Better Detection and Treatment for Mold Exposure: Improving Outcomes for Patients by Matt Pratt-Hyatt, PhD

One of the hallmarks of integrative medicine is to focus on determining the underlying causes of chronic illnesses for patients, not just to treat symptoms. This has led many practitioners of integrative medicine to use testing that helps provide more personalized care protocols to their patients. It is the duty of diagnostic laboratories to deliver the most accurate test results possible and provide interpretations of the results to help practitioners understand the correlations between results and their patients' conditions. At The Great Plains Laboratory, we have been working on finding new ways to detect underlying causes of many illnesses and studying correlations between the results and conditions. Using results from both our MycoTOX Profile as well as our Organic Acids Test (OAT) (both urine tests), we have discovered correlations between them and uncovered new routes of treatment for patients with mold exposure.

For several decades, new research has gone into the detection and treatment of mold mycotoxins. These are the toxic metabolites produced by certain types of fungi. These small molecules are often carried on dust particles or food or are present in water-damaged buildings and homes. The most common routes of exposure are inhalation, skin contact, or ingestion through contaminated food.¹ Exposure can lead to different types of chronic health conditions depending on the age, sex, genetics, and health status of the patient. Common symptoms of mycotoxin exposure are fatigue, headaches, rashes, food sensitivities, joint pain, and cough.²⁻⁴ Mycotoxins can induce disease through different routes such as cytotoxicity, immunosuppression, and DNA damage.

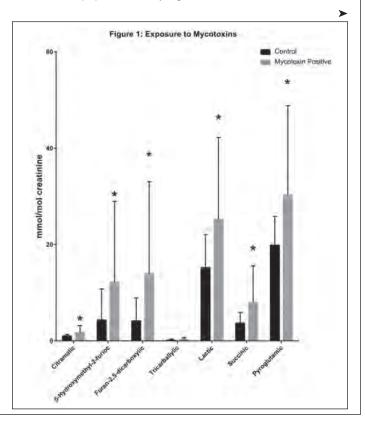
Mycotoxins can cause cytotoxicity by inhibiting multiple pathways within the cell. Mycotoxins inhibit the production of proteins by interfering with the function of the ribosome. Multiple mycotoxins bind to different subunits that are involved in protein biosynthesis, such as the 60S subunit of the ribosome.^{5,6} In addition to inhibition translation, mycotoxins can also activate Jun N-terminal Kinase (JNK), mitogen-activated protein kinases (MAPKs), and p38.^{7,8} Activation of these stress response elements in the cell can lead to cellular damage.

Through their mycotoxin metabolites, pathogenic fungi can reduce the efficacy of the immune system by suppressing multiple different components of it. Mycotoxin metabolites can suppress T and B lymphocyte activity, inhibit immunoglobulin production, and reduce antibody production. Some species of fungi release mycotoxins which can interfere with which genes are activated. Some also release polysaccharides which can result in the induction of neutrophil apoptosis. All these processes result in the body's inability to fight off the fungi as well as other infections.

DNA damage caused by mycotoxins has been shown to be carcinogenic.¹² There are two different pathways in which mycotoxins can damage DNA. First is the ability for some mycotoxins to covalently bind to DNA nucleotides. Mycotoxins can covalently bind to the N-7 position of guanine and the N-3 position of adenine, both of which can interfere with DNA synthesis. These adducts can cause an incorrect substitution of a nucleotide or cause deletion in the DNA code.¹³ The second cause of DNA damage from mycotoxins is the inhibition of DNA topoisomerase I and II. These enzymes are required to untangle DNA during replication, and preventing this activity can lead to the accumulation of DNA breaks.¹⁴

Types of Mold and Mycotoxins

Mycotoxins are produced by filamentous fungi, which are ubiquitous because of their ability to thrive in many different types of environments.¹⁵ The most common mycotoxins are produced from the fungi genera of *Aspergillus, Fusarium, Penicillium,* and



