Chameleon-Inspired Strain-Accommodating Smart Skin

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Supporting Information

ABSTRACT: Stimuli-responsive color-changing hydrogels, commonly colored using embedded photonic crystals (PCs), have potential applications ranging from chemical sensing to camouflage and anti-counterfeiting. A major limitation in these PC hydrogels is that they require significant deformation (>20%) in order to change the PC lattice constant and generate an observable chromatic shift (~100 nm). By analyzing the mechanism of how chameleon skin changes color, we developed a strain-accommodating smart skin (SASS), which maintains near-constant size during chromatic shifting. SASS is composed of two types of hydrogels: a stimuli-responsive, PC-containing hydrogel that is patterned within a second hydrogel with robust mechanical properties, which permits strain accommodation. In contrast to conventional “accordion”-type PC responsive hydrogels, SASS maintains near-constant volume during chromatic shifting. Importantly, SASS materials are stretchable (strain ~150%), amenable to patterning, spectrally tunable, and responsive to both heat and natural sunlight. We demonstrate examples of using SASS for biomimicry. Our strategy, to embed responsive materials within a mechanically matched scaffolding polymer, provides a general framework to guide the future design of artificial smart skins.

KEYWORDS: magnetic particles, chameleon-inspired structural color, light-responsive hydrogels, strain accommodating, photonic crystals, chromatic materials

Any living creatures have evolved the ability to change color, such as chameleons,† neon tetra fishes,‡ and Morpho sulikowskyi butterflies.§ The ability to rapidly change color is beneficial for camouflage, mating, and intimidating predators. Coloration in these organisms is described as “structural” because it is generated by periodic arrays of nano- and microstructures, known as photonic crystals (PCs), which generate color through optical interference effects. Unlike common coloration strategies using dyes and pigments, which rely on wavelength-specific absorption of light, structural coloration is immune to photobleaching and can be easily manipulated by controlling particle spacing as opposed to synthetic strategies. In accordance with Bragg’s law of diffraction,5,6 the periodic distance between elements in these natural PCs dictates the color of the material. Accordingly, the mechanism of dynamic color change for many of these species employs responsive matrices that tune the spacing of the PCs.

Inspired by natural examples, there has been significant effort focused on generating synthetic responsive PC materials that can change color on demand.8–13 Such color-changing materials, which are sometimes described as “smart skins”, are envisioned to become important in various applications ranging from camouflage and communications to anti-counterfeiting and biosensing. The fundamental mechanism for adjusting the color of synthetic PCs mirrors that of natural systems and requires modulating the periodicity of the PC lattice using an “accordion” mechanism.14–17 This strategy is usually realized by embedding the PC in a hydrogel18–21 or flexible polymer matrix22,23 that responds to an input by expansion or contraction, resulting in a shift in the PC periodicity. The major problem with this accordion strategy of creating artificial smart skins pertains to the large volume change, which leads to structural instability and buckling of the material.24,25 For example, a 20% or greater linear expansion or contraction (strain) of a responsive PC is required to produce a significant color shift (∆λ = 100 nm) (Supplementary S1). This magnitude of strain and deformation impedes their application and adoption. However, this led us to consider how color-changing
animals, such as chameleons, address this problem (Figure 1a). On the basis of recent time-lapse imaging of chameleon skin,1 we noticed that only a small fraction of the skin cells contain the guanine PC arrays, while the remainder of the skin cells are colorless. In other words, chameleon skin tissue includes nonresponsive cells that may contribute to reduce the global volume change during chromatic response.

This general concept is shown in Figure 1b, which highlights the contrast between artificial responsive PCs that have been extensively studied and natural responsive PCs, which employ a tunable PC element embedded within a matrix that lacks PCs and accommodates the strain. This structural difference, which was shown in natural responsive PCs, leads to a significant reduction in the total change in volume of the material, while maintaining the observable dynamic coloration.

