

Abstract
3221

Kevertin induces p53-dependent and independent cell cycle arrest and apoptosis in ovarian cancer cell lines representing heterogeneous histologies

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Introduction

Ovarian cancer (OC) is a molecularly and histologically heterogeneous disease; however, standard treatment is the same for all subtypes. High-grade serous OC initially responds to chemotherapy; however, low-grade serous and clear cell OC are relatively chemo-resistant. Limited treatment options are available upon recurrence. p53 mutations are found in over 90% of high-grade serous OC. Low-grade serous OC harbor wild type p53, but contain other mutations. During later stages of OC, tumors are a heterogeneous population of mutant cells; thus development of a novel drug that addresses these molecular differences is highly desirable.

Previously, we showed that Kevertin stabilized wild type p53 and induced transcriptional targets in human lung carcinoma. We sought to validate Kevertin as a potential treatment for OC with varied p53 status. Endometrioid carcinoma (A2780, wt p53), high-grade serous (OVCAR-3, mutant p53; OV-90, mutant p53) and atypical non-serous clear cell (SKOV-3, partially deleted p53) OC cell lines and tumors were treated with Kevertin for our studies.

Methodology

RNA Isolation and Quality Control: Human cell culture samples were submitted to Ocean Ridge Biosciences (ORB, Deerfield Beach, FL) for mRNA-Sequencing. Cell lines, A2780, OVCAR-3, OV90 and SKOV-3, were treated with 100 μM, 200 μM, or 400 μM Kevertin (KCI salt) and compared to untreated controls. Xenograft tissue samples treated with Kevertin were also submitted to ORB for mRNA-Sequencing. Total RNA was isolated from cell culture samples using the TRI Reagent® method. Following isolation, RNA was quantified by OD measurement, and assessed for quality on a 1% agarose 2% formaldehyde RNA Quality Control (QC) gel. RNA was digested with RNase-free DNase I and re-purified using Agencourt RNAClean XP bead purification kit. The newly digested RNA samples were then quantified by OD measurement.

For small RNA: RNA was then digested with RNase free DNase I and filtered through Nanospin 100K Omega columns and re-purified on RNeasy MinElute columns using an alternative high-ethanol binding condition to preserve Low Molecular Weight (LMW) RNA. The newly prepared LMW RNA samples were then quantified by OD measurement.

Library Preparation (mRNA): Amplified cDNA libraries suitable for sequencing were prepared from 250-500 nanograms (ng) of DNA-free total RNA using the TruSeq Stranded Total RNA Library Prep Kit with Ribo-Zero Gold (Illumina Inc., San Diego, CA). The quality and size distribution of the amplified libraries were determined by chip-based capillary electrophoresis (LabChip GX microfluidic system, Caliper Life Sciences). Libraries were quantified using the KAPA Library Quantification kit.

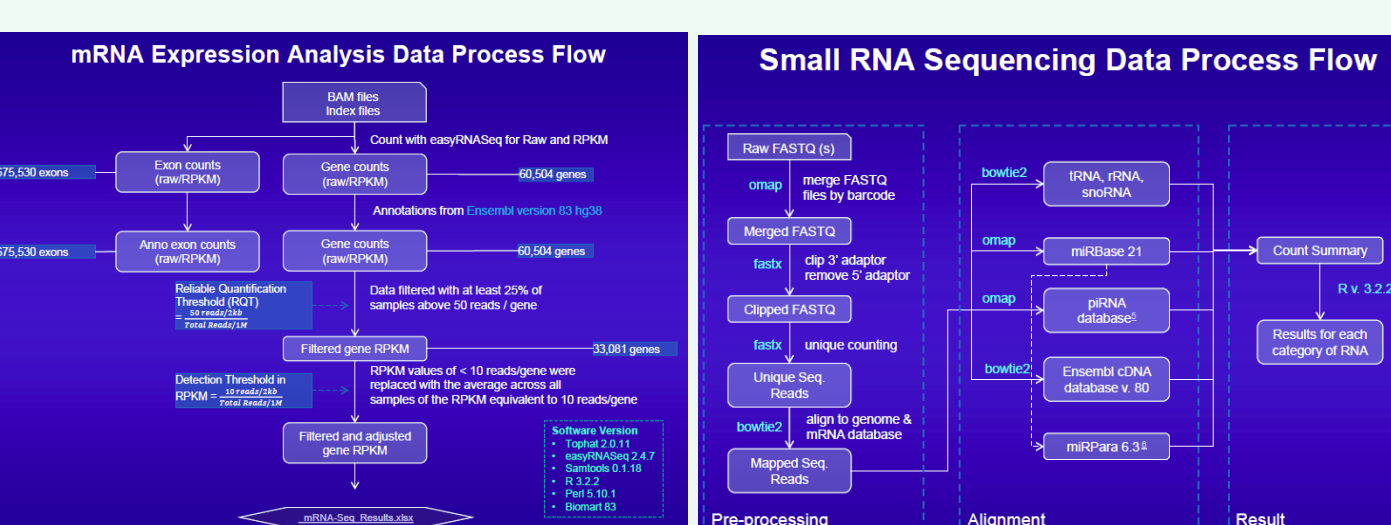
Library Preparation (small RNA): Templated DNA molecules suitable for cluster generation were prepared from the isolated RNA samples using the CleanTag Small RNA Library Prep Kit according to the manufacturer's instructions. Following the CleanTag preparation, purified libraries were electroporated through a freshly cast 6% native acrylamide gel. Library fragments of 130-160 nt corresponding to inserts of 11-41 nt were excised from the gel and recovered by overnight agitation at 37°C at 200 RPM in elution buffer, passage of the eluate through a 0.45μm filter, and ethanol precipitation. The quality and size distribution of the amplified libraries were determined by chip-based capillary electrophoresis.