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Stachybotrys. Many of fungi species can produce more than one type of mycotoxin (Table 1). In this table, we list ten of the most common mycotoxins, and we can depict which mold species produce these mycotoxins. *Aspergillus* is the most prevalent mold genus in the environment. The two most common *Aspergillus* mycotoxins are aflatoxin and ochratoxin, and their main target is the liver.^{16,17} *Aspergillus* species are commonly associated with indoor air problems.¹⁸ The most common route of transmission for *Aspergillus* is inhalation.¹⁹

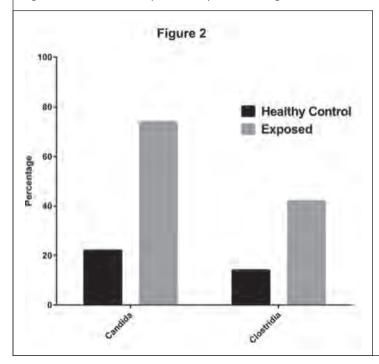
Fusarium fungi grow best in temperate climate conditions.²⁰ *Fusarium* is present in both indoor as well as outdoor environments.²¹ It also grows worldwide on many different types of grains including corn and wheat.²² Most trichothecene mycotoxins are produced from *Fusarium* species.

Penicillium is a blue-green mold found on fruits, vegetables, and indoor environments. Many different types of citrus fruits can become contaminated with *Penicillium*, but it can also contaminate seeds and grains. Factors leading to a contamination of *Penicillium* in the home, work, or school environment include inadequate heating and ventilation, water leaks, and low sunlight.²³

Stachybotrys is a greenish-black mold. This mold can grow on materials with high cellulose and low nitrogen content such as gypsum board, paper, fiberboard, and ceiling tiles. The humidity requirements for *Stachybotrys* are much higher than other fungi, around 93%, whereas other fungi grow at approximately 75%, and it often occurs in the presence of other fungi.^{3,24} Toxicity caused by *Stachybotrys* is mostly caused by the toxins and other compounds produced, and less from particle penetration from the spores.

Detection of Mold Exposure

For decades, it has been difficult to determine if patients have been exposed to mold toxins. Research and testing in the agricultural market has proven helpful in testing and treatment



methods for humans. One method of detection, which was developed a decade ago, is Enzyme Linked Immunosorbent Assay (ELISA), which has been used to detect mycotoxins in many different types of crops. There are two barriers in transferring this technology to human samples. These barriers are insufficient sensitivity and matrix effect (ME) in human samples. The first barrier is caused by the small amounts of mycotoxins in human blood and urine, which are both much less than seen in crops. The FDA action level for aflatoxin is 20 ppb (20 μ g/L), which is a factor 100 times higher than seen in human samples.^{25,26} Sample ranges for many mycotoxins in humans are normally in the 60-2000 ng/L (.06-2 μ g/L) range.²⁶

The matrix effect barrier is caused by compounds in the sample matrix that are not related to the analyte being measured but can cause a result to be either higher or lower than the actual amount. These MEs can influence any type of test, and protocols must be performed in order to separate the analyte of interest from these interferences.²⁷ Studies in the past eight years have focused on a new technology to access mycotoxin exposure, which is liquid chromatography mass spectrometry (LC-MS/MS).

The introduction of LC-MS/MS has allowed scientists and practitioners to quantitatively measure many different chemicals. These measurements can be done on compounds either individually or simultaneously if the compounds have similar molecular characteristics. A mass spectrometer is a device that measures the molecular mass of compounds. Many different fields have used this technology and two of the most common are drug testing and chemical exposure testing, which both need the sensitivity and selectiveness that LC-MS/MS provides.²⁸ Over the last 10 years in the scientific community, LC-MS/MS has become the gold standard for the measurement of mycotoxins.²⁹

At The Great Plains Laboratory, we use two methods of detection to determine the best course of treatment for patients. To detect mycotoxins, we utilize LC-MS/MS technology because of its sensitivity as well as its selectivity in determining analytes. Our laboratory experts have developed a method for extracting mycotoxins out of human matrixes that provides the most sensitive and accurate data available. We recommend that all patients who take our LC-MS/MS test for mycotoxins, also take an Organic Acid Test (OAT), and both tests can be run on the same urine sample. The Organic Acids Test is a metabolic test performed on the gas chromatography mass spectrometer (GC/ MS). The OAT provides a snapshot of how the body is performing metabolically and it offers four distinct areas of information that can assist in the treatment of mycotoxin exposure. These areas are fungal markers, glutathione markers, mitochondrial markers, and yeast/clostridia markers.

