagnostic yield.106

Panel 1: Recommendations included in the WHO guidelines for the treatment of drug sensitive tuberculosis

First-line 6 month treatment regimen

New patients with pulmonary tuberculosis should receive a regimen containing 6 months of rifampicin: 2HRZE/4HR (high grade of evidence)

Alternative first-line continuation phase

In populations with known or suspected high levels of isoniazid resistance, new tuberculosis patients can receive HRE as treatment in the continuation phase as an acceptable alternative to HR (weak evidence)

Optimum dose frequency

- Wherever feasible, the optimum dose frequency for new patients with pulmonary tuberculosis is daily throughout the course (high grade of evidence)
- Alternatively, new patients with pulmonary tuberculosis can receive a daily intensive phase and then three-times weekly continuation phase provided that each dose is directly observed: 2HRZE/4(HR)₃ (high/moderate grade of evidence)
- Alternatively, new patients with pulmonary tuberculosis can receive three-times
 weekly dosing throughout treatment, provided that every dose is directly observed
 and the patient is not living with HIV or living in a high HIV prevalence setting:
 2(HRZE)₃/4(HR)₃ (high/moderate grade of evidence)

Retreatment regimens and detection and treatment of drug resistance

- Ideally, DST is done for all patients at the start of treatment, so that the most appropriate treatment for each individual can be established
- Specimens for culture and DST should be obtained from all previously treated patients with tuberculosis at or before the start of treatment; DST should be done at least for isoniazid and rifampicin
- In settings where rapid molecular DST is available, the results should guide the choice of regimen
- In settings where rapid molecular-based DST results are not routinely available to guide the management of individual patients, empirical treatment should be started as follows:
 - Tuberculosis patients whose treatment has failed or other patient groups with high likelihood of MDR tuberculosis should be started on an empirical MDR regimen
 - Tuberculosis patients returning after defaulting or relapsing from their first treatment course can receive the retreatment regimen containing first-line drugs 2HRZES/1HRZE/5HRE if country-specific data show low or medium levels of MDR in these patients or if such data are not available
- In settings where DST results are not yet routinely available to guide the management
 of individual patients, the empirical regimens will continue throughout the course of
 treatment

Numbers in the treatment algorithms are the months of treatment and subscript number is the dose frequency per week. H=isoniazid. R=rifampicin. Z=pyrazinamide. E=ethambutol. DST=druq susceptibility testing. MDR=multidruq resistant.

The initial clinical presentation of patients with drugresistant *M tuberculosis* strains does not typically differ from that of patients with resistant strains. Drug-resistant tuberculosis should be suspected in patients who do not respond to the intensive phase of standard short course therapy, have previously had tuberculosis, have a history of poor adherence aggravated by social deprivation or substance abuse, are known contacts of patients with drug-resistant tuberculosis, or who live in regions where the prevalence of drug-resistance is known to be high. However, more than half of the patients with drugresistant tuberculosis have none of these risk factors and

Panel 2: Principles underlying the treatment of multidrugresistant tuberculosis adapted from WHO guidelines

Number of drugs

Treatment regimens should consist of at least four drugs with either certain, or almost certain, effectiveness. Often more than four drugs are started if the susceptibility pattern is unknown or questionable.

Reliability of DST

In general, susceptibility testing for isoniazid, rifampicin, the fluoroquinolones, and the injectable drugs is fairly reliable. For other drugs this is less reliable and basing individualised treatments on DST for these drugs should be avoided.

Treatment administration

Each dose of an MDR regimen should be given by directly observed therapy throughout the treatment.

Monitoring treatment response

To assess treatment response, smears and cultures should be done monthly until smear and culture conversion (two negative smears and cultures taken 30 days apart). Thereafter smears should be monitored at least monthly and cultures quarterly.

Duration of intensive phase

The intensive phase of treatment for MDR tuberculosis is defined by the duration of treatment with the injectable drug. This should be given for a minimum of 6 months and for at least 4 months after the patient first becomes and remains smear or culture negative.

Total duration of therapy

Treatment for MDR tuberculosis should be given for a minimum of 18 months after culture conversion, but extension to 24 months might be indicated in patients with chronic disease with extensive pulmonary damage.

DST=drug susceptibility testing. MDR=multidrug resistant.

thus ideally every patient with proven tuberculosis should be tested for drug susceptibility.

Treatment

The WHO revised international guidelines for the treatment of tuberculosis in 2010,107 specifically responding to the growing evidence base 108-111 and escalating problem of drug-resistant disease worldwide.32 Earlier guidelines emphasised the use of two main standardised treatment regimens, one for new (previously untreated) cases and one for patients with sputum smear-positive disease who had previously received treatment (retreatment regimen). The drug combinations used in these two regimens differed only by the addition of a single drug—a far from optimum situation with regard to prevention of emergence of drug resistance. Lack of laboratory infrastructure for culture and DST in many settings with a high burden of tuberculosis resulted in widespread empirical use of the

retreatment regimen. Blind therapy combined with intermittent treatment adherence might have inadvertently fuelled the emergence of multidrug-resistant strains.¹¹²

Panel 1 summarises the key recommendations from the 2010 WHO tuberculosis treatment guidelines. 107 Rifampicin should now always be given throughout the total 6 months duration of the first-line regimen. New emphasis is placed on the crucial role of DST for guiding the individual management of patients who have previously received treatment for tuberculosis. At present infrastructure for routine DST is scarce, precluding widespread adoption of this recommendation. For settings where this is not routinely possible, national tuberculosis programmes are strongly encouraged to assess country-specific drug resistance data for patients with treatment failure, relapse, and default to inform local policy on empirical use of either retreatment or MDR tuberculosis treatment regimens. Recommendations for the management of drug-resistant tuberculosis are contained within the WHO 2010 tuberculosis treatment guidelines.¹⁰⁷ Panel 2 summarises key principles of treatment, which need carefully informed choices of optimum drug combinations.113

Treatment for drug-resistant disease is very costly and prolonged, and is associated with high rates of drugrelated toxic effects. However, successful outcomes for MDR tuberculosis are achievable in about two-thirds of patients.¹¹⁴ Outcomes of treatment for XDR tuberculosis are very heterogeneous;114-117 although HIV-negative patients in Peru had similar outcomes to those of patients with MDR tuberculosis, 117 XDR tuberculosis was almost universally fatal in a localised outbreak in patients infected with HIV with advanced immunodeficiency in South Africa.¹¹⁶ In most parts of the world, access to such therapy is very poor, with less than 2% of patients with MDR tuberculosis worldwide treated according to WHO standards in 2008.2 Thus, in May, 2009, the World Health Assembly passed a resolution urging member states to provide universal access to diagnosis and treatment of MDR and XDR tuberculosis.32