Herein, we demonstrate a strain-accommodating smart skin (SASS) inspired by the structure of natural responsive PCs. The total volume of our SASS structure is maintained because the PC contraction is accommodated by local elastic deformation of a supporting polymer layer. SASS materials display excellent mechanical properties including high stretchability and good mechanical strength that is primarily dependent on the supporting layer. SASS demonstrates a generalizable concept of how to create responsive PCs that maintain constant size during chromatic shifting. Furthermore, we show that SASS color change can be optically triggered and SASS films rapidly changed color upon exposure to ambient sunlight (t = 10 min). These features suggest the potential utility of SASS in applications such as camouflage and anti-counterfeiting.

RESULTS AND DISCUSSION
To recapitulate the strain-accommodating behavior of natural responsive PCs and predict the optimal material design, we first performed finite element analysis modeling of mechanical strain.

Figure 1. Design and fabrication of strain-accommodating smart skin (SASS) material. (a) Photographs of a chameleon displaying color change (adapted with permission from Teyssier, J., Saenko, S. V., van der Marel, D., and Milinkovitch, M. C. Nat. Commun. 2015, 6, 6368; copyright 2015 Springer Nature).1 (b) Schematic comparing the typical design of artificial responsive PCs to responsive PCs utilized by nature. The dots represent the PC, yellow and green represent the swollen and deswollen states of the responsive hydrogel, respectively, and gray indicates the supporting, nonresponsive polymer matrix. (c) Model used for finite element analysis (FEA) of SASS (also see Figure S1). (d) Rendering of FEA model mapping the deformation of a bilayer material with a supporting nonresponsive layer and a responsive upper layer divided into sections. (e) Plot of the FEA-determined average force and maximal displacement within a plane of the supporting polymer layer as a function of the number of squares in the responsive PC array layer. (f) Photograph and schematic of SASS that includes dimensions and chemical structure of polymers. (g) Flow diagram illustrating the steps used in SASS fabrication, where TEM and SEM images show representative structures of the magnetic nanoparticles and their organization with the responsive PC (scale bar of SEM image: 5 and 1 μm (inset)).
Specifically, we modeled a patterned array of responsive PC hydrogels that were embedded within a nonresponsive supporting polymer layer (Figure 1c and d, also see Figure S1 in Supplementary S2). The geometry of the model was such that the responsive PC elements were localized to the top face of the material. When we fixed the Young’s modulus of the two materials and assumed a 50% change in length of the responsive layer, we found that a single planar layer of responsive hydrogel placed atop a supporting layer led to significant strain and deformation at the edges (Figure 1d, 1×1 case). Note that in our model a free-standing responsive hydrogel film displayed the expected deformation (Figure S2). Representative uniaxial tensile measurement of a SASS sample (red) and a sample of a standalone film comprised of the strain-accommodating polymer (black). (h) Photographs demonstrating the elasticity of SASS and its strain-induced color change. (i) Photographs showing the chromatic response of a traditional responsive pNIPAM PC film (upper) triggered by adjusting the temperature (20 to 40 °C) compared to the chromatic response of SASS (lower) under the same conditions. The dimensions of the materials in the initial and final states are provided to the left of each image. All error bars represent the standard deviation of three independent measurements.