To better understand how the OAT and mycotoxin test overlap, we ran a study of 100 healthy patients and 150 mycotoxinexposed patients. These results are depicted in Figure 1. All of these values were statistically significant with p-values below 0.05. The first insights that we observed in patients that took both the mycotoxin test and the OAT was that two markers in the yeast/fungal section of the OAT were statistically elevated over healthy controls. These two markers were 5-hydroxymethyl-2-furioc and Furan-2,5-dicarboxylic, which are both produced by species of *Aspergillus* (Figure 1).^{30,31} Elevations in these two markers could indicate that the patient has a colonization of *Aspergillus* in the body. *Aspergillus* is the most common fungi to colonize the body. Because Aspergillus spores are smaller than most other species, 2 μ m to 10 μ m, they can reach the lower airways. This can lead to fungal colonization, which results in prolonged mycotoxin exposure and allergic responses. The most common areas of colonization are the sinus cavities, the lungs, and the gut.³² Our data indicates that 90% of patients with elevations in these two markers also have elevations of mycotoxins. These data have been shown to be extremely beneficial in determining the best treatment of patients.

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The second benefit for using the OAT in combination with the mycotoxin test is to measure detoxification capacity. In Figure 1, we demonstrate that pyroglutamic acid is elevated in mold-exposed patients, which indicates these patients have a higher usage of glutathione (GSH). Detoxification of many mycotoxins is

	The 40 Species of Wold and 10 Wycotoxins They Produce									
	Aflatoxin	Gliotoxin	Ochratoxin	Sterigmatocystin	Zearalenone	Roridin E	Verrucarin A	Enniatin B	Mycophenolic Acid	Citrinin
Species										
Acremonium sp.				present						present
Alternaria		present								
A. flavipes				present						present
Aspergillus flavus	present									present
A. fumigatus		present								present
A. niger			present							present
A. ochraceus			present	present						present
A. parasiticus	present									present
A. sydowii				present						present
A. versicolor				present						present
A. viridictum			present	present						present
Aureobasidium				present						
Chaetomium				present						
Cladosporium				present						
Cunninghamella				present						
Cylindrocarpon						present				
Dendrodochium						present	present			
Exophiala				present						present
Fusarium avenaceum				present	present			present		present
F. cerealis				present	present					present
F. clumonrum				present	present					present
F. equiseti				present	present					present
F. graminearum				present	present					present
F. incarnatum				present	present					present
F. moniliforme				present	present			present		present
F. solani								present		
F. verticillioides				present	present					present
Myrothecium roridum						present				
M. verrucaria						present	present			
Penicillium carbonarius		present	present	present					present	present
P. nordicum		present	present	present					present	present
P. stoloniferum		present	present	present					present	present
P. verrucosum		present	present	present					present	present
Phoma sp.				present						present
Rhodotorula				present						present
Scopulariopsis				present						present
Stachybotrys				present		present	present			present
S. chartarum				present						
Trichoderma viride		present		present						
Ulocaldium										
Verticillium				present						present
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The 40 Species of Mold and 10 Mycotoxins They Produce

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dependent on the endogenous molecule GSH. GSH is a tripeptide, which is made of glycine, cysteine, and glutamate. Mycotoxin toxicity is intensified in patients with glutathione transferase mutations (GSTP1 and GSTM).³³ Glutathione status is measured in the OAT with pyroglutamic acid and 2-hydroxybutyric acid. Patients that are exposed to mycotoxins can have elevated

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amounts of both compounds (Figure 1). High pyroglutamic acid occurs when the body has been depleted of glutathione, which then activates the γ -glutamyl cysteine synthetase, which leads to the production of pyroglutamic acid.³⁴ 2-Hydroxybutyric has also been linked to glutathione production. Patients with higher amounts could be glutathione deficient.³⁵ Using this information will allow practitioners to determine what dose of glutathione would be most appropriate for a patient.