Although patients with HIV-associated tuberculosis receive the same antituberculosis treatment regimens as patients not infected with HIV, an additional package of care added to their management regimen can reduce their high mortality risk. HIV testing serves as the crucial gateway for accessing such care and yet only 26% of tuberculosis patients worldwide were tested in 2009. Provider-initiated counselling and HIV testing can greatly increase access to appropriate care 119,120 and rates of testing reached 53% in sub-Saharan Africa in 2009. Mortality risk is reduced by 22–48% with the use of co-trimoxazole prophylaxis 121 and by 64–95% with antiretroviral therapy (ART). 122

Data are now emerging from controlled clinical trials of the optimum time to start ART in such patients. These studies have shown that irrespective of the CD4 cell count, deferral of ART to the end of treatment for tuberculosis is associated with high mortality risk 124 and

that mortality is reduced by 34% in patients in Cambodia with very low CD4 cell counts (median 25 cells per $\mu L)$ who are started on ART within the first 2 weeks of treatment rather than after 2 months. 125 The 2010 revision of the WHO ART guidelines recommends that, irrespective of CD4 cell count, all patients with HIV-associated tuberculosis should receive ART as soon as possible during the first 2–8 weeks of treatment for tuberculosis. 126

Despite pharmacokinetic interactions and potential cotoxicity,127 excellent virological responses and low rates of treatment limiting toxic effects are noted with overlapping efavirenz-based ART and tuberculosis treatment. 118,128,129 However, use of rifampicin in patients that receive protease-inhibitor-based treatment remains more problematic, needing either dose adjustment or preferably substitution of rifampicin with rifabutin where available. 130 Tuberculosis immune reconstitution disease might complicate ART when started during treatment for tuberculosis, with rapid restoration of pathogen-specific immune responses resulting in the deterioration in the clinical characteristics of tuberculosis.¹³¹ A systematic review¹³² reported that this complication developed in 16% (95% CI 10-25) of patients. In most patients immune reconstitution disease is self-limiting, but a small proportion (about 3%) of such patients die. 131,132 A randomised controlled trial has shown that morbidity associated with tuberculosis immune reconstitution disease can be effectively reduced with the use of corticosteroids. 133

Future successes in the control of tuberculosis will depend on the development of new antituberculosis drugs used in treatment regimens that are shorter, easier to deliver, safe, and low in cost. After decades of neglect, ten new drugs for the treatment of tuberculosis are in the clinical development pipeline of which six were specifically developed for tuberculosis.134 Examples include the diarylquinolone, TMC-207, which targets mycobacterial ATP synthase and in a phase 2 clinical trial greatly increased sputum smear conversion in patients with MDR tuberculosis.¹³⁵ Nitroimidazoles, such as PA-824 and OPC-67683, are equally active against drug-susceptible and drug-resistant tuberculosis136 and are also being assessed in clinical trials. Since nitroimidazoles are active against both replicating and non-replicating organisms, they could potentially shorten treatment of active disease and provide activity against latent infection with tuberculosis. 134 Assessment of the use of moxifloxacin, gatifloxacin, and TMC-207 in treatment regimens to shorten the duration of chemotherapy are ongoing.

Vaccines for tuberculosis

There is a dire need for a universally effective vaccine for the control of tuberculosis.^{137,138} The only licensed vaccine, BCG, was first given to a human infant in 1921. The vaccine has been given to 4 billion people so far and to more than 90% of the children in the world today, making

it the most widely used vaccine in the world. However, it has done little to contain the current tuberculosis pandemic. Despite evidence of confirmed efficacy against childhood tuberculous meningitis and miliary tuberculosis, protection induced by BCG can wane within a decade and thus the efficacy against adult pulmonary tuberculosis is variable.¹³⁹ Current pre-exposure vaccination strategies can only aim at reducing the initial M tuberculosis bacterial burden and at preventing reactivation of latent infection. Post-exposure vaccines are also needed to target these dormant bacteria and prevent their reactivation. Post-exposure vaccines would ideally also prevent reinfection of individuals living in regions of high tuberculosis prevalence. Whole heatkilled Mycobacterium vaccae, an environmental saprophyte, has been tested as an immunotherapeutic agent for the treatment of tuberculosis in three trials in Africa with discrepant outcomes. 140 A randomised, placebocontrolled, double-blind study of M vaccae given in several doses to BCG-vaccinated participants infected with HIV, showed a reduction in cases of tuberculosis.141 A dozen new vaccine candidates are in clinical trials, with the aim of either replacing the existing BCG vaccine or enhancing immunity induced by BCG. 137 Several phase 1 and 2 clinical trials are underway in South Africa.112

Tuberculosis control

After the declaration in 1993 that tuberculosis was a global emergency, WHO launched the directly observed treatment, short-course (DOTS) strategy, which was successfully expanded as the principal tuberculosis control strategy, focusing primarily on detection and effective treatment of infectious cases. Between 1995 and 2008, 43 million people were treated under DOTS, 36 million were cured, case-fatality rates decreased from 8% to 4%, and an estimated 6 million deaths were potentially averted.² After a decade of implementation, the new STOP TB Strategy and the Global Plan to Stop TB (2006-15) were launched in 2006 to address important challenges that included the HIV-associated tuberculosis epidemic, the emergence of the MDR tuberculosis epidemic, weak health systems, and insufficient engagement with private healthcare providers and with communities. 142,143

Progress is being monitored against an established series of goals and targets (panel 3). He The earlier worldwide target of an 85% treatment success rate for sputum smear-positive cases was first achieved in 2007, and the Millennium Development Goal 6 target to reverse the rising incidence in tuberculosis incidence rates has been fulfilled since 2004. However, although the worldwide case detection rate increased substantially between 1995 and 2008, it has stabilised at around 60%, falling substantially short of the 70% target. Moreover, the targets of halving the 1990 tuberculosis prevalence and mortality rates by 2015 are unlikely to be met worldwide, with the epidemics of HIV-associated tuberculosis in Africa and MDR tuberculosis in eastern Europe being key stumbling blocks.²

Panel 3: Performance targets for tuberculosis control

World Health Assembly, 1991

Targets originally set for 2000, later postponed to 2005 and now deemed obsolete in view of the call for universal detection and cure:

- Achieve a worldwide case detection rate of 70%
- Achieve a worldwide cure rate of 85%

MDG 6

Target 6.c: halt and begin to reverse the incidence of tuberculosis by 2015

Targets linked to the MDGs and endorsed by the STOP TB Partnership

By 2015: reduce the prevalence of tuberculosis and deaths due to tuberculosis by 50% compared with the baseline of 1990

By 2050: to eliminate tuberculosis as a public health problem as defined by achieving a worldwide incidence of tuberculosis of less than 1 case per million population per year

Adapted from the WHO Global Plan to STOP TB 2011–15. 144 MDG=Millennium Development Goal.

