First Coho Salmon caught on Piledriver Slough in August, 2021!

Introduction
This report discloses the findings of the 2021 study of fisheries, beaver activity, and water quality on Piledriver Slough and 23 mile Slough. The study was conducted by the Tanana Valley Watershed Association (TVWA) contractors with assistance from Salcha Elementary School. Survey site results are discussed below.

Purpose
2021 was the tenth year of a ten-year study to fulfill Mitigation Measure 56 of the Service Transportation Board. The measure states, “prior to construction of Salcha Alternative Segment 1, ARRC shall develop appropriate mitigation in consultation with ADF&G to prevent blockage of Piledriver and 23 Mile Sloughs by beaver dams (as a result of flushing flows caused by ARRC-proposed channel plugs). Mitigation may include monitoring conducted by ARRC at a frequency agreed to by ADF&G.” The Piledriver Slough Mitigation Plan was created to assess impacts of the Northern Rail Extension Project-Phase 1.

Need
In 2011 a levee was put into place to alleviate blockage to spring flow flushing from the Tanana River into the Piledriver Slough due to construction of the new rail extension. With the construction of the levee, concerns were raised about the potential alteration in flow-rate because of the lacking ability of natural flushing of debris or ice build-up by spring flows. Resulting concerns include ice
and log jams and beaver dams impeding fish passage. This study was created to assess the risk that such obstructions pose to fish passage.

**Objectives**

The Alaska Department of Fish and Game (AKDFG) consults TVWA in action through a Memorandum of Agreement implementing fish monitoring within the Piledriver and 23 mile sloughs. TVWA is charged to manage the Piledriver Slough Beaver Activity Survey program until 2022, in which a final report will be submitted to AKDF&G and the Alaska Rail Road (AKRR). The report will compile results and conclusions drawn from outlined objectives and accomplishments achieved during the 10-year study.

*TVWA field technician Lauren Schour shows a Salcha student how to use the turbidimeter during June 2021 sampling.*

**Methodology**

The ten-mile section of the Piledriver Slough was divided into two sections for managing monitoring based upon distance from the levee site to the Bailey Bridge. These sections were the *Upper Piledriver* and *Lower Piledriver*. Upper Piledriver surveying began from the levee site and ended at the Old Valdez Trail road crossing. This section was surveyed by TVWA staff with the assistance of citizen scientists from Salcha Elementary. Lower Piledriver surveying began from the
Old Valdez Trail road crossing and ended at the Bailey Bridge, adjacent to Eielson Airforce Base. This section was surveyed by TVWA staff. Undivided, the 23 Mile Slough site was located and surveyed in its entirety off of Old Eielson Farm Road. All surveys took place late spring, summer, and late fall, with exact dates dependent on staff and school availability.

For the study of Upper Piledriver, TVWA trained volunteers and student citizen scientists to collaborate in the research process. The students were trained in water safety, fish, plant and invertebrate identification, fish handling, water quality, invasive species, aquatic invertebrates and habitat assessment. Each child was equipped with a tool kit containing supplies and safety for the field surveying. Algae and aquatic plant identification education curriculum was added in 2014. Fish factsheets, tracks sheets, and more complex habitat assessments were added in 2015. Compasses and magnifying boxes were added to the curriculum in 2016. In 2017, a curriculum was added on water velocity and flow. 2018’s new curriculum was centered on turbidity, pH in the classroom and insects. Our 2019 focus was on elements of water quality, especially dissolved oxygen. In 2020 the Salcha teachers Tori Brannan and Samantha Wills were trained in the GLOBE citizen science program and began using some new data protocols to measure cloud cover, turbidity, and using a dissolved oxygen probe. During June, sampling Salcha teachers were trained by UAF scientist Christina Buffington in GLOBE protocols including weather surveys. During September, sampling a DO meter was used for the first time to compare results from the kids. In 2021 TVWA purchased a HATCH turbidimeter and trained students to use this under supervision. TVWA also conducted a staff training with Salcha Elementary on August 13, 2021 to assist the school with staff turnover.

