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# Program: twoACEvj.R
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# Date: 10 22 2018
#
# Twin Bivariate ACE model to estimate causes of variation across multiple groups
# Matrix style model - Raw data - Continuous data
# -----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
# Load Libraries & Options
rm(list=ls())
library(OpenMx)
library(psych); library(polycor)
source("miFunctions.R")

# Create Output
filename <- "twoACEvj"
sink(paste(filename, ".R", sep=""), append=FALSE, split=TRUE)

# -----
# PREPARE DATA

# Load Data
data(twinData)
dim(twinData)
describe(twinData[,1:12], skew=F)
twinData[, 'ht1'] <- twinData[, 'ht1']*10
twinData[, 'ht2'] <- twinData[, 'ht2']*10
twinData[, 'wt1'] <- twinData[, 'wt1']/10
twinData[, 'wt2'] <- twinData[, 'wt2']/10

# Select Continuous Variables
varsc <- c('ht') # list of continuous variables names
nvc <- 1 # number of continuous variables
ntvc <- nvc*2 # number of total continuous variables
conVars <- paste(varsc, c(rep(1, nvc), rep(2, nvc)), sep="")

# Select Ordinal Variables
nth <- 3 # number of thresholds
varso <- c('wt') # list of ordinal variables names
nvo <- 1 # number of ordinal variables
ntvo <- nvo*2 # number of total ordinal variables
ordVars <- paste(varso, c(rep(1, nvo), rep(2, nvo)), sep="")
ordData <- twinData
wtquant <- quantile(ordData[, c('wt1', 'wt2')], (0:(nth+1))/(nth+1), na.rm=TRUE)
for (i in c('wt1', 'wt2')) { ordData[, i] <- cut(ordData[, i], breaks=wtquant, labels=c(0:nth)) }

# Select Variables for Analysis
vars <- c('ht', 'wt') # list of variables names
nv <- nvc+nvo # number of variables
ntv <- nv*2 # number of total variables
oc <- c(0, 1) # 0 for continuous, 1 for ordinal variables
selVars <- paste(vars, c(rep(1, nv), rep(2, nv)), sep="")

# Select Data for Analysis
mzData <- subset(ordData, zyg==1, selVars)
dzData <- subset(ordData, zyg==3, selVars)
mzDataF <- cbind(mzData[, conVars], mxFactor( x=mzData[, ordVars], levels=c(0:nth) ))
dzDataF <- cbind(dzData[, conVars], mxFactor( x=dzData[, ordVars], levels=c(0:nth) ))

# Generate Descriptive Statistics
apply(mzData, 2, myMean)
apply(dzData, 2, myMean)
sapply(mzData[, ordVars], table)
sapply(dzData[, ordVars], table)
hetcor(mzData)$cor
hetcor(dzData)$cor

# Set Starting Values
frMV <- c(TRUE, FALSE) # free status for variables
svMe <- c(15, 0) # start value for means
svPa <- .2 # start value for path coefficient
svPe <- .5 # start value for path coefficient for e
svLTh <- -.5 # start value for first threshold
svITH <- 1 # start value for increments
svTh <- matrix(rep(c(svLTh, (rep(svITH, nth-1)))), nrow=nth, ncol=nv) # start value for thresholds
lbTh <- matrix(rep(c(-3, (rep(0.001, nth-1))), nv), nrow=nth, ncol=nv) # lower bounds for thresholds

# -----
# PREPARE MODEL

# Create Algebra for expected Mean & Threshold Matrices

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meanG    <- mxMatrix( type="Full", nrow=1, ncol=ntv, free=frMV, values=svMe, labels=labVars("mean",vars), name="meanG" )
thinG    <- mxMatrix( type="Full", nrow=nth, ncol=ntvo, free=TRUE, values=svTh, lbound=lbTh, labels=labTh("th",vars,nth),
name="thinG" )
inc      <- mxMatrix( type="Lower", nrow=nth, ncol=nth, free=FALSE, values=1, name="inc" )
threG    <- mxAlgebra( expression= inc %*% thinG, name="threG" )

# Create Matrices for Variance Components
covA     <- mxMatrix( type="Symm", nrow=nv, ncol=nv, free=TRUE, values=valDiag(svPa,nv), label=labLower("VA",nv), name="VA" )
covC     <- mxMatrix( type="Symm", nrow=nv, ncol=nv, free=TRUE, values=valDiag(svPa,nv), label=labLower("VC",nv), name="VC" )
covE     <- mxMatrix( type="Symm", nrow=nv, ncol=nv, free=TRUE, values=valDiag(svPa,nv), label=labLower("VE",nv), name="VE" )

# Create Algebra for expected Variance/Covariance Matrices in MZ & DZ twins
covP     <- mxAlgebra( expression= VA+VC+VE, name="V" )
covMZ    <- mxAlgebra( expression= VA+VC, name="cMZ" )
covDZ    <- mxAlgebra( expression= 0.5*x%VA+ VC, name="cDZ" )
expCovMZ <- mxAlgebra( expression= rbind( cbind(V, cMZ), cbind(t(cMZ), V)), name="expCovMZ" )
expCovDZ <- mxAlgebra( expression= rbind( cbind(V, cDZ), cbind(t(cDZ), V)), name="expCovDZ" )