Despite substantial progress in worldwide tuberculosis control, it is unclear why tuberculosis incidence is only falling at a rate of less than 1% per year. The potential benefits of DOTS might be offset by changing population risk factors145 or late tuberculosis diagnosis might mitigate the effect on transmission rates. Rates of decline might also be more strongly related to social and economic factors and general population health than the performance of national tuberculosis control programmes.3,146 To eliminate tuberculosis as a public health problem by 2050, incidence must fall by an average of 16% yearly over the next 40 years.145 However, even if the Global Plan to Stop TB were successfully implemented, incidence would only decrease at around 6% yearly, meaning that worldwide incidence rates in 2050 would remain 100-times higher than the elimination target.27 In addition to biomedical interventions related to tuberculosis, improvements in tuberculosis control will also need to progress in the development and strengthening of health systems and progress in the broader development agenda.7,147

One of the greatest challenges to tuberculosis control is the HIV-associated tuberculosis epidemic, for which the DOTS strategy is insufficient. ^{5,118} Preventive interventions are also needed and the four principal tuberculosis prevention methods available include intensified case finding, isoniazid preventive therapy (IPT), tuberculosis infection control, and ART. ¹¹⁸ So far ART is the only one of these that has been implemented at scale, provided to an estimated 5·3 million people in low-income and middle-income countries by the end of 2009. ¹⁴⁸ By contrast, only 4·1% of HIV-infected patients benefited from intensified tuberculosis case finding and 0·2% of eligible individuals received IPT in 2008. ² To galvanise

greater momentum in the implementation of these interventions, WHO launched in 2008 the 3Is policy, which consists of IPT, intensified case finding, and infection control to be scaled up in parallel with ART. 149,150

A key reason underlying the failure of DOTS to control the HIV-associated tuberculosis epidemic in Africa is that it fails to address the fundamental epidemiological interactions between tuberculosis and HIV in which progressive immunodeficiency fuels high rates of disease. Immune recovery during ART is associated with a 67% (95% CI 61–73) reduction in tuberculosis incidence rates in cohorts in both high and low tuberculosis burden settings and irrespective of tuberculin skin test status. 122,150 The overall effect at a population level will depend greatly on the time that patients spend at low CD4 cell counts both before and during ART. 150,151 Much more aggressive scale-up of ART in combination with the 3Is interventions would potentially have a greater effect on this epidemic. 152

A major impediment to achieving control of tuberculosis is the lack of resources to effectively implement the Global Plan to Stop TB. The estimated funding needed for the period 2006-15 was US\$60 billion. Funding for tuberculosis control in the 22 high burden countries increased from \$1.84 billion in 2006 to an estimated \$2.64 billion in 2010. 153 However, during the same period, the projected funding shortfall also increased from \$145 million to \$500 million. Although donor funds might facilitate the provision of essential tuberculosis and HIV services, local ownership and identification of sustainable local solutions are the key to the control of tuberculosis and for donor-dependent programmes to transfer responsibility to local governments and long-term sustainable funding models. Securing adequate funding to meet the huge demand in the current economic recession, presents a formidable challenge.

Conclusions

Tuberculosis remains a major cause of death and morbidity worldwide, and control efforts so far have not adequately controlled the epidemic in many parts of the world, especially in the countries of sub-Saharan Africa and parts of eastern Europe. Absence of a cheap point of care diagnostic test, the long duration of treatment, lack of an effective vaccine, emergence of drug-resistant tuberculosis, and weak health systems in resource-poor developing countries are all factors that continue to hamper progress towards achieving control of tuberculosis worldwide. Despite this, there is growing momentum in basic and applied research activity that is starting to yield new diagnostic, treatment, and prevention methods, and now provide grounds for optimism. However, this growing scientific momentum must be matched by massive political and funder commitment to provide adequate funding to ensure that the aims of the WHO Global Plan to STOP TB 2006-15 are achieved.153

Contributors

SDL and AZ contributed equally to the preparation of this Seminar.

Conflicts of interests

We declare that we have no conflicts of interest.

