Exceptional Simian Hemorrhagic Fever Virus Diversity in a Wild African Primate Community

Michael Lauck,a Samuel D. Sibley,b David Hyeroba,c Alex Tumukunde,c Geoffrey Weny,c Colin A. Chapman,c,d Nelson Ting,e William M. Switzer,f Jens H. Kuhn,g Thomas C. Friedrich,b,h David H. O’Connor,a,h Tony L. Goldberg,b,c,h

Department of Pathology and Laboratory Medicine, University of Wisconsin—Madison, Madison, Wisconsin, USA; Department of Pathobiological Sciences, University of Wisconsin—Madison, Madison, Wisconsin, USA; Makerere University, Kampala, Uganda; Department of Anthropology and School of Environment, McGill University, Montreal, Quebec, Canada; Department of Anthropology and Institute of Ecology and Evolution, University of Oregon, Eugene, Oregon, USA; Laboratory Branch, Division of HIV/AIDS Prevention, National Center for HIV, Hepatitis, STD, and TB Prevention, Centers for Disease Control and Prevention, Atlanta, Georgia, USA; Integrated Research Facility at Fort Detrick, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Fort Detrick, Frederick, Maryland, USA; Wisconsin National Primate Research Center, Madison, Wisconsin, USA

Simian hemorrhagic fever virus (SHFV) is an arterivirus that causes severe disease in captive macaques. We describe two new SHFV variants subclinically infecting wild African red-tailed guenons (Cercopithecus ascanius). Both variants are highly divergent from the prototype virus and variants infecting sympatric red colobus (Procolobus rufomitratus). All known SHFV variants are monophyletic and share three open reading frames not present in other arterviruses. Our data suggest a need to modify the current arterivirus classification.

Simian hemorrhagic fever virus (SHFV) is a member of the family Arteriviridae, together with equine arteritis virus (EAV), lactate dehydrogenase elevating virus (LDV), and porcine reproductive and respiratory syndrome virus (PRRSV) (1). SHFV was first isolated from captive macaques (Macaca sp.) in 1964 after nearly simultaneous outbreaks in Soviet and American primate centers (2–4), possibly having originated from subclinically infected wild-caught patas monkeys (Erythrocebus patas), green monkeys (Chlorocebus aethiops), or guinea baboons (Papio papio) (5,6). Much of what is known about SHFV comes from prototype variants LVR 42-0/M6941 and Sukhumi, isolated during the original two outbreaks, and their derivatives (4,7).

We recently discovered two novel SHFV variants infecting a male red colobus monkey (Procolobus rufomitratus) from Kibale National Park, Uganda (8). These viruses are highly divergent from each other and the prototype variants but, like the prototype variants, contain three unique open reading frames (ORFs) (2a, 2b, and 3) downstream from the replicase-encoding ORFs (8,9). This genomic architecture may be characteristic of the SHFV taxon.

Here, we report the discovery of two novel SHFV variants in red-tailed guenons (Cercopithecus ascanius) from the same location where we previously reported SHFV in a red colobus (8). In 2010, we sampled 13 Kibale red-tailed guenons as part of a larger study of primate ecology, conservation, and health (10). Animals were anesthetized and samples were collected as previously described (8). Viral RNA was prepared from blood plasma for direct sequencing as previously described (8), with minor modifications for sequencing on an Illumina MiSeq instrument. De novo assembly of sequence reads yielded complete SHFV coding genomes from three individuals (two females, RT05 and RT11, and one male, RT10). A fourth nearly complete SHFV coding genome was recovered from another male individual, RT09, and small gaps were filled by PCR and 3’ rapid amplification of cDNA ends (RACE) followed by Sanger sequencing (8).

Viral genomes were annotated with CLC Genomics Workbench version 5.5 (CLC Bio, Aarhus, Denmark), and putative ORFs were confirmed by querying the NCBI GenBank database (11). Open reading frames were individually aligned with prototype variant LVR 42-0/M6941 and red colobus variants SHFV-krc1 and SHFV-krc2 using a codon-based version of the MAFFT algorithm (12) implemented in Translator X (13). Individual ORF alignments were then concatenated, nucleotide-level similarities of resulting full-length coding genomes were calculated using MEGA5 (14), and sliding-window plots of inferred amino acid similarity were created with SimPlot version 3.5.1 (15). Phylogenetic relationships within the family Arteriviridae were estimated from nucleotide sequences of homologous ORFs of representative full-length coding genomes, aligned as described above but with regions of ambiguous alignment removed using Gblocks (16). Bayesian trees were constructed from concatenated alignments using MrBayes version 3.2.1 (17), with a substitution model of the form GTR+I+Γ selected using jModelTest (Akaike information criterion [AIC], ΔAIC to second-best model GTR+Γ=71.7) (18), model parameters estimated from the data under default priors, and Markov chains run for one million generations, with the first 25% of sampled trees discarded as burn-in.

