CT Based Semi-Automated Method for Pneumonia Severity in Mice

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Abstract
Community-acquired pneumonia is an important clinical problem, with high rates of misdiagnosis and mortality. Diagnoses are typically conducted using two-dimensional chest x-rays, which have shown to be generally time-consuming and inaccurate, so current diagnostic methods should be improved. The goal of this research was to utilize Micro-Computed Tomography (MicroCT) and image analysis software to develop a diagnostic algorithm to quantitatively assess the severity of pneumonia in mice. This method provides immediate, more precise, and more accurate diagnoses as opposed to the qualitative assessments done by radiologists at present. MicroCT provides opportunities for non-invasive radiographic endpoints for pneumonia studies. A quantitative scoring of previously obtained Computed Tomography (CT) scans of pneumonia infected and control mice lungs was developed with a semi-automated image segmentation algorithm. At the endpoint of 168 hours, each of the mice was categorized as either a) a Saline (control)-injected mouse (total=13), a Pneumonia-injected Survivor (total=11), or a Pneumonia-injected Non-survivor (total=11). The scores demonstrated that the semi-automated algorithm was better able to distinguish certain pneumonia groups than could radiologists. The three comparison tests that were performed were Saline vs. All Pneumonia Injected Mice, Pneumonia Survivors vs. Pneumonia Non-survivors, and All Survivors (both Saline & Pneumonia) vs. Pneumonia Non-survivors. In all three comparisons, the semi-automated method did a better job discerning these groups than did the radiologists, with p-values of 0.001, 0.039, and 0.001 for the semi-automated algorithm, and 0.004, 0.581, 0.058 for the radiologists, respectively. The newly developed algorithm using CT scans and imaging techniques to assess early pneumonia in this pilot study shows promise and merits further testing.

Introduction
In the United States, pneumonia is the sixth leading cause of death and the number one infectious disease killer (Niederman et al., 1993). The disease is an inflammation of one or both lungs caused by an infection from bacteria, viruses or fungi. The infection causes the alveoli of the lungs to become inflamed and filled with fluid, which leads to symptoms such as cough, fever and respiratory breathing difficulties (Jelic, 2008). Often times, pneumonia occurs as a secondary infection when the immune system of a person is already weakened due to prior infection, such as an upper respiratory tract infection. This primary infection causes inflammation in the inner lining of airways that leaves the patient susceptible to the secondary infections such as pneumonia (Boone, 2004; Niederman et al., 1993).

Pneumonia can be classified by the population affected. Hospital-acquired pneumonia is acquired when a patient breathes germs during a hospital stay for another illness. People are most prone to hospital-acquired pneumonia while on a mechanical ventilator, since potentially pneumonia-causing bacteria and viruses may be blown directly into the lungs. The most common type of pneumonia is community-acquired.

Community-acquired pneumonia is acquired outside of hospitals and other health care settings, with about 5.6 million people getting infected every year in the USA and 1.1 million requiring hospitalization (Niederman, McCombs, 1998; Niederman et al., 1993). Community-acquired pneumonia is an important clinical problem, with high rates of misdiagnosis and mortality. Current methods to diagnose pneumonia rely on two dimensional (2D) chest X-rays, which are known to have low sensitivity early in the course of pneumonia (Mohd, 2010). Radiologists typically score 6 lung zones (upper, middle, and lower, on the right and left sides) for each mouse on a scale of 0 to 4, such that zero is normal, and the maximum possible abnormal score is 24; 0 represents 0% pneumonia involvement, 1 represents up to 25% involvement, 2 represents up to 50%, 3 represents up to 75%, and 4 represents up to 100% (Mosier, 2004). These chest X-rays may take days to diagnose the severity of pneumonia, in which time immunocompromised patients, such as patients with HIV/AIDS, cancer, diabetes, or sickle cell anemia, may reach a severity beyond curing (Smrga, 2008; Stuart, 2008). Because of their decreased ability to fight the bacteria, patients with chronic illnesses are more likely to require
immediate diagnosis and care for pneumonia. For example, immunocompromised patients with pneumonia have a mortality rate of 12% (Mohd, 2010). Thus, imaging techniques to evaluate pneumonia earlier and with more accuracy would be important diagnostic tools for clinicians. Imaging information can then be used to guide decisions on the clinical care needed, such as whether to hospitalize or to treat the patient at home, thus improving pneumonia diagnosis.