*Equipment:* Equipment used in the study by TVWA staff were a Garmin GPS 62s, PentaxWGIII SR Adventure Proof GPS Camera, GoPro video camera, Android telephone camera for capturing photos and videos to be used for analysis and reporting. GPS units were used for marking identified dams and lodges as well as geo-referencing photos.
Water Quality Sampling: TVWA’s “Adopt-a-Stream” water quality sampling protocol was used to record water quality at each Upper Piledriver Site and five sites on Lower Piledriver Slough. This data was then submitted to the Dept. of Environmental Quality. This protocol is detailed below:

**Step 1: Perform a Hanna meter pre-sampling check with tap water.** Using the pH 4 and 1413 conductivity standards provided, test your meter’s accuracy. Turn on your meter. Place a small amount of the pH 4 standard into a plastic cup marked “pH4 check” (just enough to cover the sensor). Take a pH reading and record the result. It should fall between 3.8 and 4.2. Rinse the meter in tap water and shake it gently to remove excess water. Then, place a small amount of the 1413 conductivity standard into the plastic cup marked “conductivity check” and take a reading. Note the conductivity level. It should fall between 1342 and 1484. Rinse the meter again in tap water and shake it gently to remove excess water before replacing the cap. The standards are safe to pour down the drain with a little tap water. DO NOT pour them into the stream.

**Step 2: Collect water sample:** A few yards away (preferably downstream or down current) from your exact sampling site, rinse the plastic bucket three times with stream water. Then go to your site and, facing upstream, lower the bucket gently into the water, and fill it to a level about 2 inches from the lip of the bucket. If you are working in very shallow water, do not disturb the bottom while collecting the sample.

**Step 3: Measure pH and Conductivity with Hanna Meter:** Turn on the meter. Hold it or clip it to the side of the bucket in the sample water for 5 minutes. Turn on the meter. Press SET/HOLD.
until it is in conductivity (μ) mode, wait 15 seconds, then record three (3) sequential readings for Conductivity at 15 second intervals. Press SET/HOLD until it is in pH mode and wait 15 seconds. Record three (3) pH readings at 15 second intervals. Finally, press SET/HOLD until it is in temperature mode and wait 15 seconds. Record three (3) water temperature readings at 15 second intervals. Turn the meter off. Put the cover back on the meter, making sure to moisten the pH sensor before doing so.

Step 4: Record the air temperature: Hang the air thermometer somewhere where it will not lean against any solid object and where it is protected as much as possible from direct wind and sunlight. The thermometer will take at least five minutes to equilibrate. It might take longer if it has to adjust for large changes in temperature. Recording the air temperature after you have completed the water quality sampling, which should ensure that the thermometer has had ample time to adjust.

Step 5: Perform the meter post-sampling check in office with tap water: Using the pH 10 and 1413 conductivity standards provided, test your meter’s accuracy. Turn on your meter. Place a small amount of the pH 10 standard into a plastic cup marked “pH10 check” (just enough to cover the sensor). Take a pH reading and record the result. It should fall between 9.8 and 10.2. Rinse the meter in tap water and shake it gently to remove excess water. Then, place a small amount of the 1413 conductivity standard into the plastic cup marked “conductivity check” and take a reading. Note the conductivity level. It should fall between 1342 and 1484. Rinse the meter again in tap water and shake it gently to remove excess water before replacing the cap. The standards are safe to pour down the drain with a little tap water. DO NOT pour them into the stream.
Fish Sampling: TVWA’s “Chena Salmon” sampling protocol was used for recording information on fish. Gee-type minnow traps (23 x 45 cm, 0.64 cm wire mesh, with 2.5 cm diameter openings) were baited with salmon roe and set 5-10 mm apart for a 24-hour soak time (Swales, 1987). After the 24 hour soak, scientists identify and count all fish in the trap and determine length using a Photarium viewing box (Duvall, WA, USA). Fish were released after identification and measurements were taken. Any incidental fish deaths were labeled and brought to the USFWS laboratory in Fairbanks for further processing.

Sampling procedures:

1. Set Traps:
   - Place bait ball in the trap.
   - Put trap in suitable location length-wise to current. Slow moving water with in-stream cover is best but this may not be possible at all sites. Put traps in the slowest moving water available at your site because fish will get exhausted swimming against current.
   - Let your trap soak overnight and check on it 24 hours later.
   - Be as consistent as possible with length of soak.
   - Get traps in deep enough water to cover the trap (deeper is better).
   - Don’t put traps in a high use area because they may get vandalized or stolen.