All four new viruses from red-tailed guenons contain two large replicase-encoding ORFs and several smaller downstream ORFs, each similar in size to homologous ORFs of the prototype variants and variants from red colobus (Fig. 1). As in other arterviruses, ORF1a and ORF1b in the new SHFV variants contain a canonical heptanucleotide “slippery sequence” (UUUAAC) and predicted downstream pseudoknot structure (19). The new viral genomes contain three ORFs, 2a, 2b, and 3, immediately downstream of the replicase, indicating conservation of this characteristic 3’ genomic architecture among all SHFV
variants. Across the aligned coding genomes, the viruses from individuals RT05 and RT11 were similar to each other at the nucleotide level (94.3%), as were the viruses from individuals RT09 and RT10 (93.7%). However, these two variants were only 79.4% similar to each other at the nucleotide level and less similar still to prototype variant LVR 42-0/M6941 (54.1%) or variants from Kibale red colobus (50.1%), with variable amino acid conservation across the genomes (Fig. 1).

Our Bayesian phylogeny is consistent with established relationships of arteriviruses (8,20, 21), supporting the monophyly of SHFV variants, the sister taxon relationship of LDV and PRRSV, and the divergence of EAV (Fig. 2). Within the SHFV clade, the new variants are highly divergent from the prototype variant and from SHFV variants found in sympatric red colobus. Based on their phylogenetic positions (Fig. 2), we designate the new viruses SHFV-krtg1 and SHFV-krtg2, to indicate their origins in Kibale red-tailed guenons and to reflect nomenclature previously used to describe simian immunodeficiency virus (SIV) and SHFV variants infecting Kibale red colobus (8,22).

In Kibale, red-tailed guenons frequently form multispecies social groups with red colobus, in which occasional direct contact occurs (23,24). Nevertheless, the phylogenetic divergence between SHFV from red-tailed guenons and SHFV from sympatric red colobus is approximately equivalent to that between PRRSV and LDV, which are currently assigned to different viral species. This observation strongly suggests that virus-host co-evolution, rather than geography or ecological overlap, shapes the phylogeny of SHFV. Primates of the subfamilies Cercopithecinae and Colobinae diverged approximately 18 million years ago (25); cocirculating, divergent SHFV variants in both red-tailed guenons and red colobus may indicate ancient diversification of SHFV within sympatric host species or, possibly, more recent admixture of viruses due to transmission from other as-yet-unidentified hosts.

The unique genomic architecture of SHFV is conserved even across highly divergent SHFV variants. Furthermore, all SHFV variants described to date are monophyletic, even though SHFV ORFs 2a, 2b, and 3 have no homologs in the other arteriviruses and could not be included in our phylogenetic analyses of the *Arteriviridae*. Currently, the family *Arteriviridae* includes a single genus, *Arterivirus* (26). Given the unique and characteristic genomic architecture of all SHFV variants described to date, the monophyly of SHFV within the family *Arteriviridae*, and the association of SHFV with simian hosts, we suggest the reclassification of SHFV into a new genus: Simartevirus. The discovery of additional arteriviruses will help clarify the appropriateness of this proposed taxonomy, as well as any other taxonomic subdivisions within the family *Arteriviridae* that may be justified.
FIG 2 Bayesian phylogenetic tree of newly discovered simian hemorrhagic fever viruses (SHFV) from red-tailed guenon and other arteriviruses, based on a 9,891 nucleotide alignment of homologous ORFs (1a, 1b, 2a, 2b, 3, 4, 5, 6, and 7, with reference to the EAV genome). New SHFV variants SHFV-krtg1a and -krtg1b (SHFV-krtg1a/b) (from individuals RT05 and RT11) and SHFV-krtg2a/b (from individuals RT09 and RT10) are highlighted (GenBank accession numbers JX473847 to JX473850). Other viruses represent the known diversity within each viral species based on available full-genome sequences, as follows (GenBank accession numbers in parentheses): SHFV-LVR, the SHFV prototype variant LVR 42-0/M6941 (NC_003092); SHFV-krc1 and SHFV-krc2 from Kibale red colobus (HQ845737 and HQ845738, respectively); PRRSV-Lelystad, the European (type 1) type strain (M96262); PRRSV-VR2332, the North American (type 2) type strain (U87392); EAV-Bucyrus (NC_002532); EAV-s3685 (GGQ903794); LDV-P, the Plagemann strain (U15146); and LDV-C, the neurovirulent type C strain (L13298). Posterior clade probabilities are shown on branches; scale bar indicates nucleotide substitutions per site.

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REFERENCES