The third benefit of using the OAT for patients is assessing how the mold mycotoxins are affecting the mitochondrial pathways in the patient's body. Many different types of mycotoxins can bind with ribosomes in the cell as well as ribosomes in the mitochondria. These interactions can inhibit proper cellular functions.⁶ The OAT assesses multiple different mitochondrial pathways. Studies done by The Great Plains Laboratory have demonstrated that patients who have been exposed to mold toxins accumulate lactic acid and succinic acid in their urine (Figure 1).

The fourth benefit of using the OAT for patients with possible mycotoxin exposure is the yeast and bacteria markers. Exposure to mycotoxins can lead to the inhibition of T helper cell function and inhibition of antibody synthesis.³⁶ Crucial immune organs have also been shown to decrease in size from exposure, such as the thymus, bursa of Fabricius, the spleen, and lymph nodes.³⁷ This can lead to the overgrowth of intestinal pathogens such as Candida or Clostridia.³⁸ Our data dictates that patients with high levels of mycotoxins have high levels of yeast 75% of the time and Clostridia 40% of the time (Figure 2). In this figure, we show that elevations in tartaric, arabinose, or carboxycitric acid are more common when the patient has been exposed to mycotoxins. These are serious co-morbidities that can lead to significant health issues. Diagnosis of these issues can lead to improved outcomes.

Treatment of Mycotoxins

Treatment of mold and mycotoxins must take into account possible routes of exposure; either internal colonization of mold or external exposure. Mycotoxin and fungi exposure can lead to many different symptoms including fatigue, headaches, difficulty breathing, rashes, gastrointestinal issues, joint pain, decreased libido, and more. As previously mentioned, these mycotoxins can come from water-damaged buildings or from food contamination. The most important element for treatment is avoidance of further exposure to water-damaged environments as well as contaminated foods. Using a licensed company to inspect and remediate the area is recommended. Remediation of contaminated areas is difficult because heat, ultraviolet (UV) light, bleach, ammonia, and ozone do not completely remove mold from contaminated areas.³⁹ Air spore counts are frequently done; however they are usually done over a short period of time and often result in false negative results. Another problem is that killing the mold does not eliminate the risk, because mycotoxins will remain in the environment.

Patients exposed to mycotoxins from mold may have two different conditions to treat. The first is the mycotoxin exposure, which can cause problems in many different tissues mentioned previously, and second is a possible infection and colonization of the fungi. To detoxify the mycotoxins, it is recommended to use a combination of glutathione, sauna, sequestering/binding agents, and antioxidants. Patients may also need to treat fungi infections and colonization. Glutathione (GSH) can be introduced in several ways, including treating with GSH precursors such as N-acetyl cysteine and whey protein or with an oral liposomal, transdermal, intravenous, nebulized, or intranasal form of GSH. It is recommended to use GSH with sequestering agents, because treatment with GSH has been shown to increase the mobilization of mycotoxins in the bloodstream.⁴⁰ Sauna therapy has been utilized for centuries for the removal of toxins from the body. Some of the most commonly used saunas are dry heat radiant saunas and far infrared saunas. Both have been shown to be equally good at removing toxins from the body. Infrared saunas have the advantage of inducing sweating at lower body temperatures. Sweating is important because many mycotoxins are lipophilic and can bind to lipid proteins, which will allow the mycotoxins to persist in the body for extend periods of time.⁴¹

Patients who have high levels of mycotoxins in their bodies may also be suffering from fungal infections and colonization. This is usually caused by direct exposure to elevated levels of mold spores. Fungal infections can occur in many different parts of the body. The most common areas of infection are the sinus cavities, lungs, and intestine. One Mayo Clinic study found fungal growth in 96% of patients with chronic sinusitis. The types of mold found included *Aspergillus, Penicillium, Alternaria, Cladosporium, Fusarium,* and *Cryptococcus*. All of these molds are known to produce mycotoxins and cause significant diseases.^{42,43} One study demonstrated that antifungal drug therapies were useful in combating microbial pathogens detected by gas chromatographymass spectrometry.⁴⁴