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References

- 1 Daniel TM. The history of tuberculosis: past, present and challenges for the future. In: Schaaf S, Zumla A, eds. Tuberculosis: a comprehensive clinical reference. Philadelphia, PA: Saunders, 2009: 1–7.
- 2 WHO. Global tuberculosis control: a short update to the 2009 report. Geneva: World Health Organization, 2009. http://www.who.int/tb/publications/global_report/2009/update/tbu_9.pdf (accessed Feb 1, 2010).
- 3 Dye C, Williams BG. The population dynamics and control of tuberculosis. *Science* 2010; 328: 856–61.
- 4 WHO. Global tuberculosis control. Geneva: World Health Organization, 2010. http://whqlibdoc.who.int/publications/2010/ 9789241564069_eng.pdf (accessed Dec 15, 2010).
- 5 Lawn SD, Churchyard G. Epidemiology of HIV-associated tuberculosis. Curr Opin HIV AIDS 2009; 4: 325–33.
- 6 Bates I, Fenton C, Gruber J, et al. Vulnerability to malaria, tuberculosis, and HIV/AIDS infection and disease: part 1 determinants operating at individual and household level. Lancet Infect Dis 2004; 4: 267–77.
- 7 Lonnroth K, Castro KG, Chakaya JM, et al. Tuberculosis control and elimination 2010–50: cure, care, and social development. *Lancet* 2010; 375: 1814–29.
- 8 WHO. Global tuberculosis control 2009: epidemiology, strategy, financing. Geneva: World Health Organization, 2009. http://www.who.int/tb/publications/global_report/2009/en/index.html (accessed Nov 29, 2010).
- 9 Jeon CY, Murray MB. Diabetes mellitus increases the risk of active tuberculosis: a systematic review of 13 observational studies. PLoS Med 2008; 5: e152.
- 10 Dooley KE, Chaisson RE. Tuberculosis and diabetes mellitus: convergence of two epidemics. *Lancet Infect Dis* 2009; 9: 737–46.
- 11 Cegielski JP, McMurray DN. The relationship between malnutrition and tuberculosis: evidence from studies in humans and experimental animals. *Int J Tuberc Lung Dis* 2004; 8: 286–98.
- 12 Chocano-Bedoya P, Ronnenberg AG. Vitamin D and tuberculosis. Nutr Rev 2009; 67: 289–93.
- 13 Vynnycky E, Fine PE. Interpreting the decline in tuberculosis: the role of secular trends in effective contact. *Int J Epidemiol* 1999; 28: 327–34.
- 14 Gajalakshmi V, Peto R, Kanaka TS, Jha P. Smoking and mortality from tuberculosis and other diseases in India: retrospective study of 43 000 adult male deaths and 35 000 controls. *Lancet* 2003; 362: 507–15.
- 15 Lin HH, Ezzati M, Murray M. Tobacco smoke, indoor air pollution and tuberculosis: a systematic review and meta-analysis. PLoS Med 2007; 4: e20.
- Bates MN, Khalakdina A, Pai M, Chang L, Lessa F, Smith KR. Risk of tuberculosis from exposure to tobacco smoke: a systematic review and meta-analysis. Arch Intern Med 2007; 167: 335–42.
- 17 Cowie RL. The epidemiology of tuberculosis in gold miners with silicosis. Am J Respir Crit Care Med 1994; 150: 1460–62.
- 18 Lonnroth K, Williams BG, Stadlin S, Jaramillo E, Dye C. Alcohol use as a risk factor for tuberculosis—a systematic review. BMC Public Health 2008; 8: 289.
- 19 Neyrolles O, Quintana-Murci L. Sexual inequality in tuberculosis. PLoS Med 2009; 6: e1000199.
- 20 Donald PR, Marais BJ, Barry CE III. Age and the epidemiology and pathogenesis of tuberculosis. *Lancet* 2010; 375: 1852–54.
- 21 Hussein MM, Mooij JM, Roujouleh H. Tuberculosis and chronic renal disease. Semin Dial 2003; 16: 38–44.

- 22 Kim HR, Hwang SS, Ro YK, et al. Solid-organ malignancy as a risk factor for tuberculosis. Respirology 2008; 13: 413–19.
- 23 Silva FA, Matos JO, de Q Mello FC, Nucci M. Risk factors for and attributable mortality from tuberculosis in patients with hematologic malignances. *Haematologica* 2005; 90: 1110–15.
- 24 Moller M, de Wit E, Hoal EG. Past, present and future directions in human genetic susceptibility to tuberculosis. FEMS Immunol Med Microbiol 2010; 58: 3–26.
- 25 Brassard P, Kezouh A, Suissa S. Antirheumatic drugs and the risk of tuberculosis. Clin Infect Dis 2006; 43: 717–22.
- 26 Wallis RS. Tumour necrosis factor antagonists: structure, function, and tuberculosis risks. Lancet Infect Dis 2008; 8: 601–11.
- 27 Watt CJ, Hosseini SM, Lonnroth K, Williams BG, Dye C. The global epidemiology of tuberculosis. In: Schaaf S, Zumla A, eds. Tuberculosis: a comprehensive clinical reference. Philadelphia, PA: Saunders, 2009: 17–27.
- 28 Stevenson CR, Forouhi NG, Roglic G, et al. Diabetes and tuberculosis: the impact of the diabetes epidemic on tuberculosis incidence. BMC Public Health 2007; 7: 234.
- 29 Shah NS, Wright A, Bai GH, et al. Worldwide emergence of extensively drug-resistant tuberculosis. *Emerg Infect Dis* 2007; 13: 380–87.
- 30 Gandhi NR, Nunn P, Dheda K, et al. Multidrug-resistant and extensively drug-resistant tuberculosis: a threat to global control of tuberculosis. *Lancet* 2010; 375: 1830–43.
- 31 Migliori GB, De Iaco G, Besozzi G, Centis R, Cirillo DM. First tuberculosis cases in Italy resistant to all tested drugs. Euro Surveill 2007; 12: E070517.
- 32 WHO. Multidrug and extensively drug-resistant TB (M/XDR-TB): 2010 global report on surveillance and response. Geneva: World Health Organization, 2010. http://whqlibdoc.who.int/ publications/2010/9789241599191_eng.pdf (accessed Aug 18, 2010).
- 33 Aerts A, Habouzit M, Mschiladze L, et al. Pulmonary tuberculosis in prisons of the ex-USSR state Georgia: results of a nation-wide prevalence survey among sentenced inmates. Int J Tuberc Lung Dis 2000: 4: 1104–10.
- 34 O'Grady J, Hoelscher M, Atun R, et al. Tuberculosis in prisons in sub-Saharan Africa - the need for improved health services, surveillance and control. *Tuberculosis (Edinb)* 2011; published online Jan 18. DOI:10.1016/j.tube.2010.12.002.
- 35 Wells CD, Cegielski JP, Nelson LJ, et al. HIV infection and multidrug-resistant tuberculosis: the perfect storm. J Infect Dis 2007; 196 (suppl 1): S86–107.
- 36 Grange JM. The genus Mycobacterium and the Mycobacterium tuberculosis complex. In: Schaaf S, Zumla A, eds. Tuberculosis: a comprehensive clinical reference. Philadelphia, PA: Saunders, 2009: 44–59.
- 37 Smith NH, Hewinson RG, Kremer K, Brosch R, Gordon SV. Myths and misconceptions: the origin and evolution of Mycobacterium tuberculosis. Nat Rev Microbiol 2009; 7: 537–44.
- 38 Gagneux S, Small PM. Global phylogeography of Mycobacterium tuberculosis and implications for tuberculosis product development. Lancet Infect Dis 2007; 7: 328–37.
- 39 Domenech P, Kolly GS, Leon-Solis L, Fallow A, Reed MB. Massive gene duplication event among clinical isolates of the Mycobacterium tuberculosis W/Beijing family. J Bacteriol 2010; 192: 4562–70.
- 40 Cole ST, Brosch R, Parkhill J, et al. Deciphering the biology of Mycobacterium tuberculosis from the complete genome sequence. Nature 1998; 393: 537–44.
- 41 Hernandez-Pando R, Jeyanathan M, Mengistu G, et al. Persistence of DNA from Mycobacterium tuberculosis in superficially normal lung tissue during latent infection. Lancet 2000; 356: 2133–38.
- 42 Dye C, Scheele S, Dolin P, Pathania V, Raviglione MC. Consensus statement—global burden of tuberculosis: estimated incidence, prevalence, and mortality by country—WHO Global Surveillance and Monitoring Project. JAMA 1999; 282: 677–86.
- 43 Eley BS, Beatty DW. The basic immunology of tuberculosis. In: Schaaf S, Zumla A, eds. Tuberculosis: a comprehensive clinical reference. Philadelphia, PA: Saunders, 2009: 75–86.
- 44 Ottenhoff TH, Verreck FA, Hoeve MA, van de Vosse E. Control of human host immunity to mycobacteria. *Tuberculosis (Edinb)* 2005; 85: 53–64.

