

Physics of Transcranial Direct Current Stimulation Devices and Their History

Dennis Q. Truong, MS and Marom Bikson, PhD

Transcranial direct current stimulation (tDCS) devices apply direct current through electrodes on the scalp with the intention to modulate brain function for experimental or clinical purposes. All tDCS devices include a current controlled stimulator, electrodes that include a disposable electrolyte, and headgear to position the electrodes on the scalp. Transcranial direct current stimulation dose can be defined by the size and position of electrodes and the duration and intensity of current applied across electrodes. Electrode design and preparation are important for reproducibility and tolerability. High-definition tDCS uses smaller electrodes that can be arranged in arrays to optimize brain current flow. When intended to be used at home, tDCS devices require specific device design considerations. Computational models of current flow have been validated and support optimization and hypothesis testing. Consensus on the safety and tolerability of tDCS is protocol specific, but medical-grade tDCS devices minimize risk.

Key Words: brain stimulation, neuromodulation, review, tDCS, tES

(J ECT 2018;00: 00–00)

HISTORICAL DEVELOPMENT OF tDCS DEVICES

This history of electrical stimulation dates to the discovery of electrical phenomena, and static voltage sources are among the earliest examples of electrical technology,¹ although with unclear relation to modern transcranial direct current stimulation (tDCS) dose. There have been a continuous history of transcranial electrical stimulation technology development and testing, much of it on non-DC waveforms such as pulsed stimulation.^{2–4} Human trial investigated tDCS for neuropsychiatric disorders continued through the middle of the 20th century, typically with current intensities lower and durations longer than modern tDCS.⁵ The importance of canonical trials circa 2000 (showing tDCS is a polarity-specific modulator of brain excitability) is evidenced by these trials establishing modern tDCS dose: 1 mA applied over tens of minutes with relatively large electrodes.^{6,7} Subsequent pilot trials instituted a 2-mA intensity for therapeutic interventions^{8–10} maintained for almost all subsequent clinical evaluation.^{11–16} These developments established contemporary tDCS dose and hence the specification of modern tDCS devices (Fig. 1). Iontophoresis devices were adopted for some tDCS trials as an off-label medical device, although they may not provide a steady output.^{17,18}

Ongoing refinements in dose (eg, use of 1.5 mA in cognitive neuroscience¹⁹), electrodes (eg, HD-tDCS²⁰), integration with imaging (eg, functional magnetic resonance imaging²¹), and home use (eg, remote supervised²²) are reflected in specific tDCS device

features. Usability device features such as enhanced programming (microcontroller), control systems (eg, response to impedance changes), rechargeable batteries, disposable electrodes, enhanced headgear materials, wireless connectivity, or integration of monitoring technology²³ reflect general progress in available technologies but maintain tDCS dose.

BASICS OF tDCS DEVICES AND POLARITY

All tDCS devices include a battery-powered current-controlled stimulator that generates the stimulation waveform: a sustained direct current of several mA for up to tens of minutes, with a ramp up/down at the beginning/end. This current is applied through wires (leads) to electrodes. All tDCS devices have a minimum of 2 electrodes, with at least 1 electrode placed on the scalp. At an anode electrode, current enters the body, and at a cathode electrode, current exits the body.²⁴ There must be at least 1 anode and 1 cathode; such tDCS devices with only 2 electrodes have 1 anode and 1 cathode. When there are more than 2 electrodes, the summed current across anode electrodes must equal the summed current across the cathode electrodes²⁵; that is because the total current entering the body must equal the total current exiting the body. A majority of tDCS interventions, and thus devices, are limited to 2 mA, which historically is the highest amplitude tested (safety), but protocols, and devices, with higher (3–4 mA) current limits are explored,^{26,27} which remain within accepted safety limits.^{15,28,29}