Furthermore, radiologists are often inconsistent with their diagnoses; two radiologists may judge the severity of pneumonia in patients very differently, leading to possible misdiagnosis (Hsu et al., 2007).

In order to address these limitations of inaccuracy, inconsistency, and delayed diagnosis, a different diagnostic method is required. Computed Tomography (CT) scans use X-rays that pass through the specimen and are received by sensors on the other end. Denser portions of the specimen result in a reduced amount of radiation received by the other end, since the specimen hinders the radiation. This disparity in densities, or attenuation, can be reconstructed to produce a 3D image with different grayscale values. Hounsfield Units (HU) are grayscale values that correspond to the density of each voxel. In the Hounsfield scale, -1000 represents air, 0 represents water, and 1000 represents bone density. Notably, fluid or pus in the alveolar sacs would be approximately 0 Hounsfield Units, normal lung alveoli have a mixture of air and tissue with near -500 HU, and voxels in lung with a mixture of air and fluid would be between -500 and 0 HU. CT scans, which can visualize the entire lung as opposed to the 2D projection scans in a chest radiograph, might have the sensitivity to assess the severity of pneumonia as early as 24 hours after onset. This earlier timeframe for treatment would allow immunocompromised patients to receive immediate treatment, thus decreasing their mortality rate. Since CT scans provide a more detailed depiction of the lung, they are potentially more accurate than the current chest X-ray method. Finally, by developing a semi-automated method that uses CT scans to diagnose the severity of pneumonia, more precise diagnoses can be conducted, since the procedure is more automated and less prone to human error (Muller, 2006).

The purpose of this particular research was to develop an algorithm to measure the severity of pneumonia in mice through Micro-Computer Tomography (MicroCT) Scan Analysis and test its effectiveness through comparison with radiologists’ diagnosis. MicroCT works in the same way as a regular CT scanner, but is typically used to image smaller specimens, such as rodents, as opposed to human beings. There were three goals for the image analysis algorithm. The first was to achieve high reproducibility in repeated analysis of the same MicroCT scan. Current methods typically have two radiologists independently score the chest X-rays and have the final score be the average of the independent scores. The second goal was to achieve higher accuracy using image segmentation algorithm to quantify the amount of pneumonia in the lungs. This is different from current methods which require radiologists to qualitatively assess multiple images of pneumonia. The quantification would be done by loading the CT scans in an imaging software, and determining the voxel distribution in order to compare densities. Finally, the project aimed to increase efficiency in diagnosis. A semi-automated computer algorithm would allow more measurements to be taken in a smaller amount of time than with current methods, without special expertise in radiology. The radiologist would have to have a couple of necessary skills. The first is the ability to use Amira, the software used in this paper. The second is knowledge of basic lung anatomy, such as the location of the trachea, stomach bubble, and mediastinum. Finally, the radiologist would need the ability to use a quantitative diagnosis performed by the computer to give the correct treatment to the patient. The automation is due to the computer’s automatic calculation of the severity of pneumonia, which, under current circumstances, would be done by the radiologist himself.

We hypothesized that in vivo MicroCT scans of mice with early bacterial pneumonia could be scored quantitatively by semi-automated imaging methods, with good reproducibility and correlation with the bacterial dose inoculated, pneumonia survival outcome, and radiologists’ scores previously obtained.