- 45 Lawn SD, Butera ST, Shinnick TM. Tuberculosis unleashed: the impact of human immunodeficiency virus infection on the host granulomatous response to Mycobacterium tuberculosis. Microbes Infect 2002; 4: 635–46.
- 46 Gringhuis SI, den Dunnen J, Litjens M, van der Vlist M, Geijtenbeek TB. Carbohydrate-specific signaling through the DC-SIGN signalosome tailors immunity to Mycobacterium tuberculosis, HIV-1 and Helicobacter pylori. Nat Immunol 2009; 10: 1081–88.
- 47 Ishikawa E, Ishikawa T, Morita YS, et al. Direct recognition of the mycobacterial glycolipid, trehalose dimycolate, by C-type lectin Mincle. J Exp Med 2009; 206: 2879–88.
- 48 Coulombe F, Divangahi M, Veyrier F, et al. Increased NOD2-mediated recognition of N-glycolyl muramyl dipeptide. J Exp Med 2009; 206: 1709–16.
- 49 Eum SY, Kong JH, Hong MS, et al. Neutrophils are the predominant infected phagocytic cells in the airways of patients with active pulmonary TB. Chest 2010; 137: 122–28.
- 50 Martineau AR, Newton SM, Wilkinson KA, et al. Neutrophil-mediated innate immune resistance to mycobacteria. *J Clin Invest* 2007; 117: 1988–94.
- 51 Persson YA, Blomgran-Julinder R, Rahman S, Zheng L, Stendahl O. Mycobacterium tuberculosis-induced apoptotic neutrophils trigger a pro-inflammatory response in macrophages through release of heat shock protein 72, acting in synergy with the bacteria. Microbes Infect 2008; 10: 233–40.
- 52 Baeke F, Takiishi T, Korf H, Gysemans C, Mathieu C. Vitamin D: modulator of the immune system. Curr Opin Pharmacol 2010; 10: 482–96
- 53 Wang TT, Nestel FP, Bourdeau V, et al. Cutting edge: 1,25-dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression. J Immunol 2004; 173: 2909–12.
- 54 Martineau AR, Timms PM, Bothamley GH, et al. High-dose vitamin D, during intensive-phase antimicrobial treatment of pulmonary tuberculosis: a double-blind randomised controlled trial. *Lancet* 2011; 377: 242–50.
- Paige C, Bishai WR. Penitentiary or penthouse condo: the tuberculous granuloma from the microbe's point of view. Cell Microbiol 2010; 12: 301–09.
- 56 Zumla A, James DG. Granulomatous infections: etiology and classification. Clin Infect Dis 1996; 23: 146–58.
- 57 Russell DG. Who puts the tubercle in tuberculosis? Nat Rev Microbiol 2007; 5: 39–47.
- 58 Rubin EJ. The granuloma in tuberculosis—friend or foe? N Engl J Med 2009; 360: 2471–73.
- 59 Russell DG, Barry CE III, Flynn JL. Tuberculosis: what we don't know can, and does, hurt us. Science 2010; 328: 852–56.
- 60 Russell DG, VanderVen BC, Lee W, et al. Mycobacterium tuberculosis wears what it eats. Cell Host Microbe 2010; 8: 68–76
- 61 Behar SM, Divangahi M, Remold HG. Evasion of innate immunity by Mycobacterium tuberculosis: is death an exit strategy? Nat Rev Microbiol 2010; 8: 668–74.
- 62 Bezuidenhout J, Schneider JW. Pathology and pathogenesis of tuberculosis. In: Schaaf S, Zumla A, eds. Tuberculosis: a comprehensive clinical reference. Philadelphia, PA: Saunders, 2009: 117–128.
- 63 Lawn SD, Bekker L-G. Co-pathogenesis of tuberculosis and HIV. In: Schaaf S, Zumla A, eds. Tuberculosis: a comprehensive clinical reference. Philadelphia, PA: Saunders, 2009: 96–106.
- 64 Barry CE III, Boshoff HI, Dartois V, et al. The spectrum of latent tuberculosis: rethinking the biology and intervention strategies. Nat Rev Microbiol 2009; 7: 845–55.
- 65 Young DB, Gideon HP, Wilkinson RJ. Eliminating latent tuberculosis. *Trends Microbiol* 2009; 17: 183–88.
- 66 Zumla A, Atun R, Maeurer M, et al. Scientific dogmas, paradoxes and mysteries of latent Mycobacterium tuberculosis infection. Trop Med Int Health 2011; 16: 79–83.
- 67 Lawn SD, Wood R, Wilkinson RJ. Changing concepts of "latent tuberculosis infection" in patients living with HIV infection. Clin Devel Immunol 2011; 2011: 980594.
- 68 Wallis RS, Doherty TM, Onyebujoh P, et al. Biomarkers for tuberculosis disease activity, cure, and relapse. *Lancet Infect Dis* 2009; 9: 162–72.