Antifungal medications have been shown to be efficacious against mycotoxins. Multiple studies have utilized amphotericin B for the treatment of fungal infections.⁴⁵⁻⁴⁷ Other antifungals that have been used include ketoconazole, fluconazole, and itraconazole. For intestinal fungal infections, nystatin or liposomal nystatin may be a good alternative.^{48,49}

Application of Antifungal Treatment in Autism and Parkinson's Disease

Our recent work with mycotoxins and antifungal treatment has focused on two different patient populations; those with autism and those with Parkinson's disease. Our studies have found numerous patients that exhibit symptoms of either autism or Parkinson's who also exhibit colonization with mold. Results indicate that treatment with antifungal therapies has greatly improved the symptoms of patients in these categories, which strongly suggests that mold exposure may play a role in exacerbating both autism and other neurological disorders.

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Mycotoxins are metabolites produced by fungi like mold, which can infest buildings, vehicles, and food. Diseases and symptoms linked to mycotoxin exposure include pneumonia-like symptoms, heart disease, rheumatic disease, asthma, sinusitis, cancer, memory loss, vision loss, chronic fatigue, skin rashes, depression, and liver damage.

The MycoTOX Profile now screens for 10 different mycotoxins, all in one urine sample. It uses the power of advanced mass spectrometry (MS/MS), which is necessary to detect lower levels of these fungal toxins.



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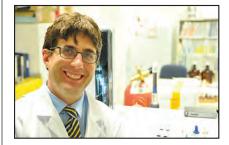
Conclusions

Exposure to mold and mycotoxins can result in many different types of illnesses depending on the patient's age, sex, genetics, and environment. Determining the root causes of these illnesses is our goal at The Great Plains Laboratory. Many in the health community in the last decade have come to realize the role that mold and mycotoxins can play in the health of our patients. We believe that using the Organic Acid Test along with the LC/MS mycotoxin test can produce the information necessary to provide optimum personalized care to patients with mold exposure.

References

- 1. Bennett JW, Klich M. Mycotoxins. Clin Microbiol Rev. 2003;16(3):497-516.
- Auger PL, Gourdeau P, Miller JD. Clinical experience with patients suffering from a chronic fatigue-like syndrome and repeated upper respiratory infections in relation to airborne molds. *Am J Ind Med.* 1994;25(1):41-42.
- Hodgson MJ, Morey P, Leung WY, et al. Building-associated pulmonary disease from exposure to Stachybotrys chartarum and Aspergillus versicolor. J Occup Environ Med. 1998;40(3):241-249.
- Howard DH. Pathogenic fungi in humans and animals. 2nd ed. New York: Marcel Dekker; 2003.