The polarity of each electrode can be described by anode or cathode. Because an anode and cathode are always present, the terms anodal or cathodal tDCS refer to a hypothesis that neurophysiological or behavioral changes reflect stimulation of brain regions near the anode or cathode, respectively.^{30,31} Similarly, the term reference or return electrode refers to a hypothesis that brain regions near these electrodes are not central in any neurophysiological or behavioral changes.^{31,32} However, during tDCS, current passes through all brain regions between electrodes.^{20,33,34} “Extracerebral electrode” indicates a position on or below the neck, which does not cancel the effect this electrode can produce (changes in excitability) on the ventral surface of the brain and in deep brain structures.^{32,35}

tDCS ELECTRODES

The traditional electrodes used for tDCS are each made from a conductive rubber or metal plate separated from the skin by a saline-soaked sponge or paste.³¹ Note that in electrochemistry the conductive rubber or plate would be the electrode, whereas the saline, gel, or paste would be the electrolyte,²⁴ but in tDCS literature, the entire assembly is called the electrode. Therefore, in tDCS, when electrode size is described (eg, $5 \times 5 \text{ cm}^2$), it is the interface between the skin and the electrolyte. Nonetheless, the configuration of all electrode component dimensions and materials is important to control and document as this affects tolerability.^{31,36–39} The thickness of the sponge or paste effectively controls the minimum distance between the conductible rubber or metal and the skin. Contact of conductive rubber or metal with skin during tDCS is avoided as this decreases tolerability and introduces risk of lasting skin irritation.

From the Department of Biomedical Engineering, The City College of New York of CUNY, New York, NY.

Received for publication March 10, 2018; accepted June 15, 2018.

Reprints: Dennis Q. Truong, MS, Center for Discovery and Innovation,

85 Saint Nicholas Ter, CDI 3366, New York, NY 10031-1246

(e-mail: dtruong@ccny.cuny.edu).

The City University of New York has patents on brain stimulation with M.B. as inventor. M.B. has equity in Soterix Medical Inc and serves as a scientific advisor to Boston Scientific Inc. D.Q.T. has no conflicts of interest or financial disclosures to report.