# Constrain Variance of Binary Variables
matUnv   <- mxMatrix( type="Unit", nrow=nvo, ncol=1, name="Unv1" )
matOc    <- mxMatrix( type="Full", nrow=1, ncol=nv, free=FALSE, values=oc, name="Oc" )
var1     <- mxConstraint( expression=diag2vec(Oc%&%V)=Unv1, name="Var1" )

# Create Data Objects for Multiple Groups
dataMZ   <- mxData( observed=mzDataF, type="raw" )
dataDZ   <- mxData( observed=dzDataF, type="raw" )

# Create Expectation Objects for Multiple Groups
expMZ    <- mxExpectationNormal( covariance="expCovMZ", means="meanG", dimnames=selVars, thresholds="threG",
threshnames=ordVars )
expDZ    <- mxExpectationNormal( covariance="expCovDZ", means="meanG", dimnames=selVars, thresholds="threG",
threshnames=ordVars )
funML    <- mxFitFunctionML()

# Create Model Objects for Multiple Groups
pars     <- list(meanG, thinG, inc, threG, matUnv, matOc, covA, covC, covE, covP )
modelMZ  <- mxModel( pars, covMZ, expCovMZ, dataMZ, expMZ, funML, name="MZ" )
modelDZ  <- mxModel( pars, covDZ, expCovDZ, dataDZ, expDZ, funML, name="DZ" )
multi    <- mxFitFunctionMultigroup( c("MZ","DZ" ) )

# Create Algebra for Standardization
matI     <- mxMatrix( type="Iden", nrow=nv, ncol=nv, name="I" )
invSD    <- mxAlgebra( expression=solve(sqrt(I*V)), name="iSD" )

# Calculate genetic and environmental correlations
corA     <- mxAlgebra( expression=solve(sqrt(I*VA))%&%VA, name="rA" ) #cov2cor()
corC     <- mxAlgebra( expression=solve(sqrt(I*VC))%&%VC, name="rC" )
corE     <- mxAlgebra( expression=solve(sqrt(I*VE))%&%VE, name="rE" )

# Create Algebra for Unstandardized and Standardized Variance Components
rowUS    <- rep('US',nv)
colUS    <- rep(c('VA','VC','VE','SA','SC','SE'),each=nv)
estUS    <- mxAlgebra( expression=cbind(VA,VC,VE,VA/V,VC/V,VE/V), name="US", dimnames=list(rowUS,colUS) )

# Create Confidence Interval Objects
odd      <- seq(1+3*nv,2+3*nv,nv)
even     <- seq(2+3*nv,2+3*nv,nv)
ciACE    <- mxCI( c("US[1,odd]","US[2,odd]","US[2,even]" ) )

# Build Model with Confidence Intervals
calc     <- list( matI, invSD, corA, corC, corE, estUS, ciACE )
modelACE <- mxModel( "twoACEvj", pars, var1, modelMZ, modelDZ, multi, calc )

# -----
# RUN MODEL

# Run ACE Model
fitACE   <- mxRun( modelACE, intervals=T )
sumACE   <- summary( fitACE )

# Compare with Saturated Model
#mxCompare( fitSAT, fitACE )

# Print Goodness-of-fit Statistics & Parameter Estimates
fitGofs(fitACE)
fitEstCis(fitACE)

# -----
# RUN SUBMODELS

# Run AE model
modelAE <- mxModel( fitACE, name="twoAEvj" )

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modelAE <- omxSetParameters( modelAE, labels=labLower("VC",nv), free=FALSE, values=0 )
modelAE <- omxSetParameters( modelAE, labels=labLower("VA",nv), free=TRUE, values=.6 )
fitAE <- mxRun( modelAE, intervals=T )
fitGofs(fitAE); fitEstCis(fitAE)

# Run CE model
modelCE <- mxModel( fitACE, name="twoCEvj" )
modelCE <- omxSetParameters( modelCE, labels=labLower("VA",nv), free=FALSE, values=0 )
modelCE <- omxSetParameters( modelCE, labels=labLower("VC",nv), free=TRUE, values=.6 )
fitCE <- mxRun( modelCE, intervals=T )
fitGofs(fitCE); fitEstCis(fitCE)

# Run E model
modelE <- mxModel( fitAE, name="twoEvj" )
modelE <- omxSetParameters( modelE, labels=labLower("VA",nv), free=FALSE, values=0 )
fitE <- mxRun( modelE, intervals=T )
fitGofs(fitE); fitEstCis(fitE)

# Print Comparative Fit Statistics
mxCompare( fitACE, nested <- list(fitAE, fitCE, fitE) )
#round(rbind(fitACE$US$result, fitAE$US$result, fitCE$US$result, fitE$US$result),4)

# -----
sink()
save.image(paste(filename, ".Ri", sep=""))

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