Salcha students with lake chub at site 4 during August 2021 sampling.
• Make sure that traps are well-secured to something on the bank.

2. Checking Traps
• Have all of your equipment ready before removing any traps from the water.
• Fill your counting and holding buckets half full of river water.
• Remove one of your traps from the water and gently pour fish into your counting buckets.
• Catch one fish at a time with the dip net and place it in the viewing box to identify it.
• Use an identification guide to identify fish.
• Record length of each first using length markings on the photarium.
• After identification, put fish into the holding bucket.
• After you are finished counting and identifying all of the fish from one trap gently pour the holding bucket into the river and start counting your next trap.
• Record total numbers for each species on the datasheet, and if no fish are caught record that.

3. Fish Handling Guidelines
• Keep your hands wet at all times.
• Use bare hands, gloves can damage scales.
• Handle fish as little as possible.
• Only empty one trap into the counting bucket at a time (to maximize oxygen content).
• Release fish in the same place where you caught them.

Beaver Survey: Beaver dams and lodges were surveyed visually by foot on Upper Piledriver Slough and by canoe on Lower Piledriver. Beaver dams were defined as dams built by beavers to provide ponds as protection against predators such as coyotes, wolves, and bears, and to provide easy access to food during winter. Beaver lodges were defined as dwellings constructed on the side of the stream that do not impede passage. All dams and lodges were photographed, GPS locations were recorded, and the sites were described. Dams were measured for height, diameter of logs and width of passage. Dams were categorized based on activity by beavers (active, inactive) and type of dwelling (primary dam, secondary dam, lodge). Active was defined as dams or lodges that exhibited signs of recent activity including fresh chews, moved materials, feed piles, tracks, beaver slides, or beaver presence, etc. Inactive dams and lodges were defined as places which did not exhibit the signs of use identified in the “active” definition. Primary dam was considered the largest dam in a ½ mile area that displayed the most use. Secondary dam was determined to be a smaller dam.

Follow Up:
All equipment was inventoried, cleaned, and serviced before and after the surveying season. Fish data reports were sent to the Alaska Dept of Fish and Game, in compliance with our permit requirement.
Salcha Elementary principal Tori Brannan assists a Salcha student with measuring a lake chub caught during June 2021 sampling.

**Study Survey Results**

The 2021 sampling was altered by the Covid-19 pandemic but we still managed to maintain strong community involvement. Social distancing, mask-wearing, and hand-sanitizing best practices were followed. In May, Salcha Elementary school assisted TVWA contractors Heather Mirczak and David Jonas in the sampling of Upper Piledriver. In June, TVWA contractor David Jonas and employees Jenna Jonas and Lauren Schour ran sampling with the assistance of a small group of Salcha students and parent volunteers. In August, the Salcha Elementary school braved cold temperatures to assist a full crew of Jenna and David Jonas, Heather Mirczak, Ashley Carrick, Christy Everett and Janet Kidd with sampling.

The study had a total of twenty-eight survey sites. Eight survey sites (with 2 traps each) on the Upper Piledriver were within the periods of May 13-14, June 20-21 and August 26-27. The Upper Piledriver was monitored with the assistance of the Salcha Elementary School through the citizen scientist collaboration. Sixteen sites (with one trap each) were surveyed on Lower Piledriver by TVWA field technicians and volunteers on May 13-14, June 20-21 and August 26-27. Undivided, 23 Mile Slough had four survey locations that took place on June 20-21 and September 3-4 by TVWA field technicians and volunteers. TVWA staff included supervision by Ashley Carrick. Staff included
Jenna Jonas and Lauren Schour. Contractors were David Jonas and Heather Mirczak. Interns (who were Salcha Elementary alumni) included Rusty Baker and Teslin Brannan. TVWA volunteers included Christy Everett and Janet Kidd. 67 students and 27 teachers/volunteers participated in the study this year.

In 2015, TVWA staff began recording qualitative data after each float and continued this practice in 2021. Water quality data was also recorded and submitted to the Alaska Dept. of Environmental Conservation’s ACWA program.