Materials and Methods

The project used MicroCT scans to evaluate a murine model of bacterial pneumonia through image analysis by semi-automated segmentation and comparison of results to radiologists’ interpretation. The following steps were used in the research to prepare mice with pneumonia, and then determine and validate the severity of their pneumonia:

1. Inoculation of mice with different severities of pneumonia bacteria and
acquisition of CT scans (Hsu et al., 2007).
2. Radiologists’ diagnosis of pneumonia (Hsu et al., 2007).
3. Pneumonia diagnosis using semi-automated diagnostic algorithm

The first two steps were done prior to this project, whereas the third step was conducted specifically in this research. Mercury Computer Systems’ Amira 4.1 software was used in this project to perform image analysis of the CT scans. Amira can perform image segmentation, 3D visualization, and other image processing (Schimel, 2007).

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Since the severity of pneumonia can be assessed by the volume distribution of materials within the lung due to inflammation, it was expected that more lung volume distribution in the range of high attenuation for greater severities of pneumonia would be observed (Hsu et al., 2007). After loading the stack of typically near 400 axial slices for 3D reconstruction in Amira, the lung was reconstructed by selecting Hounsfield units -510 to 0 (Muller, 2006). However, non-lung anatomical structures overlapping or in the vicinity of the lungs in the mice CT scans needed to be excluded from the lung reconstruction, because these structures are not involved with pneumonia infection (Figures 3a, 3b, and 4) (Iwaki, 2001). The non-lung components that overlapped with the lungs were primarily the stomach bubble, trachea, and mediastinum (Mosier, 2004).

The algorithm excludes each of these components systematically using the image manipulation features of Amira. Note that all subsequent non-3D images are axial cross-sections of the upper mouse body. The stomach bubble is a pocket of air that lies right below the lung.

Figure 2a: Axial Lung Slice – Manual Exclusion of Stomach Bubble
A projection view of Amira being used to eliminate the stomach bubble from the lung.

Figure 2b: 3D Reconstructed Lung – Manual Exclusion of Stomach Bubble
A 3D view of Amira selecting the stomach bubble, which will then be removed from the lung.

The contents in this bubble fall in the Hounsfield Unit range of -510 to 0 and also border the lung. Therefore, when a region in the lung within this range was clicked, the stomach bubble was included in the selection as well. The stomach bubble had to be manually removed, as shown in Figures 3a and 3b. Like the stomach bubble, the trachea is attached to the lung and gets selected in the -510 to 0 range. To remove it, the trachea was cut off at the carina, the point at which the two bronchi join to become the trachea. Then, the same procedure used in the stomach bubble removal was used for the trachea removal (Figure 3). The mediastinum is the anatomic region with a group of structures between the lungs that...
includes blood vessels and the esophagus. It was at times hard to distinguish between lung and mediastinum, making separation of mediastinum difficult in many scans. However, since the mediastinum accounts for such a small percentage of the lung volume, its effect on the voxel quantification was negligible, thus essentially eliminating human error.

The procedures for removing of stomach bubble and trachea were used to remove the mediastinum as well. At this point, the trachea, stomach bubble, and mediastinum had been removed from the lungs. Therefore, the image reconstructed contained only those voxels that could be affected by pneumonia. This image was now ready for segmentation (Figure 4).

The lung was then segmented into 8 regions using the following steps. The density ranges of each of the 8 materials, named Well-Aerated Lung, B, C, D, E, F, G, and H were created, with Hounsfield values of -510 to -350, -350 to -300, -300 to -250, -250 to -200, -200 to -150, -150 to -100, -100 to -50, and -50 to 0, respectively. These materials were entered into the “Label Voxel” tool, generating these 8 distinct density ranges within the entire CT scan. Figure 5 illustrates this procedure: The materials “Well-Aerated Lung,” “B,” and “C,” encapsulate all voxels inside the entire scan within the density ranges of -510 to -350, -350 to -300 and -300 to -250, respectively.