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

DOI: 10.1097/YCT.0000000000000531

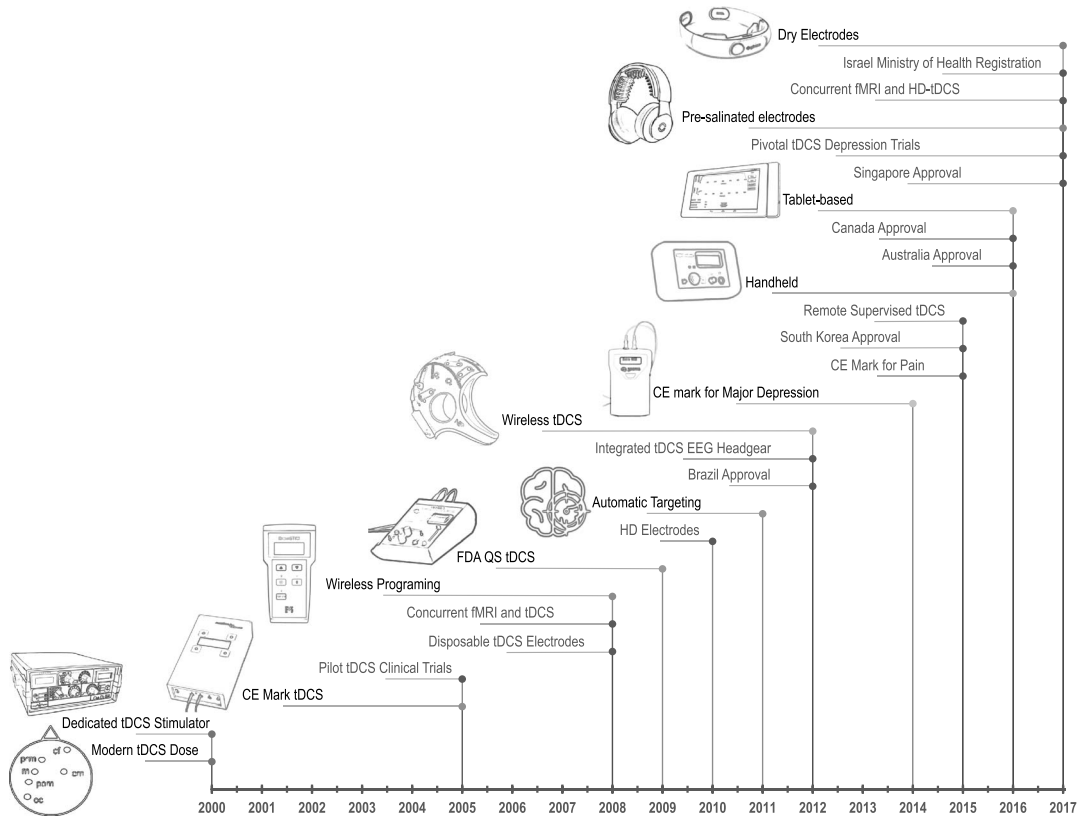


FIGURE 1. Timeline of tDCS innovations: technology and regulatory milestones. CE mark indicates Conformité Européenne Marking; FDA QS, Food and Drug Administration Quality Systems; fMRI, functional magnetic resonance imaging; HD, high definition/density.

Single-use electrodes are advantageous. In any case, the electrolyte is not reused (as it dehydrates, its properties will change). Electrodes typically positioned based on the electroencephalography (EEG) 10/10 system (eg, anode of F3) through customized placement based on neuronavigated,^{40–43} functional,⁴⁴ nonneuronavigated,⁴⁵ or image-based approaches (eg, EEG reciprocity^{23,46–48}) have been developed. Either the headgear is designed to support the determination electrodes positions (eg, a cap or marked straps^{49,50}), or the headgear is generic (eg, rubber bands⁵¹), and independent measurement is used to position the electrodes.

HD-tDCS ELECTRODES AND MONTAGES

In transcranial electrical stimulation, smaller electrodes are called high-definition (HD) electrodes⁵² and typically are made from small circular Ag/AgCl electrode separated from the skin by a gel contained in a plastic cylinder.⁵³ The plastic cylinder controls the distance between the electrode and the skin. Because they are smaller, use of HD electrodes for tDCS allows more precision in electrode position⁵⁴ and the option to use more electrodes.^{25,55–57}

The 4 × 1 HD-tDCS montage uses 1 center electrode surrounded by 4 electrodes (Fig. 2) of the opposite polarity, with the intention to target cortical regions.^{20,58–63} Bipolar HD-tDCS montages (1 anode and 1 cathode) when electrodes are positioned closely can be used to probe the role of current direction across the cortex⁵⁴ and when electrodes are positioned across the head to maximize brain current flow.²⁵ Increasing the number of HD electrodes can support multifocal stimulation.^{56,57,64–67}

tDCS DOSE AND CURRENT FLOW MODELING

Electrode size and position on the scalp along with the current applied to each electrode conventionally define tDCS dose.⁶⁸

Transcranial direct current stimulation dose, along with head anatomy, determines the resulting current flow (intensity and pattern) in the brain and in turn affects resulting neurophysiological and behavioral changes.⁶⁹ Indeed, the canonical studies establishing the neuromodulation actions of tDCS did so by showing dose (electrode montage)–specific effects.⁶ Yet, systematic ongoing studies have characterized additional factors beyond electrode montage such as brain state,^{70,71} interindividual differences,^{72–74,74,75} and nonmonotonic (eg, “more is not always better”^{76–78}) dose response. This complexity of tDCS dose-response is in line with other forms of brain stimulation,^{79–81} whereas the sensitivity to brain state is consistent with hypothesized mechanisms of actions.⁸² Notably, in contrast to other common forms of clinical brain stimulation (repetitive transcranial magnetic stimulation, electroconvulsive therapy, deep brain stimulation, etc), tDCS is not typically titrated on subject-specific basis; ongoing research on methods to individualize tDCS dose is warranted.^{46,83–85}

For a given dose and anatomy, computational models predict the resulting current flow (electric field distribution) in the brain (Fig. 2). Computational models have been developed^{20,25,34,57,86–89} and repeatedly validated^{33,90–93} over a decade. It is important not to conflate established montage-specific effects (eg, “shaping” the outcomes of stimulation⁹⁴) with demonstration of focality (eg, current delivery to 1 region of interest). Rather, models of conventional tDCS and HD-tDCS support testing hypothesis linking brain regions to neurophysiologic or behavioral changes.⁹⁵ This includes registering results from current flow models with imaging data.⁹⁶

tDCS BIOPHYSICS

Although there are open questions about the mechanisms and efficacy of tDCS for varied indications and the biophysics