*Fish:* AKDF&G issued TVWA a Fish Resource Permit for the study (See Appendix A). Surveying took place post-permit issuance. Data collection recorded fish species identified, relative size, and location assisted by equipment (minnow traps, viewer, bucket, and identification book). The compilation of fish parameters was reported to AKRR as the *Fish Collection Report* (See Appendix B). Fish monitoring was conducted at 28 sites with a total of 87 caught-and-release fish recorded.

*Beaver:* Beaver dams were categorized based on whether or not it was actively used by beavers, which simply were active or inactive. Secondary categorization was based on dwelling type of dam, which consisted of primary dam, secondary dam and lodge. Dam activity and dwelling type was recorded as well as coordinates.

## Fish Collection Report

### Summary

In 2021, 87 fish were caught, identified and released in Piledriver and 23 mile sloughs. Of these, 77 were caught on Upper Piledriver with the Salcha Elementary students, 9 on Lower Piledriver; and 1 on 23 mile slough. This was a big catch on Upper Piledriver compared to other years. Most of the fish caught were lake chub which were aggregating during August sampling.

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<th>Place</th>
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<th>Slimy Sculpin</th>
<th>Lake Chub</th>
<th>Coho Salmon</th>
<th>Arctic Grayling</th>
<th>Chinook Salmon</th>
<th>Days Sampled</th>
<th># traps set</th>
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<td>3</td>
<td>1</td>
<td>0</td>
<td>May 13-14, June</td>
<td>16</td>
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</table>
Equipment Used

Gee-type minnow traps (23 x 45 cm, 0.64 cm bar mesh, with 2.5 cm diameter opening) were baited with disinfected salmon roe and set for 24 hours for each sampling event. Traps were placed in a variety of habitat types including cut banks, slough mouths, in woody debris, and on either side of beaver dams. All captured fish were identified to species. The fork length of the fish identified at each site each week was measured using the ruler on a medium Photarium viewing box (Duvall, WA). Fish were released after identification and measurement.

Species Diversity

The most commonly caught fish in 2021 was the Lake Chub (57 fish). The second most commonly caught fish was Slimy Sculpin (26 fish). The third most common fish caught was Coho Salmon (3 fish). The coho salmon captured on lower piledriver slough during the August sampling were the first recorded in this study. These fish were photographed and sent to AKDF&G staff to confirm the identification. Per our permit requirement, TVWA submitted this finding to the anadromous waters catalog and it was confirmed on October 28. In past years, the Slimy Sculpin has been the most commonly caught fish except for when sampling coincided with lake chub spawning events. This
Year’s high catch of lake chub occurred during the August sampling which is outside of the normal spawning period for this species. We are uncertain why the large aggregation of lake chub were caught then but they were visually observed during this period on Lower Piledriver as well.

The slimy sculpin (Cottus cognatus), a bottom-dwelling fish, can be found throughout most of the northern United States, Canada and Alaska. The slimy sculpin is an ambush predator. It feeds primarily on insects, but also eats crustaceans, fish eggs, and small fish. The slimy sculpins’ size and poor swimming ability makes it a great prey item for larger fish. The slimy sculpin has been studied in waters where there is current acidification (water that is more acidic). The sculpin were found to be less active and have lower rates of reproduction when found in these waters. For these reasons, the slimy sculpin has been identified as a good indicator species (a species that indicates a change in environment by a difference in behavior or population size) for acidification in lakes and ponds and possibly for streams.¹

Our water quality data thus far does not show that acidification is occurring in Piledriver slough, the challenge that we have observed is an increased sedimentation and increasing temperature. Interestingly, the Slimy Sculpin do fan their nests to remove silt, an adaptation that may allow them to thrive in the changing Piledriver Slough. The slimy sculpin moves to shallower waters during the spawning season, which is in the spring, usually after break-up. Males establish a nesting spot under a rock or log and groom the area by fanning fine sediment and moving small pebbles with its mouth out of the area. Males are territorial and can be aggressive towards other males. A male courts a female until she deposits her eggs, which are yellow to pink, on the underside of the rock or log. The female does this from the upside-down position while the male fertilizes the eggs with his milt. The female leaves after egg deposition. A single male may spawn with several females. Once the eggs are fertilized, the male guards his nest until the young fish are ready to leave. During this time the male fans the eggs to remove silt and provide oxygen and keeps the nest clean. The eggs hatch about 30 days after being fertilized. The sac-fry stay in the nest, usually resting on the bottom. They remain there for about a week while the yolk is being absorbed. Once the yolk-sac is gone, the sculpin leave the nest as fry.