Figure 2. Characterization of SASS. (a) Plot relating the cross-linker (PEGDA) concentration in pNIPAM PC gels to their area before and after heating to 45 °C (a 350 G magnetic field was applied during polymerization). The right y-axis represents the area change % upon heating for each composition. (b) Plot showing the change in reflection λ_max for the samples prepared in (a). The Δλ_max was determined using reflection spectra of standalone PC films at RT and 45 °C. (c) Plot of reflection Δλ_max (collected at RT and 45 °C) for responsive pNIPAM PCs polymerized under different magnetic field strengths. These gels were synthesized with 120 mM PEGDA. (d) Plot of temperature-dependent strain of pNIPAM hydrogels prepared without (black) and with magnetic nanoparticles (red). (e) Normalized reflection spectra of a SASS sample collected at different temperatures. (f) Plot showing λ_max of reflectance for SASS samples as a function of temperature. (g) Representative uniaxial tensile measurement of a SASS sample (red) and a sample of a standalone film comprised of the strain-accommodating polymer (black). (h) Photographs demonstrating the elasticity of SASS and its strain-induced color change. (i) Photographs showing the chromatic response of a traditional responsive pNIPAM PC film (upper) triggered by adjusting the temperature (20 to 40 °C) compared to the chromatic response of SASS (lower) under the same conditions. The dimensions of the materials in the initial and final states are provided to the left of each image. All error bars represent the standard deviation of three independent measurements.
flavin photoinitiator, NIPAM monomer), and while applying a magnetic field, UV light was used to cross-link the hydrogel, thus locking nanoparticles into defined periodic arrays. Subsequently, a mold (Figure S5) was used to create a square array of responsive PC hydrogels (4 × 4 mm squares, spaced by 0.5 mm). Finally, the tetra-PEG supporting layer precursor was cured onto the responsive PC array at room temperature. A photograph of a SASS film with a 5 × 5 responsive PC array demonstrating its color and elasticity is shown in Figure 1f. Cross-section scanning electron microscopy (SEM) imaging of the responsive PC layer confirms a periodic chain structure formed by magnetic nanoparticles (MNPs) (Figure 1g), which generates the colorful Bragg diffraction of the sample. Notably, the chain structures appear to be distorted in the SEM images likely due to drying-induced compression of the hydrogel matrix during imaging.

To optimize the chromatic shift of the responsive pNIPAM PC layer, we tested the role of its mechanical properties (by changing cross-linker concentration) in tuning the swelling/deswelling ratio (Figure 2a) and generating a maximal chromatic shift (Figure 2b). Optimization was performed using a reflection-mode spectrometer to record the chromatic shift of pNIPAM PC films in response to thermal heating (45 °C). We found that an intermediate concentration of cross-linker (120 mM PEGDA) produced the greatest magnitude of chromatic shift while minimizing area change. Additionally, optimization of magnetic field strength during synthesis is shown in Figure 2c. We found that if the magnetic field was too strong (385 G), the peak wavelength shift was diminished likely because the periodic distance between particles approaches a minimum, and the collapse of the responsive hydrogel can no longer reduce interparticle distance further. On the contrary, if the magnetic field is too weak (175 G), no color is observed since the field strength is not sufficient to assemble photonic crystals (Figure 2c). Therefore, we fixed our protocol to use 245 G, as it produced a maximum chromatic shift.

These experiments provided the optimal parameters for fabricating the responsive PC hydrogel. Using these optimized conditions, we measured the thermally induced volume transition of PC-containing and non-PC-containing responsive films, as shown in Figure 2d (120 mM PEGDA, 245 G). PC-containing gels displayed a sharper and more significant thermally induced deswelling strain compared to the pNIPAM films lacking the MNPs. This may be due to the strong UV absorption cross-section of MNPs which impacts UV-initiated radical polymerization. Note that the volumetric transition of these films is broader compared to that of conventional pNIPAM films, which is likely due to the PEG chains in the cross-linker (PEGDA) that modulate dehydration of the hydrogel (Figure 2d). The data in Figure 2a support this conclusion since gels with higher PEGDA concentrations display lower deswelling ratios.