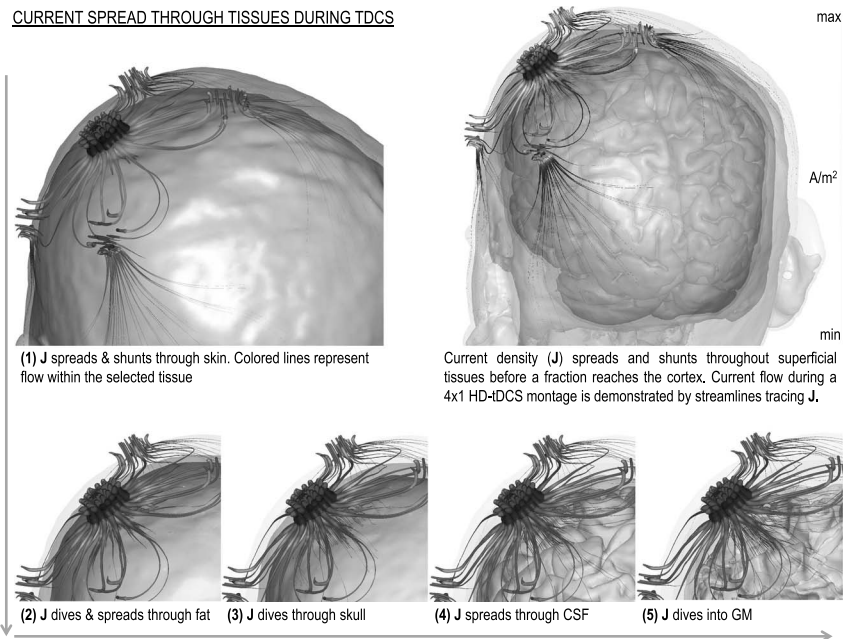


FIGURE 2. Illustration of current spreading and shunting through superficial tissues. Streamlines seeded under the electrodes trace current density as it loops from anode to cathode. Line diameter is logarithmic with intensity. Insets (1) through (5) isolate and desaturate current outside the tissue of interest. Only a fraction of the current delivered during tDCS reaches the cortex. In the 4×1 HD-tDCS example, the center anode and surround cathodes (or vice versa) are in close proximity on the scalp surface. Much of the current shunts through skin but a fraction of the current spreads into deeper tissues and eventually into the cortex.

of tDCS related to current delivery to the brain, the resulting polarization of neuronal membranes is well established.^{97,98} Current that is passed through tDCS electrodes takes a path through the head determined by the head anatomy and the resistivity of each tissue type. A fraction of the current never crosses the resistive cranium, instead shunting across the relatively-conductive (low resistivity) scalp.⁹¹ Of the current fraction that crosses the skull, a further portion is shunted by the high-conductivity cerebrospinal fluid. The current component that reaches the brain crosses the gray and then white matter. As current crosses brain tissue, it generates an electric field on the local tissue. Neurons are exposed to and so stimulated by local electric field. The current intensity is not uniform across the brain, and so the electric field intensity is also distributed. The peak electric field in the brain during 2-mA tDCS is 0.5 to 1 V/m based on intracranial recording in subjects validating current flow models.^{33,92,93} For conventional tDCS, this peak may be in a brain region between electrodes.

The direction of current flow across the gray matter can be radial inward (from the pial surface toward gray/white matter boundary), radial outward, or tangential (along the gray matter).⁹⁹ Current flow will polarize neuron in a compartment-specific manner (ie, the soma, dendrites, axon of a single neuron may be polarized differently^{100,101}). The magnitude and direction of the electric field generated in the gray matter determine the polarization of neuronal compartments.¹⁰² Radial inward current will depolarize the somas of cortical pyramidal neurons ~ 0.2 mV per V/m of electric field, whereas radial outward current will hyperpolarize the cortical pyramidal neurons somas by -0.2 mV per V/m.¹⁰³ Radial inward/outward current is expected to increase/decrease the firing rate of these neurons because of somatic polarization.^{104,105} Each neuronal compartment will be polarized, depending on the morphology of the neuron.^{106,107} Electric field will polarize axon terminals (synapses) oriented parallel to the field direction ~ 1 mV per V/m,¹⁰⁸ which can then influence synaptic function.^{107,109–111}