- Hassan YI, Watts C, Li XZ, Zhou T. A novel Peptide-binding motifs inference approach to understand deoxynivalenol molecular toxicity. *Toxins (Basel)*. 2015;7(6):1989-2005.
- Pace JG, Watts MR, Canterbury WJ. T-2 mycotoxin inhibits mitochondrial protein synthesis. *Toxicon.* 1988;26(1):77-85.
- Chung YJ, Zhou HR, Pestka JJ. Transcriptional and posttranscriptional roles for p38 mitogenactivated protein kinase in upregulation of TNF-alpha expression by deoxynivalenol (vomitoxin). *Toxicol Appl Pharmacol.* 2003;193(2):188-201.
- Zhou HR, Islam Z, Pestka JJ. Rapid, sequential activation of mitogen-activated protein kinases and transcription factors precedes proinflammatory cytokine mRNA expression in spleens of mice exposed to the trichothecene vomitoxin. *Toxicol Sci.* 2003;72(1):130-142.
- Fontaine T, Delangle A, Simenel C, et al. Galactosaminogalactan, a new immunosuppressive polysaccharide of Aspergillus fumigatus. *PLoS Pathog.* 2011;7(11):e1002372.
- Pahl HL, Krauss B, Schulze-Osthoff K, et al. The immunosuppressive fungal metabolite gliotoxin specifically inhibits transcription factor NF-kappaB. J Exp Med. 1996;183(4):1829-1840.
- Corrier DE. Mycotoxicosis: mechanisms of immunosuppression. Vet Immunol Immunopathol. 1991;30(1):73-87.
- Huynh VL, Gerdes RG, Lloyd AB. Synthesis and degradation of aflatoxins by Aspergillus parasiticus. II. Comparative toxicity and mutagenicity of aflatoxin B1 and its autolytic breakdown products. *Aust J Biol Sci.* 1984;37(3):123-129.
- 13. Cavalieri EL, Li KM, Balu N, et al. Catechol ortho-quinones: the electrophilic compounds that form depurinating DNA adducts and could initiate cancer and other diseases. *Carcinogenesis.* 2002;23(6):1071-1077.
- 14. Fehr M, Pahlke G, Fritz J, et al. Alternariol acts as a topoisomerase poison, preferentially affecting the llalpha isoform. *Mol Nutr Food Res.* 2009;53(4):441-451.
- More TT, Yan S, Tyagi RD, Surampalli RY. Potential use of filamentous fungi for wastewater sludge treatment. *Bioresour Technol.* 2010;101(20):7691-7700.
- Liu Y, Wu F. Global burden of aflatoxin-induced hepatocellular carcinoma: a risk assessment. Environ Health Perspect. 2010;118(6):818-824.
- Tao Y, Xie S, Xu F, et al. Ochratoxin A: Toxicity, oxidative stress and metabolism. Food Chem Toxicol. 2018;112:320-331.
- Nielsen KF, Gravesen S, Nielsen PA, Andersen B, Thrane U, Frisvad JC. Production of mycotoxins on artificially and naturally infested building materials. *Mycopathologia*. 1999;145(1):43-56.
- 19. Vonberg RP, Gastmeier P. Nosocomial aspergillosis in outbreak settings. J Hosp Infect. 2006;63(3):246-254.
- Doczi I, Gyetvai T, Kredics L, Nagy E. Involvement of Fusarium spp. in fungal keratitis. Clin Microbiol Infect. 2004;10(9):773-776.
- Jarvis BB, Lee YW, Comezoglu SN, Yatawara CS. Trichothecenes produced by Stachybotrys atra from Eastern Europe. Appl Environ Microbiol. 1986;51(5):915-918.