Representative reflection spectra and plots of the peak reflection wavelength versus temperature of the as-prepared SASS samples show a clear thermochromatic shift, in agreement with the reported behavior of responsive hydrogels (Figure 2e and f).28,29 The heat-triggered response of the SASS film was similar to that of the responsive PC films, suggesting that the SASS architecture did not significantly modulate the transition temperature of pNIPAM. However, compared to the mechanical strength of conventional pNIPAM hydrogels30 which
display rupture forces of a few kPa, SASS displayed superior mechanical strength. Specifically, uniaxial tensile testing (Figure 2g) showed that SASS has a rupture force of 60 kPa with a strain of 1.5. This was primarily driven by the mechanical properties of the supporting layer, which similarly had an 83 kPa rupture force with a strain of 1.8. We also found that SASS samples could be handled manually and displayed a color change upon stretching (Figure 2h). Note that the chromatic shift generated by the Fe3O4@SiO2 particle chains is sensitive to the orientation of the mechanical strain. Given that the thermal or optically triggered deswelling response generates in-plane forces, we focused on strains parallel to the SASS film. Of course, one can apply an external strain that is perpendicular to the SASS film, and such forces can lead to an opposite red-shift in the reflection peak of the material.

Critically, SASS displayed significant strain-accommodating behavior, as was especially clear when comparing its volume transition to conventional responsive PC films (Figure 2i). To illustrate the difference, SASS and conventional responsive PC films were sandwiched between two plastic Petri dishes such that only in-plane deformations were allowed. To the best of our ability to measure the dimensions of the SASS sample, its size was identical following heating-induced chromatic shifting, in contrast to the 8 × 8 film, which displays reduced deformation.

In addition to its thermoresponsive properties, another intriguing feature of SASS is its light-responsive behavior. Due to the strong light absorption of magnetite nanoparticles (Figure S8),31 we hypothesized that illumination of SASS would also trigger a chromatic response (Figure 3a). Upon exposure of SASS to a hand-held white light LED source, we observed a rapid and reversible reflectance peak shift that mirrored the response to thermal heating (Figure 3b and c). In the samples tested, we found that the reflection λmax of SASS shifted from 600 nm to 525 nm within a few minutes of illumination. To further quantify the photothermal conversion efficiency of MNPs, a calibrated 370 nm light source with ∼20 mW/cm2 intensity was used to heat an aqueous dispersion of MNPs in a quartz cuvette (schematic shown in Figure 3d). As plotted in Figure 3e, the photothermal ideal heating rate can be inferred by extrapolating a tangential line to the initial phase of the heating curve. Combining this data with the equations shown in the Supplementary S4, we inferred a conversion efficiency of 81.8%.

We next aimed to validate the dynamic remodeling of the PC arrays within the responsive hydrogel film upon illumination. To achieve this, we employed high-resolution in situ optical microscopy to excite and simultaneously image the PC layer within the SASS film (Figure 4a). We found that exciting the sample using a 405 nm laser coupled with a quad cube filter set
(405/488/561/647 nm) revealed the weak autofluorescence of the hydrogel. By happenstance, we found that continuous illumination in the 405 nm channel also triggered photothermal heating, localized deswelling of the SASS film, and reorganization of the PC microstructure (Figure 4b). Conveniently, the nanochain structure has significant scattering when imaged using a 535 nm reflection interference contrast microscopy (RICM) filter set. This allowed for concurrent imaging of the PC nanostructures along with the surrounding responsive hydrogel during photothermal heating and cooling (Figure 4c and Figure S10). As expected, the PC arrays (RICM channel) were antilocalized with the polymer hydrogel (405 nm channel). End points and time-lapse images recording a single photothermal heating/cooling cycle are shown in Figure 4b and c (also see Supplementary Movies S1 and S2). In the time-lapse images, the heating phase (0–13.9 s) was recorded using the 405 nm channel, while the cooling phase (13.9–27.8 s) was captured in the RICM channel. The PC chains are displaced and indeed go out of focus due to the isotropic (3D) nature of deswelling. Importantly, the images comparing the positions of the nanochain structure before and after the heating/cooling cycle demonstrate that the SASS microstructure recovers to its initial configuration.

To confirm that the photothermal effect was exclusively attributed to absorption by Fe₃O₄@SiO₂ particles rather than other components of SASS, we fabricated SASS films doped with pure silica particles lacking the Fe₃O₄ core (Figure 4d) and compared the time-dependent PC array displacement in SASS samples to that of films with silica nanoparticles (Figure 4e and Figure S11; also see Supplementary Movies S3 and S4). The lack of response in the silica nanoparticle-containing samples confirms the key role of magnetite in driving the light-triggered chromatic response of SASS.