Sequencing: The libraries were pooled at equimolar concentrations and diluted prior to loading onto the flow cell of the Illumina HiSeq cluster station. The libraries were extended and bridge amplified to create sequence clusters using the Illumina HiSeq PE Cluster Kit v4 and sequenced on an Illumina HiSeq Flow Cell v4 with 50 nt paired-end reads plus dual index reads using the Illumina HiSeq SBS Kit v4. Real time image analysis and base calling were performed on the instrument using the HiSeq Sequencing Control Software version 2.2.56.

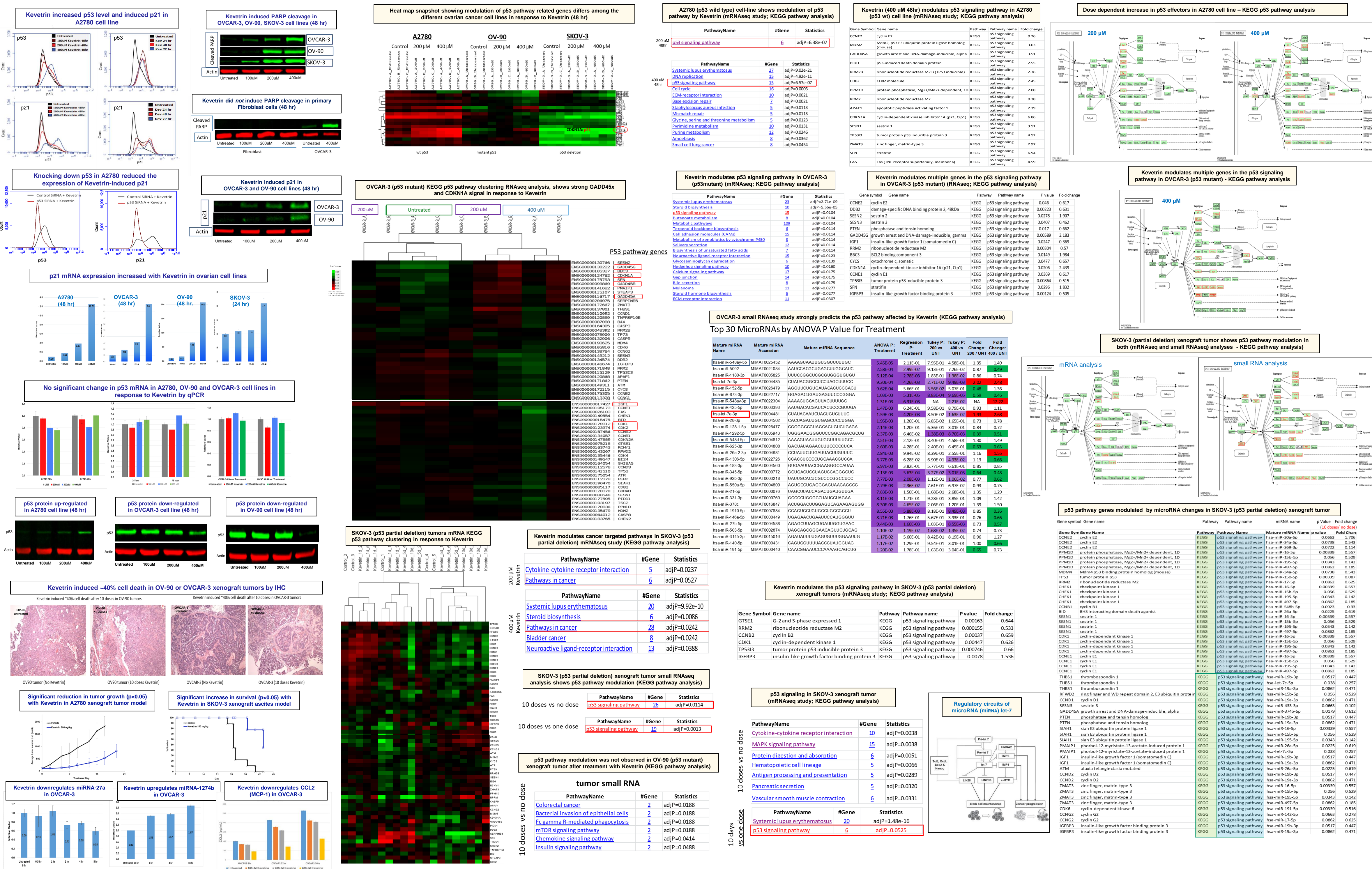
Statistical Analysis: A two-way ANOVA was performed to compare the overall effects of the cell line, treatment and the interaction of cell line and treatment on gene expression. Replicates for each cell line were treated as random factors in the ANOVA. Tukey's Honest Significant Difference (HSD) tests were performed to compare cell lines within each treatment, and to compare treatments within each cell line. Fold changes were also calculated for the same comparisons as the Tukey tests using the mean of each group being compared. If the mean of both groups considered in the fold change comparison was below RQT, 'NA' was reported. All statistical analysis was performed using R version 3.2.2 statistical computing software.