The lake chub (Couesius plumbeus) belongs to the largest freshwater fish family, the minnows (Cyprinidae). They are a small fish, with adults averaging from 5-10 cm long. The lake chub is found in all types of freshwater bodies (lakes and streams), but in Alaska it has been found more often in silty waters. It tends to prefer shallow water, although it will move to deeper water during hot weather. The lake chub is usually abundant wherever it is found. Young lake chubs feed primarily on zooplankton. Older lake chubs feed on terrestrial and aquatic insects, but also feed on algae, occasionally small fishes, and have been known to scavenge on decaying fish.² This makes sense as our surveys of Piledriver’s aquatic invertebrates have found the habitat to be host to an extensive array of aquatic insects and we have observed increasing amounts of algae in the past few years. We additionally believe that we encountered the Lake Chub during their spawning period, which is known to occur between spring and early summer. This would account, in part, for their abundance although lake chub prefer spawning areas with shallow water and rocky or gravelly bottoms.

Visual Fish Survey

¹ “Slimy Sculpin” Alaska Dept. of Fish and Game Wildlife Notebook Series, Kelly Mansfield, 2004
² “Lake Chub” Alaska Dept. of Fish and Game Wildlife Notebook Series, Kelly Mansfield, 2004
In 2021 TVWA field staff did not observe any adult or juvenile salmon on upper or lower Piledriver Slough. 2021 was a notably bad year for Summer and Fall Chum Salmon in the Tanana River drainage, the run was late and far smaller than usual, and fishing was closed on these species. For the second year in a row, we observed fewer than usual of the schools of Arctic Grayling on Lower Piledriver Slough and attribute this lack to the prevalence of large beaver dams.

*Salcha Elementary student and teacher with 2 lake chub at Upper Piledriver Site 1 during August sampling.*
Appendix A: Fish Resource Permit: Fish Resource Permit

STATE OF ALASKA
DEPARTMENT OF FISH AND GAME
PO Box 115526
JUNEAU, ALASKA 99811-5526

AQUATIC RESOURCE PERMIT
(For Scientific/Educational/Collection Purposes)

This permit authorizes:
Jenna Jonas
(whose signature is required on page 3 for permit validation)

Of
Tanana Valley Watershed Association
2240 Railroad Drive, Fairbanks, AK 99709
(907) 568-0245 jennasjonas@gmail.com

to conduct the following activities from May 1, 2021 to October 1, 2021 in accordance with AS 16.05.010, AS 16.05.340(b), and 5 AAC 41.

Purpose: To examine fish presence and abundance in the target locations in fulfillment of Mitigation Measure 56 of the Service Transplanter Board while incorporating a Citizen Science project with Salcha Elementary School students.

Location: Placidriver Slough (AVC #334-40-11000-2490-3315), 23 Mile Slough (334-40-11000-2490-3315-4010)

Species: Local species

Method of Capture: Minnow trap (see Stipulations #5-8)

Final Disposition: Any number of fish may be captured, identified, and released alive at the site of capture during each sampling event.
350 individuals of each species encountered at each sample location may also be measured for length and/or weight prior to release.
2 individuals of each unknown species may be killed and saved for later identification.
All unprovided mortalities must be recorded and returned to capture site waters.

COLLECTION REPORT DUE: November 1, 2021 and RESEARCH REPORT DUE: April 1, 2022. See Stipulations #2 and #3 for more information. Data from such reports are considered public information. Reports must be submitted by email (dfs.divfish@alaska.gov) or by mail to: Alaska Department of Fish and Game, Division of Sport Fish-HQ, PO Box 115526, Juneau, AK 99811-5526. Attention: Permit Coordinator. A report is required whether or not collecting activities were undertaken.