Though the tool can only analyze three materials at a time, because the analysis is almost instantaneous, it had a negligible effect on the time it took to do the quantification analysis. However, we were only interested in the volume of each material within the lung, not the entire scan, which includes bone, muscle, fat, fur, and other tissues. The units of the volume measurements are irrelevant, since we eventually determined the percentage distribution of each material within the lung. Amira allowed us to measure how much of each of the 8 materials was present in the lung volume. In Figure 6, the volume of each of the first 3 materials can be seen: outside the lung (Exterior), inside the lung (Lung), and in the whole scan (Total).
Essentially, the semi-automated method was broken down into the three following steps. The first was the isolation of the lung from the rest of the CT scan. The second was the removal of extraneous anatomical features, such as the stomach bubble, trachea, and mediastinum, which cannot get affected by pneumonia and would therefore skew our calculations. The final step involved finding the percentage of each of the lung materials (Well-Aerated, B, C…). The voxel distribution was analyzed for each of the survival groups.

In the animal facility prior to this project, 24 mice inoculated with bacteria developed symptoms of pneumonia, 11 of which died by the endpoint of 168 hours. The 11 mice inoculated with saline had few or no signs of pneumonia, and none died. No mice died during the actual MicroCT scan. After monitoring the mice for 7 days, each of these 35 mice was assigned to one of three experimental groups based on survival outcome and inoculation: (1) Pneumonia Survivor (total = 13), (2) Pneumonia Non-Survivor (total = 11), and (3) Saline-Inoculated Control (total = 11).

Results
In the development of a semi-automated method, three trials were conducted for each of the 35 mice, and the coefficient of variation (CV) was calculated to evaluate reproducibility. The semi-automated segmentation was reproducible, with the trials for each MicroCT scan resulting in the same segmentation volumes within a coefficient of variation of 2%. Grouping the MicroCT segmentation results showed the expected findings for the three experimental groups (Figure 7).

In the lower ranges of attenuation, Saline-Inoculated mice had the greatest percentage of lung volume, followed by Pneumonia Survivors, then Pneumonia Non-survivors. For example, the material with the lowest attenuation, “Well-Aerated Lung” (Hounsfield units -510 to -350), on average, made up 18.3% of lung volume in the Saline-Inoculated group, 10.9% of Pneumonia Survivors, and 8.2% of Pneumonia Non-surrors. On the other side of the density spectrum, the mice that eventually died of pneumonia had the highest percentages of lung volume in the higher ranges of attenuation, followed by Pneumonia Survivors, and finally the Saline-Inoculated group.

Figure 8 presents the p-value distribution from a t-test for each of the 8 density ranges, for the following three experimental group comparisons: (1) Saline-Inoculated vs. All Pneumonia Infected, (2) Pneumonia Survivors vs. Pneumonia Non-Survivors, and (3) All Survivors vs. Pneumonia Non-Survivors.

Comparisons (1) and (3) were significantly different (p-value < 0.05) for all the materials except for material D (Figure 8, highlighted yellow).

This may indicate the density range of material D (-250 to -200) includes regions of the lung that are not affected by pneumonia, since the three experimental groups did not show significant difference in their average percentages for this material. Excluding material D, the Pneumonia Survivors vs. Pneumonia Non-Survivors comparison, however, showed no statistical significance except for in material H (Figure 8, highlighted orange). Material H, as can be seen by Figure 9, showed far greater significance by the
A qualitative assessment of the scans can also be made. Figures 10-12 show frontal pictures of three mice, each from the saline, pneumonia survivors, and pneumonia non-survivors groups, respectively. The red color represents voxels in the highest third density range of the lung, the orange color represents voxels in the middle third density range, and the blue color represents the voxels in the lower third of the density range.

Figure 10: Example of a saline mouse lung; red represents high attenuation density; orange represents low attenuation. Frontal view of a saline mouse’s lung. Blue represents the lower third of the lung density range, orange represents the middle third, and red represents the upper third. Notice that most the lung is shaded orange or blue, indicating a relatively less dense lung.

As it can be seen, the saline mouse seems to have the greatest amount of orange and least amount of red, the pneumonia survivor group has less orange and more red, and finally, the pneumonia non-survivor group has the least orange and most red.

Figure 11: Example of a pneumonia survivor mouse lung; red represents high attenuation density; orange represents low attenuation. Frontal view of a pneumonia survivor mouse’s lung. The lung has far more red shade than the saline mouse, indicating its relative high density.