SASS chromatic response could also be spatially controlled using localized illumination. To demonstrate this capability, a 532 nm laser was focused onto a 2 mm diameter spot on the...
SASS sample at an angle of 50°, and a smart phone camera was used to record the chromatic change perpendicular to the sample (Figure 5a). The sample drastically changed color within a few seconds of laser illumination (Figure 5a; also see Supplementary Movie S5), and the color change was localized to the spot of irradiation. The response was quantified by RGB analysis of the images (Figure 5b). An increase in blue intensity was observed with a concomitant decrease in both red and green intensity. This response is consistent with the expected blue shift in the reflection spectra after deswelling. This result suggests potential applications for SASS involving dynamic chromatic patterning.

Neon tetra fish are well known for their light-responsive behavior, where the lateral stripes display different structural colors after brief exposure to sunlight (Figure 5c).2,3 To mimic this response, we first tested whether SASS is sensitive to natural sunlight. Surprisingly, we found that a 10 min exposure to sunlight triggered an observable chromatic shift (Figure 5d). Specifically, the SASS response $\lambda_{\text{max}}$ shifted from 622 ± 2.2 nm to 573 ± 7.7 nm for samples fabricated with 210 nm diameter MNPs. The response triggered by sunlight was reversible, as the amplitude of the chromatic shift (48 ± 9.6 nm) did not change significantly over the course of six illumination/cooling cycles. The range of the $\lambda_{\text{max}}$ shift response could also be tuned by employing MNPs of a different diameter. For example, particles with a 180 nm diameter showed a response $\lambda_{\text{max}}$ of 556 ± 10.1 nm that reversibly shifts to 504 ± 5.3 nm upon sunlight exposure (Figure 5e). Further mimicking the neon tetra fish response, we fabricated SASS films shaped like a fish (Figure 5f). After a 10 min exposure to natural sunlight, the SASS film shifted from orange to green, while the overall size remained constant, as determined to the best of our abilities (Figure S12).

Given that SASS can be fabricated in different shapes and is sunlight responsive, we aimed to demonstrate camouflage behavior. Hence, we fabricated a leaf-shaped SASS sample and arranged it together alongside real leaves (Figure 5g). After exposure to sunlight, the color of the SASS “leaf” shifted from orange to green and was camouflaged among the real leaves. Again, a key feature enabling camouflage is that the size of the “leaf” remained constant during chromatic shifting.

CONCLUSIONS

In summary, we present an alternative approach to fabricate smart skin materials inspired by the chameleon skin structure. The SASS design solves the long-standing problem of mechanical buckling and instability of conventional responsive PC hydrogels. The key element to SASS is a two-component polymer material where a segmented responsive PC hydrogel is embedded within a supporting polymer that accommodates the strain arising from the volume transition. By optimizing crosslinker concentration and magnetic field strength, we demonstrate a strong and reversible heat/light triggered optical response in SASS. High-resolution optical microscopy confirms light-triggered local deswelling and chromatic response of the SASS film. Moreover, SASS chromatic response can be controlled in space and time using a focused laser beam. Note that the resolution of patterning is limited by the dimensions of the illumination spot, the rate of gel response, and the thermal diffusion coefficient within the sample. Future work will define these parameters. Finally, we show that SASS is sensitive to natural sunlight and can be molded into various geometries to aid in biomimicry. This suggests potential applications in camouflage, signaling, and anti-counterfeiting.

METHODS

Synthesis of Fe3O4@SiO2 Core–Shell Nanoparticles (MNP)s.