- 69 Wallis RS, Pai M, Menzies D, et al. Biomarkers and diagnostics for tuberculosis: progress, needs, and translation into practice. *Lancet* 2010: 375: 1920–37.
- 70 Berry MP, Graham CM, McNab FW, et al. An interferon-inducible neutrophil-driven blood transcriptional signature in human tuberculosis. *Nature* 2010; 466: 973–77.
- 71 Agranoff D, Fernandez-Reyes D, Papadopoulos MC, et al. Identification of diagnostic markers for tuberculosis by proteomic fingerprinting of serum. *Lancet* 2006; 368: 1012–21.
- 72 Repsilber D, Kern S, Telaar A, et al. Biomarker discovery in heterogeneous tissue samples—taking the in-silico deconfounding approach. BMC Bioinformatics 2010; 11: 27.
- 73 Black GF, Thiel BA, Ota MO, et al. Immunogenicity of novel DosR regulon-encoded candidate antigens of Mycobacterium tuberculosis in three high-burden populations in Africa. Clin Vaccine Immunol 2009; 16: 1203–12.
- 74 Sutherland JS, de Jong BC, Jeffries DJ, Adetifa IM, Ota MO. Production of TNF-α, IL-12(p40) and IL-17 can discriminate between active TB disease and latent infection in a West African cohort. PLoS One 2010; 5: e12365.
- 75 Jacobsen M, Repsilber D, Gutschmidt A, et al. Candidate biomarkers for discrimination between infection and disease caused by Mycobacterium tuberculosis. J Mol Med 2007; 85: 613–21.
- 76 Mistry R, Cliff JM, Clayton CL, et al. Gene-expression patterns in whole blood identify subjects at risk for recurrent tuberculosis. *J Infect Dis* 2007; 195: 357–65.
- 77 Maertzdorf J, Repsilber D, Parida SK, et al. Human gene expression profiles of susceptibility and resistance in tuberculosis. Genes Immun 2010; published online Sept 23. DOI:10.1038/gene.2010.51.
- 78 Pai M, Minion J, Sohn H, Zwerling A, Perkins MD. Novel and improved technologies for tuberculosis diagnosis: progress and challenges. Clin Chest Med 2009; 30: 701–16.
- 79 Steingart KR, Ng V, Henry M, et al. Sputum processing methods to improve the sensitivity of smear microscopy for tuberculosis: a systematic review. *Lancet Infect Dis* 2006; 6: 664–74.
- 80 Steingart KR, Henry M, Ng V, et al. Fluorescence versus conventional sputum smear microscopy for tuberculosis: a systematic review. Lancet Infect Dis 2006; 6: 570–81.
- 81 WHO. Report of the ninth meeting of the strategic and technical advisory group for tuberculosis. Geneva: World Health Organization, 2009. http://www.who.int/tb/advisory_bodies/ stag_tb_report_2009.pdf (accessed Aug 16, 2010).
- WHO. New laboratory diagnostic tools for tuberculosis control. Geneva: World Health Organization, 2008. http://whqlibdoc.who. int/publications/2008/9789241597487_eng.pdf (accessed Jan 18, 2010).
- 83 Morgan M, Kalantri S, Flores L, Pai M. A commercial line probe assay for the rapid detection of rifampicin resistance in Mycobacterium tuberculosis: a systematic review and meta-analysis. BMC Infect Dis 2005; 5: 62.
- 84 Ling DI, Zwerling AA, Pai M. GenoType MTBDR assays for the diagnosis of multidrug-resistant tuberculosis: a meta-analysis. Eur Respir J 2008; 32: 1165–74.
- 85 Hillemann D, Rusch-Gerdes S, Richter E. Feasibility of the GenoType MTBDRsI assay for fluoroquinolone, amikacin-capreomycin, and ethambutol resistance testing of Mycobacterium tuberculosis strains and clinical specimens. J Clin Microbiol 2009; 47: 1767–72.
- 86 Lawn SD, Edwards DJ, Kranzer K, Vogt M, Bekker LG, Wood R. Urine lipoarabinomannan assay for tuberculosis screening before antiretroviral therapy diagnostic yield and association with immune reconstitution disease. AIDS 2009; 23: 1875–80.
- 87 Shah M, Variava E, Holmes CB, et al. Diagnostic accuracy of a urine lipoarabinomannan test for tuberculosis in hospitalized patients in a High HIV prevalence setting. J Acquir Immune Defic Syndr 2009; 52: 145–51.
- 88 Peter J, Green C, Hoelscher M, Mwaba P, Zumla A, Dheda K. Urine for the diagnosis of tuberculosis: current approaches, clinical applicability, and new developments. Curr Opin Pulm Med 2010; 16: 262–70.
- Greco S, Girardi E, Navarra A, Saltini C. Current evidence on diagnostic accuracy of commercially based nucleic acid amplification tests for the diagnosis of pulmonary tuberculosis. *Thorax* 2006; 61: 783–90.