GENERAL CONDITIONS, EXCEPTIONS, AND RESTRICTIONS
1. The permit must be carried by persons specified during approved activities and any person(s) specified during approved activities who shall allow it on request to persons authorized to enforce Alaska’s fish and game laws. This permit is not transferable and will be revoked or renewed biennially by the Commissioner of Fish and Game if the permittee violates any of its conditions, exceptions, or restrictions. No resubmission of authority may be allowed under this permit unless specifically released.
2. No specimens taken under authority hereof may be sold, bartered, or consumed. All specimens must be deposited in a public museum or a public scientific or educational institution unless otherwise stated herein. Subpermittees shall not retain possession of the animals or other specimens.
3. The permittee shall keep records of all activities conducted under authority of this permit, available for inspection at all reasonable hours upon request of any authorized state enforcement officer.
4. Permits will not be renewed unless specific reports, as specified in the Stipulations section, have been received by the department.
5. UNLESS SPECIFICALLY STATED HEREIN, this permit does not authorize the transportation of specimens or the taking of specimens outside of existing regulations.

Kesia Schneider
Division of Sport Fish

Director
Division of Sport Fish

2-2-2021

Authorized Personnel: The following persons may perform collecting activities under terms of this permit:
Jenna Jonas, David Jonas, Tori Brannan, Heather Mincanz, Christy Everett, Naomi Rambo, Sam,
Employees and volunteers under the direct supervision of, and in the presence of, one of the authorized personnel listed above may participate in collecting activities under terms of this permit.
Authorized Personnel: The following persons may perform collecting activities under terms of this permit:

Jonna Jonas, David Jonas, Tori Brannan, Heather Mirczak, Christy Everett, Naomi Rambo, Samantha Willis

Employees and volunteers under the direct supervision of, and in the presence of, one of the authorized personnel listed above may participate in collecting activities under terms of this permit.

Permit Stipulations:

1) Heather Scannell (459-7357; heather.scannell@alaska.gov) – the Tanana River (Fairbanks) Area Management Biologist (AMB), must be contacted a few days prior to engaging in any collecting activities for you to receive final authorization for conducting surveys. AMBs have the right to specify methods for collecting, as well as limiting the collections of any species by number, time, and location. The timeline of this contact must be included in your collections report (using the “data submission form” furnished by ADF&G).

2) A report of collecting activities, referencing this aquatic resource permit, must be submitted within 30 days after the expiration of this permit. This report should present the research conducted in a format similar to a scientific paper and include the following sections: introduction (objective of the study), methods, results, and discussion. The report is intended to show that the specimens were used in a scientific method, and allows for the evaluation of potential cumulative effects from multiple projects in the same area. A report is required whether or not collecting activities were undertaken.

3) An instance of >10% unintended collecting mortality (per day, all species combined) requires sampling at a site to cease and the AMB to be contacted.

4) Minnow traps must be checked and emptied regularly enough to prevent significant holding mortality (Stipulation #4). Catch and mortality must be recorded each time traps are checked.

5) Each piece of unattended sampling gear must be: 1) labeled with the permittee’s name, telephone number, and permit number, 2) properly secured to ensure retrieval, 3) placed in a location where they will not be easily noticed (e.g., under cut banks, in pools away from roads or trails), 4) allowed to soak no more than twenty-four hours at a time, 5) located with GPS coordinates, and 6) accounted for and removed at the conclusion of sampling.

6) Salmon eggs used as bait in traps must either be sterilized commercial eggs or, if raw, disinfected prior to use. A 10-minute soak in 1:100 Betadine solution or some other disinfectant is adequate. Commercial eggs must be placed into a container that does not allow the fish to consume them (e.g., film canister with holes punched in it or a perforated plastic bag).

7) Gloves, boots, and collecting gear should be cleaned and disinfected between streams to reduce the potential of pathogen transmission and the spread of invasive species. Clean all equipment free of sediment, vegetation, and seeds, and then wash/line in 1:100 Betadine solution or soak in 1% bleach solution for a minimum of 10 minutes. Felt or absorbent socks on waders and wading boots are prohibited.