Fe3O4@SiO2 core–shell nanoparticles were synthesized according to previously developed methods.22 Briefly, for particles with an average diameter of 180 nm, 0.65 g of FeCl3, 40 mL of ethylene glycol, 3.0 g of sodium acetate, 1.05 g of poly(4-styrenesulfonic acid-co-maleic acid) sodium salt, 14 mg of N-isosorbic acid, and 120 μL of deionized H2O were added consecutively into a Parafilm-sealed beaker. After vigorous magnetic stirring for about 40 min, a homogeneous mixture formed, and then 0.6 g of NaOH was added. The mixture was stirred for 1–2 h until all the NaOH was completely dissolved. The mixture was then transferred into a capped 50 mL Erlenmeyer flask and then allowed to react in a preheated oven (190 °C) for 9 h. The particles were washed with 30 mL of 50 (v/v)% ethanol in water solution three times and then washed three additional times with DI water.

To synthesize the silica shell, a 12 mL aliquot of Fe3O4 dispersion was mixed with 80 mL of ethanol and 4 mL of ammonium hydroxide (25–28 wt %) under vigorous mechanical stirring for 1 min. This solution was warmed by using a water bath at 50 °C. Subsequently, two aliquots of 0.4 mL of tetraethyl orthosilicate (TEOS) were added every 20 min. Finally, the reaction products were washed with ethanol three times by using magnetic separation for each wash.

Synthesis of Pure SILA Nanoparticles.

Silica nanoparticles (~300 nm) were synthesized by the well-established Stöber method.32 A 50 mL ammonium solution was made by mixing 9 mL of ammonium hydroxide (25–28 wt %), 16 mL of ethanol, and 25 mL of deionized H2O. This solution was swiftly added to 50 mL of 0.4 M TEOS ethanol solution under vigorous stirring. After reacting for 2 h, the particles were centrifuged and washed three times with ethanol to collect monodisperse silica nanoparticles. TEM indicated that the average particle size was approximately 300 nm.

Fabrication of Delrin Mold and Gel Cutting Tools.

Square-shaped sheets of Delrin with a thickness of 0.125 in. (0.3175 cm) were cut into the desired shapes using a laser cutter (Universal Laser Systems VLS 4.60, featuring a 60 W CO2 laser). The patterns were designed using the AutoCAD software package.

Synthesis of Responsive Photonic Film.

The as-obtained Fe3O4@SiO2 particles from the protocol (described above) were dispersed in 12 mL of ethanol, and then 3 mL of this solution of Fe3O4@SiO2 was redispersed in EG solution containing 8 mg of riboflavin, followed by adding 0.09 g of Irgacure, 225 μL of poly(ethylene glycol) diacrylate, and 0.45 g of N-isopropylacrylamide. The mixture was vortexed for 10 s to obtain a homogeneous solution which was then sonicated for 5 min in the dark. We denote this mixture as the “precursor solution” in subsequent steps. A 3 cm × 3 cm Delrin mold covered with a Petri dish was used to fabricate the responsive photonic crystal layer. For each synthesis, 350 μL of precursor solution was injected into the mold and exposed to a magnetic field of approximately 245 G (determined using a WT10A teslameter). Finally, the precursor solution was cured with a 100 W high-intensity UV lamp (365 nm, Analytik Jena US company) for 10 min. The as-obtained thin film was washed with dimethylformamide and H2O three times, respectively, then was stored in water for further use. Notably, for special shapes (fish and leaf), the Delrin molds were deliberately fabricated into the required shapes by using a laser cutter as described above using AutoCAD to draw the design.