- 90 Ling DI, Flores LL, Riley LW, Pai M. Commercial nucleic-acid amplification tests for diagnosis of pulmonary tuberculosis in respiratory specimens: meta-analysis and meta-regression. *PLoS One* 2008; 3: e1536.
- 91 Boehme CC, Nabeta P, Henostroza G, et al. Operational feasibility of using loop-mediated isothermal amplification for diagnosis of pulmonary tuberculosis in microscopy centers of developing countries. J Clin Microbiol 2007; 45: 1936–40.
- 92 Helb D, Jones M, Story E, et al. Rapid detection of Mycobacterium tuberculosis and rifampin resistance by use of on-demand, near-patient technology. J Clin Microbiol 2010; 48: 229–37.
- 93 Boehme CC, Nabeta P, Hillemann D, et al. Rapid molecular detection of tuberculosis and rifampin resistance. N Engl J Med 2010; 363: 1005–15.
- 94 Pai M, Zwerling A, Menzies D. Systematic review: T-cell-based assays for the diagnosis of latent tuberculosis infection: an update. Ann Intern Med 2008; 149: 177–84.
- 95 Adetifa IM, Ota MO, Walther B, et al. Decay kinetics of an interferon gamma release assay with anti-tuberculosis therapy in newly diagnosed tuberculosis cases. PLoS One 2010; 5: e12502.
- 96 Aiken AM, Hill PC, Fox A, et al. Reversion of the ELISPOT test after treatment in Gambian tuberculosis cases. BMC Infect Dis 2006: 6: 66.
- 97 Ribeiro S, Dooley K, Hackman J, et al. T-SPOT.TB responses during treatment of pulmonary tuberculosis. BMC Infect Dis 2009; 9: 23.
- 98 Losi M, Bossink A, Codecasa L, et al. Use of a T-cell interferon-gamma release assay for the diagnosis of tuberculous pleurisy. Eur Respir J 2007; 30: 1173–79.
- 99 Mazurek GH, Weis SE, Moonan PK, et al. Prospective comparison of the tuberculin skin test and 2 whole-blood interferon-gamma release assays in persons with suspected tuberculosis. Clin Infect Dis 2007; 45: 837–45.
- 100 Ruhwald M, Petersen J, Kofoed K, et al. Improving T-cell assays for the diagnosis of latent TB infection: potential of a diagnostic test based on IP-10. PLoS One 2008: 3: e2858.
- 101 Hoa NB, Sy DN, Nhung NV, Tiemersma EW, Borgdorff MW, Cobelens FG. National survey of tuberculosis prevalence in Viet Nam. Bull World Health Organ 2010; 88: 273–80.
- 102 Elliott AM, Halwiindi B, Hayes RJ, et al. The impact of human immunodeficiency virus on presentation and diagnosis of tuberculosis in a cohort study in Zambia. J Trop Med Hyg 1993; 96: 1–11.
- 103 Gilks CF, Brindle RJ, Otieno LS, et al. Extrapulmonary and disseminated tuberculosis in HIV-1-seropositive patients presenting to the acute medical services in Nairobi. AIDS 1990; 4-981-85
- 104 Getahun H, Kittikraisak W, Heilig CM, et al. Development of a standardized screening rule for tuberculosis in people living with hiv in resource-constrained settings: individual participant data meta-analysis of observational studies. PLoS Med 2011; 8: e1000391.
- 105 Kranzer K, Houben RM, Glynn JR, Bekker LG, Wood R, Lawn SD. Yield of HIV-associated tuberculosis during intensified case finding in resource-limited settings: a systematic review and meta-analysis. *Lancet Infect Dis* 2010; 10: 93–102.
- 106 Monkongdee P, McCarthy KD, Cain KP, et al. Yield of acid-fast smear and mycobacterial culture for tuberculosis diagnosis in people with human immunodeficiency virus. Am J Respir Crit Care Med 2009; 180: 903–08.
- 107 WHO. Treatment of tuberculosis: guidelines—4th edn. Geneva: World Health Organization, 2010. http://whqlibdoc.who.int/ publications/2010/9789241547833_eng.pdf (accessed Nov 30, 2010).
- 108 Jindani A, Nunn AJ, Enarson DA. Two 8-month regimens of chemotherapy for treatment of newly diagnosed pulmonary tuberculosis: international multicentre randomised trial. *Lancet* 2004; 364: 1244–51.
- 109 Mak A, Thomas A, Del GM, Zaleskis R, Mouzafarova N, Menzies D. Influence of multidrug resistance on tuberculosis treatment outcomes with standardized regimens. Am J Respir Crit Care Med 2008; 178: 306–12.
- 110 Menzies D, Benedetti A, Paydar A, et al. Standardized treatment of active tuberculosis in patients with previous treatment and/or with mono-resistance to isoniazid: a systematic review and meta-analysis. PLoS Med 2009; 6: e1000150.

- 111 Menzies D, Benedetti A, Paydar A, et al. Effect of duration and intermittency of rifampin on tuberculosis treatment outcomes: a systematic review and meta-analysis. PLoS Med 2009; 6: e1000146.
- 112 Pillay M, Sturm AW. Evolution of the extensively drug-resistant F15/LAM4/KZN strain of Mycobacterium tuberculosis in KwaZulu-Natal, South Africa. Clin Infect Dis 2007; 45: 1409–14.
- 113 Caminero JA, Sotgiu G, Zumla A, Migliori GB. Best drug treatment for multidrug-resistant and extensively drug-resistant tuberculosis. *Lancet Infect Dis* 2010; 10: 621–29.
- 114 Orenstein EW, Basu S, Shah NS, et al. Treatment outcomes among patients with multidrug-resistant tuberculosis: systematic review and meta-analysis. *Lancet Infect Dis* 2009; 9: 153–61.
- 115 Dheda K, Shean K, Zumla A, et al. Early treatment outcomes and HIV status of patients with extensively drug-resistant tuberculosis in South Africa: a retrospective cohort study. *Lancet* 2010; 375: 1798–807.
- 116 Gandhi NR, Moll A, Sturm AW, et al. Extensively drug-resistant tuberculosis as a cause of death in patients co-infected with tuberculosis and HIV in a rural area of South Africa. *Lancet* 2006; 368: 1575–80
- 117 Mitnick CD, Shin SS, Seung KJ, et al. Comprehensive treatment of extensively drug-resistant tuberculosis. N Engl J Med 2008; 359: 563–74.
- 118 Harries AD, Zachariah R, Corbett EL, et al. The HIV-associated tuberculosis epidemic—when will we act? *Lancet* 2010; 375: 1906–19.
- 119 Odhiambo J, Kizito W, Njoroge A, et al. Provider-initiated HIV testing and counselling for TB patients and suspects in Nairobi, Kenya. Int J Tuberc Lung Dis 2008; 12 (suppl 1): 63–68.
- 120 Lawn SD, Fraenzel A, Kranzer K, Caldwell J, Bekker LG, Wood R. Provider initiated HIV testing increases access of patients with HIV-associated tuberculosis to antiretroviral therapy. S Afr Med J (in press).
- 121 Harries AD, Zachariah R, Lawn SD. Providing HIV care for co-infected tuberculosis patients: a perspective from sub-Saharan Africa. *Int J Tuberc Lung Dis* 2009; **13**: 6–16.
- 122 Lawn SD, Kranzer K, Wood R. Antiretroviral therapy for control of the HIV-associated tuberculosis epidemic in resource-limited settings. Clin Chest Med 2009; 30: 685–99.
- 123 Lawn SD, Torok ME, Wood R. Optimum time to start antiretroviral therapy during HIV-associated opportunistic infections. Curr Opin Infect Dis 2011; 24: 34–42.
- 124 Abdool Karim SS, Naidoo K, Grobler A, et al. Timing of initiation of antiretroviral drugs during tuberculosis therapy. N Engl J Med 2010; 362: 697–706.
- 125 Blanc F-X, Sok T, Laureillard D, et al. Significant enhancement in survival with early (2 weeks) vs late (8 weeks) initiation of highly active antiretroviral treatment (HAART) in severely immunosuppressed HIV-infected adults with newly diagnosed tuberculosis. XVIII International AIDS Conference; Vienna, Austria; July 18–23, 2010. Abstract THLBB1.
- 126 WHO. Antiretroviral therapy for HIV infection in adults and adolescents: recommendations for a public health approach (2010 version). Geneva: World Health Organization, 2010. http://www.who.int/hiv/pub/arv/adult2010/en/index.html (accessed Nov 19, 2010).
- 127 McIlleron H, Meintjes G, Burman WJ, Maartens G. Complications of antiretroviral therapy in patients with tuberculosis: drug interactions, toxicity, and immune reconstitution inflammatory syndrome. J Infect Dis 2007; 196 (suppl 1): S63–75.
- 128 Lawn SD, Myer L, Bekker LG, Wood R. Burden of tuberculosis in an antiretroviral treatment programme in sub-Saharan Africa: impact on treatment outcomes and implications for tuberculosis control. AIDS 2006; 20: 1605–12.
- 129 Hoffmann CJ, Charalambous S, Thio CL, et al. Hepatotoxicity in an African antiretroviral therapy cohort: the effect of tuberculosis and hepatitis B. AIDS 2007; 21: 1301–08.
- 130 Maartens G, Decloedt E, Cohen K. Effectiveness and safety of antiretrovirals with rifampicin: crucial issues for high-burden countries. Antivir Ther 2009; 14: 1039–43.
- 131 Lawn SD, Bekker LG, Miller RF. Immune reconstitution disease associated with mycobacterial infections in HIV-infected individuals receiving antiretrovirals. *Lancet Infect Dis* 2005; 5: 361–73.