8) If anadromous fish are found in permitted streams, rivers, and lakes, the permit holder will submit a nomination form to the AMB (Stipulation #1) within 24 hours should you find any species suspected to be a non-native species during your sampling. If discovered, the organism should be killed, preserved by freezing or placing into 90% alcohol, and taken to the nearest ADF&G office. Please take a photo of the organism, as well as a photo of the organism in the environment in which it was observed, and note the location with a GPS or by describing it on a map with landmarks.

9) No fish may be possessed live or transported live under this permit.

10) Contact Tammy Davis with the ADF&G Invasive Species Program (907-465-6183 or 1-877-INVASIV), and the nearest AMB (Stipulation #1) within 24 hours should you find any species suspected to be a non-native species during your sampling. If discovered, the organism should be killed, preserved by freezing or placing into 90% alcohol, and taken to the nearest ADF&G office. Please take a photo of the organism, as well as a photo of the organism in the environment in which it was observed, and note the location with a GPS or by describing it on a map with landmarks.

14) Failure to comply with the conditions of this permit will result in the loss of future permitting privileges.

15) PERMIT VALIDATION requires permittee’s signature agreeing to abide by permit conditions before beginning collecting activities.

Signature of Permittee

cco: Heather Scannell, Division of Sport Fish, Fairbanks
Brandy Baker, Division of Sport Fish, Fairbanks
Bonnie Barba, Division of Commercial Fisheries, Fairbanks
Audra Braise, Habitat Section, Fairbanks
Michelle Morris, Commercial Fisheries Permit Coordinator, Juneau
Colonel Massie, Alaska Wildlife Troopers
Captain DeGroat, Alaska Wildlife Troopers Northern Detachment
## Appendix B: Sampling Sites

The sites used in 2021 were consistent with those used in 2020.

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## Appendix C: Beaver Report

Piledriver Slough Beaver Activity Survey Report 2021
Tanana Valley Watershed Association  
December 4 2021

Purpose:
The Piledriver slough mitigation plan monitors changes to the Piledriver slough that may be caused by beaver activity. Due to construction of the new rail extension, a levee was put in place that blocks flushing flows into the Piledriver Slough from the Tanana River. The flow rate changes may cause ice and log jams that would hinder fish passage. Beaver dams may no longer be knocked out by flushing spring flows and could cause further fish passage issues. Beavers are a natural part of the local environment and can help or hinder the other wildlife in the area. In the case of Piledriver Slough, monitoring will be conducted to evaluate the beaver dams and determine if they need to be removed to aid fish passage through the slough.

Methods
The ten-mile section of Piledriver from the levee site to the Bailey Bridge was monitored in two sections: “Upper Piledriver” from the levee site to the Old Valdez Trail road crossing and “Lower Piledriver” from the Old Valdez Trail road crossing to the Bailey Bridge adjacent to Eielson Airforce Base. Piledriver Slough was monitored on May 13-14, June 20-21 and August 26-27 2021. Identification of dam, and lodges were marked with GPS Locations. Pictures and videos were taken for further comparison and review. Beaver activity was classified as active or inactive and labeled as a primary dam, secondary dam and lodge, and lodge size, blockage and other changes were noted.

Report
In 2021 it appears that the beaver population is robust and healthy and has continued its expansion into this stretch of lower Piledriver Slough. There was no noted disruption to the dams or lodges due to flooding or flushing flows, which allowed the beavers to develop unimpeded and create a dam habitat which blocked the passage of adult salmon, grayling, and other species. The dams were observed in May and June as completely blocking passage at several locations whereas during August sampling there was water flowing over about half of the dams.

Our crew observed sign (chew sticks, vegetation removed, old dams re-constructed etc) during May, June and September sampling that indicate two major areas that are being used by beavers on the Lower Piledriver Slough. The first area of interest centers on the lodge and series of 2 dams just downstream of the Cady property. Here the two old dam sites (1 and 2) were fully constructed and blocking fish passage. An additional new dam at site 4 was also reconstructed and fully blocked passage for the first time this year. On Upper Piledriver slough site 3 a beaver dam was also constructed for the first time this year.