Patterning of SASS. The as-synthesized responsive photonic film was first cut by firmly pressing a patterned Delrin grid. This was performed on a clean Petri dish surface. Prior to the fabrication of the PEG supporting layer, two precursor solutions were prepared. The first (precursor A) was generated using 750 μL of deionized water and 0.03 g of Laponite XLG, which were mixed together in a 2 mL vial using a magnetic stir bar until the solution became clear. To this Laponite XLG solution were added 750 μL of 0.1 M Na3PO4/H3PO4 buffer (pH = 7.2) and 0.12 g of tetra-PEG-NHS, and the mixture was completely dissolved through vigorous stirring and sonication. The second precursor solution (B) was prepared using 0.12 g of tetra-PEG-NHS that was dissolved in 600 μL of Na3PO4/H3PO4 buffer (pH = 7.2) under vigorous stirring and sonication. Both precursors A and B were...
stored in an ice box for 20 min before mixing in order to slow the gelation rate. After cooling precursors A and B, the solutions were mixed together for 10 s using a magnetic stir bar, then quickly poured into a specific Delrin mold (Figure S5) and sandwiched with the grid-cut photonic film that was placed on a Petri dish. The gelation process was complete within 15 min at room temperature. More details of materials and fabrication process are shown in the Supporting Information.

Characterization. TEM images were acquired with a Hitachi HT-7700 with an 80 kV accelerating voltage. SEM images were obtained using a Topcon DS-130F field emission SEM. Digital photographs and videos were recorded with an iPhone 6s. Dynamic surface temperature was monitored by an Etekcity Lasergrip 1080 non-contact digital laser infrared thermometer temperature gun (dynamic range for instrument is −50 to 550 °C). Dynamic light scattering (DLS) was performed with a NanoPlus Zeta/Nano particle analyzer at 25 °C. UV light intensity was determined by an AMTAST UVA365 UV light meter. Gel deswelling ratio was measured by a digital caliper. White LED illumination was performed using a Cygolite Metro 800 with a light intensity of 800 lm (distance to the sample: 3 cm). Solar responses were tested with natural sunlight during the daytime between the hours of 1 to 3 p.m. during the months of June and July (location: 33 °N 84°19′36.3″ W). Laser response test was conducted with a 50–80 mW 532 nm green laser module with TTL and Fan (12 V) purchased from OdićForce Lasers (Surbiton, UK). Reflectivity spectra were measured with a FLAME-S-VIS-NIR spectrometer equipped with a Premium 400 μm reflection probe (Ocean Optics Inc., Dunedin, FL, USA). All the spectral data were recorded through the OceanView 1.6.3 software package. Uniaxial tensile tests were conducted using a TestResources 100Q universal testing machine (Shakopee, MN, USA). All optical imaging was performed using a Nikon Eclipse Ti microscope, operated with Nikon Elements software, a 1.49 NA CFI Apo 100× objective, perfect focus system, and LED light source along with a laser launch excitation source. Image] was used to analyze the intensity change of the time-lapse video. Further experimental details are included in the Supporting Information.

ASSOCIATED CONTENT

Supporting Information
The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.nano.9b04231.

Estimation of strain in conventional responsive PC gels; Detailed materials and methods; finite element analysis; characterization of Fe3O4@SiO2 nanoparticles; optical microscopy imaging; calculation of photothermal efficiency of MNPs; comparison of size change of fish-shaped conventional RPC to fish-shaped SASS (PDF)

Movie S1: Time-lapse video of aligned MNPs in the responsive layer collected during photothermal heating (AVI)

Movie S2: Time-lapse video of aligned MNPs in the responsive layer collected during cooling (AVI)

Movie S3: Time-lapse video of photothermal heating in a SASS sample in which the MNPs were replaced with pure silica nanoparticles (AVI)

Movie S4: Time-lapse video of photothermal heating in a SASS sample in which the MNPs were replaced with pure silica nanoparticles (AVI)

Movie S5: Demonstration of a small piece SASS sample responding to 535 nm laser (MOV)

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Author Contributions
Y.D. and K.S., conceived of the project. Y.D. designed and performed most of the experiments. A.A. helped with the design of the Delrin mold and gel cutting tools. A.P., F.S., and W.S. helped with the uniaxial tensile measurements. J.C. and K.C. participated in discussions and revision of the paper. Y.D. and K.S. wrote the manuscript.

Notes
The authors declare no competing financial interest.

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REFERENCES