For xenograft samples, one-way ANOVA was performed to compare the effects of the number of days of Kevertin dosing on mRNA expression. Tukey's HSD tests were used to identify significant differences among the individual treatment groups. The Tukey test generates P-values for all possible pair-wise comparisons and then adjusts the P-values upwards based on the total number of comparisons tested, which can be calculated as $(n/2 * n - 1)$, where n is the number of groups. Fold changes were also calculated for the same comparisons as the Tukey tests using the mean of each group being compared. If the mean of both groups considered in the fold change comparison was below RQT, 'NA' is reported.

Linear regression on days of Kevertin treatment was also performed. The strength of the regression fit is indicated by Regression Coef. Day, which is negative when mRNA expression decreases with time and positive when mRNA expression increases with time. The Regression Adjusted R² values indicates the fraction of variance explained by the model, taking into account the number of explanatory variables in the model.



Key Findings



Results Summary

- Kevertin induced apoptosis in wild type p53, mutant p53 and partially deleted p53 ovarian cancer cell lines as assayed by cleavage of PARP.
- p53 and p21 increased protein levels were seen in A2780 cells (p53 wild type) after 24 to 48 hr exposure to Kevertin. Depletion of p53 by siRNA reduced the p21 expression as assayed by FACS.
- Kevertin downregulated oncogenic mutant p53 in OVCAR-3 and OV-90 (p53 mutant)
- Increase in p21 mRNA in response to Kevertin was observed in all OC cell lines.
- No significant changes were observed in p53 mRNA.
- In xenograft models, Kevertin inhibited A2780 tumor growth, and increased SKOV-3 animal survival.
- Transcriptomic analyses demonstrated that Kevertin modulates p53 signaling pathways and induces apoptosis and cell cycle arrest in ovarian cancer cell lines as well as xenograft tumors as shown by RNAseq data. KEGG pathway analysis was used for these studies.
- Altered expression of miRNA-27a, miRNA-1274b, miRNA-25 (known to be dysregulated in ovarian cancer) in a time-dependent manner in OVCAR-3 cells provides possible biomarkers for Kevertin response.
- In tumors where p53 is partially deleted (SKOV-3), Kevertin modulates p53 pathways and induces apoptosis. Further detailed mechanism studies are ongoing.

Conclusions

- Kevertin regulates multiple molecular targets and signaling pathways.
- Kevertin induces wild type p53 and induces apoptosis in a non-genotoxic manner (previously shown). Most currently available chemotherapeutic drugs are genotoxic and damage DNA.
- Targeting multiple pathways such as the p53 signaling pathways has benefits of anti-tumor activity, reducing toxicity and overcoming development of drug resistance.
- In a completed Phase 1 clinical trial, Kevertin was well tolerated with minimal adverse effects. A clinical trial in ovarian cancer is underway.

Further Information

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