- 132 Muller M, Wandel S, Colebunders R, Attia S, Furrer H, Egger M. Immune reconstitution inflammatory syndrome in patients starting antiretroviral therapy for HIV infection: a systematic review and meta-analysis. Lancet Infect Dis 2010; 10: 251–61.
- 133 Meintjes G, Wilkinson RJ, Morroni C, et al. Randomized placebo-controlled trial of prednisone for paradoxical tuberculosis-associated immune reconstitution inflammatory syndrome. AIDS 2010; 24: 2381–90.
- 134 Ma Z, Lienhardt C, McIlleron H, Nunn AJ, Wang X. Global tuberculosis drug development pipeline: the need and the reality. *Lancet* 2010; 375: 2100–09.
- 135 Diacon AH, Pym A, Grobusch M, et al. The diarylquinoline TMC207 for multidrug-resistant tuberculosis. N Engl J Med 2009; 360: 2397–405.
- 136 Stover CK, Warrener P, VanDevanter DR, et al. A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis. Nature 2000; 405: 962–66.
- 137 Kaufmann SH, Hussey G, Lambert PH. New vaccines for tuberculosis. *Lancet* 2010; 375: 2110–19.
- 138 Lambert PH, Hawkridge T, Hanekom WA. New vaccines against tuberculosis. Clin Chest Med 2009; 30: 811–26.
- 139 Trunz BB, Fine P, Dye C. Effect of BCG vaccination on childhood tuberculous meningitis and miliary tuberculosis worldwide: a meta-analysis and assessment of cost-effectiveness. *Lancet* 2006; 367: 1173–80.
- 140 Stanford J, Stanford C, Grange J. Immunotherapy with Mycobacterium vaccae in the treatment of tuberculosis. Front Biosci 2004; 9: 1701–19.
- 141 Von Reyn CF, Mtei L, Arbeit R, et al. Prevention of tuberculosis in BCG-primed, HIV-infected adults boosted with an inactivated whole cell mycobacterial vaccine. AIDS 2010; 24: 675–85.
- 142 Stop TB Partnership. The global plan to stop TB 2006–2015. Geneva: World Health Organization, 2006. http://www.stoptb.org/assets/documents/global/plan/GlobalPlanFinal.pdf (accessed Nov 30, 2010).
- 143 Stop TB Partnership, WHO. The stop TB strategy: building on and enhancing DOTS to meet the TB-related Millennium Development Goals. Geneva: World Health Organization, 2006. http://www.who. int/tb/publications/2006/stop_tb_strategy.pdf (accessed Nov 30, 2010).

- 144 Stop TB Partnership. The global plan to stop TB 2011–2015. Geneva: World Health Organization, 2010. http://www.stoptb.org/assets/documents/global/plan/TB_GlobalPlanToStopTB2011-2015.pdf (accessed Nov 30, 2010).
- 145 Lonnroth K, Jaramillo E, Williams BG, Dye C, Raviglione M. Drivers of tuberculosis epidemics: the role of risk factors and social determinants. Soc Sci Med 2009; 68: 2240–46.
- 146 Oxlade O, Schwartzman K, Behr MA, et al. Global tuberculosis trends: a reflection of changes in tuberculosis control or in population health? *Int J Tuberc Lung Dis* 2009; 13: 1238–46.
- 147 Atun R, Weil DE, Eang MT, Mwakyusa D. Health-system strengthening and tuberculosis control. *Lancet* 2010; 375: 2169–78.
- 148 WHO. Towards universal access: scaling up priority HIV/AIDS interventions in the health sector—progress report 2010.
 Geneva: World Health Organization, 2010. http://www.who.int/hiv/pub/2010progressreport/summary_en.pdf (accessed Jan 12, 2011).
- 149 WHO. WHO three I's meeting: intensified case finding (ICF), isoniazid preventive therapy (IPT) and TB infection control (IC) for people living with HIV. http://www.who.int/hiv/pub/meetingreports/WHO_3Is_meeting_report.pdf (accessed Nov 30, 2010).
- 150 Lawn SD, Wood R, De Cock KM, Kranzer K, Lewis JJ, Churchyard GJ. Antiretrovirals and isoniazid preventive therapy in the prevention of HIV-associated tuberculosis in settings with limited health-care resources. *Lancet Infect Dis* 2010; 10: 489–98.
- 151 Lawn SD, Myer L, Edwards D, Bekker LG, Wood R. Short-term and long-term risk of tuberculosis associated with CD4 cell recovery during antiretroviral therapy in South Africa. AIDS 2009; 23: 1717–25.
- 152 Lawn SD, Harries AD, Williams BG, et al. Will ART do it? Antiretroviral therapy and the control of HIV-associated tuberculosis. Int J Tuberc Lung Dis (in press).
- 153 Marais BJ, Raviglione MC, Donald PR, et al. Scale-up of services and research priorities for diagnosis, management, and control of tuberculosis: a call to action. *Lancet* 2010; 375: 2179–91.