- Escobar J, Loran S, Gimenez I, et al. Occurrence and exposure assessment of Fusarium mycotoxins in maize germ, refined corn oil and margarine. *Food Chem Toxicol.* 2013;62:514-520.
- Sharpe R, Thornton CR, Osborne NJ. Modifiable factors governing indoor fungal diversity and risk of asthma. *Clin Exp Allergy*. 2014;44(5):631-641.
- 24. Grant G, Hunter CA, Flannigan B, Bravery AF. The moisture requirements of moulds isolated from domestic buildings. *Int Biodeterior*. 1989;25:259-284.
- Mitchell NJ, Bowers E, Hurburgh C, Wu F. Potential economic losses to the US corn industry from aflatoxin contamination. Food Addit Contam Part A Chem Anal Control Expo Risk Assess. 2016;33(3):540-550.
- 26. Solfrizzo M, Gambacorta L, Visconti A. Assessment of multi-mycotoxin exposure in southern Italy by urinary multi-biomarker determination. *Toxins (Basel)*. 2014;6(2):523-538.
- Stahnke H, Kittlaus S, Kempe G, Alder L. Reduction of matrix effects in liquid chromatographyelectrospray ionization-mass spectrometry by dilution of the sample extracts: how much dilution is needed? *Anal Chem.* 2012;84(3):1474-1482.
- JSC T, GL L. Drug-Testing technologies and applications. In: RC W, HY T, eds. Drug-testing technologies and applications. Totowa: Humana Press; 2005:29-69.
- Escriva L, Font G, Manyes L, Berrada H. Studies on the Presence of Mycotoxins in Biological Samples: An Overview. *Toxins (Basel)*. 2017;9(8).
- Kimura Y, Tani S, Hayashi A, et al. Nematicidal activity of 5-hydroxymethyl-2-furoic acid against plant-parasitic nematodes. *Z Naturforsch C*. 2007;62(3-4):234-238.
- Karich A, Kleeberg SB, Ullrich R, Hofrichter M. Enzymatic Preparation of 2,5-Furandicarboxylic Acid (FDCA)-A Substitute of Terephthalic Acid-By the Joined Action of Three Fungal Enzymes. *Microorganisms*. 2018;6(1).
- Horner WE, Helbling A, Salvaggio JE, Lehrer SB. Fungal allergens. Clin Microbiol Rev. 1995;8(2):161-179.
- Sun CA, Wang LY, Chen CJ, et al. Genetic polymorphisms of glutathione S-transferases M1 and T1 associated with susceptibility to aflatoxin-related hepatocarcinogenesis among chronic hepatitis B carriers: a nested case-control study in Taiwan. *Carcinogenesis*. 2001;22(8):1289-1294.
- Emmett M. Acetaminophen toxicity and 5-oxoproline (pyroglutamic acid): a tale of two cycles, one an ATP-depleting futile cycle and the other a useful cycle. *Clin J Am Soc Nephrol.* 2014;9(1):191-200.
- Darmaun D, Smith SD, Sweeten S, Hartman BK, Welch S, Mauras N. Poorly controlled type 1 diabetes is associated with altered glutathione homeostasis in adolescents: apparent resistance to N-acetylcysteine supplementation. *Pediatr Diabetes*. 2008;9(6):577-582.
- Lea T, Steien K, Stormer FC. Mechanism of ochratoxin A-induced immunosuppression. Mycopathologia. 1989;107(2-3):153-159.
- Boorman GA, Hong HL, Dieter MP, et al. Myelotoxicity and macrophage alteration in mice exposed to ochratoxin A. *Toxicol Appl Pharmacol*. 1984;72(2):304-312.
- Park SH, Kim D, Kim J, Moon Y. Effects of Mycotoxins on mucosal microbial infection and related pathogenesis. *Toxins (Basel)*. 2015;7(11):4484-4502.
- Peitzch, Bloom, Haase, Must, Larson. Remediation of mould damaged building materialsefficiency of a broad spectrum of treatments. *Journal of Environmental Monitoring*. Vol 142012:908-915.
- Hope J. A review of the mechanism of injury and treatment approaches for illness resulting from exposure to water-damaged buildings, mold, and mycotoxins. *ScientificWorldJournal*. 2013;2013:767482.
- Wild CP, Gong YY. Mycotoxins and human disease: a largely ignored global health issue. Carcinogenesis. 2010;31(1):71-82.
- 42. Praneenararat S. Fungal infection of the colon. Clin Exp Gastroenterol. 2014;7:415-426.
- Ponikau JU, Sherris DA, Kern EB, et al. The diagnosis and incidence of allergic fungal sinusitis. Mayo Clin Proc. 1999;74(9):877-884.
- 44. Shaw W, Kassen E, Chaves E. Assesssment of antifungal drug therapy in autism by measurement of suspected microbial metabolites in urine with gas chromatography-mass spectrometry. *Clinical Practice of Alternative Medicine*. 2000;1:15-20.
- Ponikau JU, Sherris DA, Weaver A, Kita H. Treatment of chronic rhinosinusitis with intranasal amphotericin B: a randomized, placebo-controlled, double-blind pilot trial. J Allergy Clin Immunol. 2005;115(1):125-131.
- Liang KL, Su MC, Shiao JY, et al. Amphotericin B irrigation for the treatment of chronic rhinosinusitis without nasal polyps: a randomized, placebo-controlled, double-blind study. *Am J Rhinol.* 2008;22(1):52-58.
- Ponikau JU, Sherris DA, Kita H, Kern EB. Intranasal antifungal treatment in 51 patients with chronic rhinosinusitis. J Allergy Clin Immunol. 2002;110(6):862-866.
- Johnson EM, Ojwang JO, Szekely A, Wallace TL, Warnock DW. Comparison of in vitro antifungal activities of free and liposome-encapsulated nystatin with those of four amphotericin B formulations. *Antimicrob Agents Chemother*. 1998;42(6):1412-1416.
- Offner F, Krcmery V, Boogaerts M, et al. Liposomal nystatin in patients with invasive aspergillosis refractory to or intolerant of amphotericin B. Antimicrob Agents Chemother. 2004;48(12):4808-4812.



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