In 2019 we saw a new lodge at 64.62057, 147.09087. This lodge supports our theory that at least two separate beaver colonies made Piledriver Slough their home this year. The first colony is based out of the lodge at “Site 0” and supported by primary dam “Site 1” and Secondary dams at “Site 2” and also refurbished this year, “Site 3”. The second colony is based out of the new lodge “2019 lodge” and supported by a primary dam at “Site 4” and a new smaller secondary dams at “2019 Dam 1” (64.60448, 147.08798) and “2019 Dam 2” (64.60552, 147.08673). This change was confirmed in 2020 observations. In 2021 this trend continued but the second colony seemed to be much more active. It seems possible that beavers have shifted their focus from the first colony farther upstream.
Dams on Upper Piledriver Slough:

Upper Piledriver site 5:
64.59399  -147.08316

The dam at Site 5 on upper Piledriver Slough first observed during the September 2019 sampling period continued to be active in 2020 and expanded in 2021. This dam caused significant changes to the hydrology of the site, raising the water about 11 inches from normal levels and changing where we could set fish trap. Students from Salcha Elementary were eager to explore this major change to the site.

Upper Piledriver Site 3:
64.60045  -147.09204

Beginning in 2020, beavers have been actively maintaining a dam at site 3, the Hoover family’s yard (formerly Principal Annie Keep-Barnes’s yard). This dam has significantly raised the water level here and is a secondary dam.

Dams on Lower Piledriver Slough:

Site 0:
64.3916  147.0647
   6       3

This lodge was active in fall of 2020 but was not being used in 2021. We are not sure why this activity ceased but it is possibly due to human activity as this lodge is adjacent to the Cady’s private property.

Site 1:
64.3612  147.0507
   1       6

A primary dam was found downstream of the lodge at site 0 and Alan Cady’s property. This dam ranged from 2-3 feet tall and blocked all passage during May, June and September floats.
Site 1 from downstream looking upstream during May 2021 sampling, note algae.

Site 1 seen from upstream during June 2021 sampling, note build-up of algae.
Site 1 from downstream looking upstream during August sampling with higher water.

Site 2:

| 64.36158 | 147.05171 |

This is the secondary dam associated with site 1 dam. It was observed during all 3 floats and passage was blocked in May and June but breached in August. TVWA did not cause this breach, it appeared to be natural and associated with higher water levels. This dam seemed to be retaining a larger than usual amount of floating algae and debris in June sampling.
Site 2 seen from below during May sampling.

Site 2 seen from below during August sampling, note natural breach.

2019 Dam 1:
64.60448, 147.08798
This was a secondary dam located just downstream of Site 2. 2019 was the first year that we have observed a dam at this location and it continued to be inhabited in 2020. It indicates the spread of the beaver colony in this area. In 2021 this dam did not appear to be actively maintained.

*2019 Dam 1 looking downstream during August sampling, passage was not being blocked during the high water.*

**2019 Lodge:**
64.62057, 147.09087

This lodge was noted during September sampling this year. It was a new lodge in 2019, created in what had possibly been an old site and continued to be used in 2021. It was complete with a large feed pile, indicating that the beavers will likely be inhabiting it this winter. This stretch of the slough contains large quantities of beaver fecal content, which can be viewed from the water.
Site 3 (2019 Dam 2):
64.60552, 147.08673

There were a series of old dam sites indicated by remaining rocks and small debris throughout Piledriver Slough. In 2019 we observed some of these sites being reclaimed and added to. This is the case with 2019 Dam 2. It was first observed during June sampling when water was lower but was added to in September. The dam was not blocking passage completely during either sampling but will be monitored. This was not being used in 2021.
Dam 3 during May sampling.
Site 4:
64.3725  147.0512  
2       6
This dam is much farther down Piledriver slough than the first two. It is at a natural narrow spot in the slough and was observed blocking passage completely during all sampling periods.
Site 4 dam from below during May sampling, note the low water.
Conclusions:

Observable beaver activity in lower Piledriver Slough in 2021 decreased slightly from 2020 and increased in upper Piledriver Slough. For most of the summer, dams were significant enough to
block most fish passage, including spawning salmon. However, by late August, juvenile salmon and grayling were found in between the two major dam/lodge complexes at lower Piledriver site 4 (salmon) and 6 (grayling). Dams seemed to be tallest in May sampling (likely due to low water flow) and most of the secondary dams were partially breached during August sampling. TVWA did not take action to remove or alter these dams, per our previous instruction from ADFG and the Railroad.