Figure 12: Example of a pneumonia non-survivor lung; red represents high attenuation density; orange represents low attenuation. Frontal view of a pneumonia non-survivor mouse’s lung. The lung is almost completely shaded red, showing that the mouse has a severely pneumonia-affected lung.

This intuitively makes sense because we expect the mice which show greater signs of pneumonia to have greater amounts of red than any other color, and vice-versa.

Discussion
Preliminary statistical analysis of the semi-automated segmentation of MicroCT detected differences between the groups of pneumonia survivors vs. pneumonia deaths. There was good reproducibility of the semi-automated segmentation, with less than 2% variability with repeated application of the methods. Radiologists’ average scores provided statistically significant differences between the mice inoculated with...
pneumonia vs. the mice inoculated with saline, but not between the groups of pneumonia survivors vs. pneumonia non-survivors, nor between all surviving mice vs. pneumonia deaths. This pilot comparison suggests that the semi-automated segmentation may provide a better method for quantitative scoring of pneumonia severity by CT scans, compared to scoring by radiologists. Only a few other studies have applied quantitative image analysis to CT scans of pneumonia in animal models (Amigoni, 2008; Armbrust, Mosier, 2005). For example, one examined another type of bacterial pneumonia and used different methods to score the pneumonia. Another examined acid-aspiration pneumonia to score the lung injury. Their studies did not examine severity in terms of survival, and did not compare the image analysis to scoring by radiologists.

Previous applications of quantitative image analysis of lung CT scans have focused on different clinical problems, rarely pneumonia: (1) emphysema, in which lungs have abnormal pockets of low attenuation, (2) lung tumors, which have much higher attenuation than normal lungs, and (3) normal and abnormal physiologic distribution of aeration and blood flow, often using tracer materials to detect flow (Ritman, 2005). Tracers to detect blood flow include xenon, intravenous iodinated contrast, and microspheres, but since such tracers were not used in this project, the problem of distinguishing normal lung vs. high-attenuation pneumonia involvement was more difficult.

Clinical use of CT imaging is widespread throughout the world because of the wealth of information that CT scans provide about abnormal fluid, tumors, aeration, etc. Clinical High Resolution CT (HRCT) is now starting to be used to diagnose pneumonia in patients, but is rarely used for community-acquired pneumonia, which affects more people than any other type of pneumonia (Jelic, 2008). Although the spatial resolution of MicroCT is better than clinical HRCT on an absolute scale (60 microns vs. 500-1000 microns), the spatial detail is better with HRCT when considering the anatomic size of mouse lungs vs. human lungs (apex to diaphragm ~3cm vs. ~50cm, alveoli 80 microns vs. 210 microns, respectively). Thus, it can be concluded that transferring the technique from mouse to humans should make the algorithm more accurate due to increased relative resolution. The current HRCT technique, though an improvement to the more common 2D chest X-ray method, may still provide insufficient detail for quantitative analysis of a 3-dimensional reconstruction because it generally captures only a few 2D slices (Uchiyama, 2003). Newer techniques of human spiral CT will capture enough information for the 3D reconstruction as used in our algorithm.

The principal limitation of this method was the accuracy of manually isolating the lung "volume of interest" from the rest of the mouse. It was sometimes hard to separate the lung from nearby structures, such as the mediastinum. In the future, iodinated or other contrast materials may help more finely define these anatomical structures, until the algorithm is applied to HRCT, which provides finer resolution and, therefore, clearer anatomical structure contrast. This way, it would be easier to isolate the lung during the steps using Amira delineated in 5.3.1 to 5.3.4. Another limitation of this animal study is the small sample size; we intend to extend this technique to another group of mice with pneumonia. Finally, these preliminary studies examined only one strain of bacteria and mice, with the inclusion of antibiotics and supportive care. Extending these techniques to other mice will require additional validation, but may help to provide a non-invasive endpoint for studies with experimental pneumonia in transgenic animals. This method of quantitative assessment of pneumonia severity by CT has potential for application in clinical trials in community-acquired pneumonia, as well as other lung diseases.

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