The neurophysiological and so behavioral consequences of tDCS will depend on how this polarization influences excitability and plasticity.¹¹² Because tDCS produces only incremental membrane polarization, the cellular effects of tDCS on brain function will depend on ongoing activity.^{82,113–115} These effects may then be amplified over time (tens of minutes^{116–118}). The organization of neurons in active networks with emergent properties such as oscillations will influence the aggregate effects of tDCS.^{114,119} The ultimate consequences of tDCS on macroscopic measures of neurophysiology (eg, transcranial magnetic stimulation) and behavior (eg, therapy) will be complex,^{54,120–123} but ongoing research about such changes should not be confused with debate about the biophysics of current flow and resulting membrane polarization.

SAFETY AND TOLERABILITY OF tDCS DEVICES

The tolerability of any intervention depends not simply on the device and dose, but on protocol including the subject's demographic and clinical characteristics (ie, inclusion/exclusion criteria [eg, age, preexisting condition], operator training and certification, ongoing monitoring, and parallel interventions). Therefore, the scientific consensus that tDCS is safe and tolerated^{12,15,31,124–126} is explicitly limited to those protocols tested. Human trials of tDCS in the United States are almost always considered nonsignificant risk (risk comparable to daily activities). But this risk designation—whether made by the Food and Drug Administration or by an institutional review board—must be made on a protocol-specific basis, emphasizing that recommendation on safety and tolerability cannot be made on any device, but must also specify the methods of use.

Transcranial direct current stimulation device design may be considered to minimize risk to the extent they reliably control dose and allow consistent electrode setup, when used within the limits of established protocols. Medical-grade tDCS devices and accessories, which are designed and manufactured to internationally recognized medical standards, regardless of region specific approval for

treatment,^{12,28,127} provide the highest standard of control in regard to reliability.

HOME-BASED tDCS DEVICES

A theoretical advantage of tDCS is deployability. Factors such as cost, portability, safety, and ease of use allow tDCS to be used in a wide range of clinical environments and at home.¹²⁸ However, devices designed for use by certified operators at research or clinical centers may not be suitable across deployed conditions. To address this concern, standards for remote-supervised tDCS have been developed¹²⁹ and validated.^{129,130} The principle of remote-supervised tDCS is, under continuous medical or research supervision, to control compliance, proper dose control, and risk. Features of suitable device include mechanisms to limit dose (eg, one 2-mA, 20-minute session per day) and simple and robust method to prepare and apply electrodes (eg, single-use presaturated snap electrodes and single-position headgear). While the ethics and merits of self-administered tDCS (outside medical or research supervision) are discussed,^{131–133} specifications for tDCS devices that minimize risk have been developed.²⁸

REFERENCES

1. Paulus W, Opitz A. Ohm's law and tDCS over the centuries. *Clin Neurophysiol.* 2013;124:429–430.
2. Guleyupoglu B, Schestatsky P, Edwards D, et al. Classification of methods in transcranial electrical stimulation (tES) and evolving strategy from historical approaches to contemporary innovations. *J Neurosci Methods.* 2013;219:297–311.
3. Steinberg H. Letter to the editor: transcranial direct current stimulation (tDCS) has a history reaching back to the 19th century. *Psychol Med.* 2013;43:669–671.
4. Wexler A. Recurrent themes in the history of the home use of electrical stimulation: transcranial direct current stimulation (tDCS) and the medical battery (1870–1920). *Brain Stimul.* 2017;10:187–195.
5. Esmailpour Z, Schestatsky P, Bikson M, et al. Notes on human trials of transcranial direct current stimulation between 1960 and 1998. *Front Hum Neurosci.* 2017;11:71.
6. Nitsche MA, Paulus W. Excitability changes induced in the human motor cortex by weak transcranial direct current stimulation. *J Physiol.* 2000;527:633–639.
7. Nitsche MA, Paulus W. Sustained excitability elevations induced by transcranial DC motor cortex stimulation in humans. *Neurology.* 2001;57:1899–1901.
8. Fregni F, Boggio PS, Mansur CG, et al. Transcranial direct current stimulation of the unaffected hemisphere in stroke patients. *Neuroreport.* 2005;16:1551–1555.
9. Fregni F, Boggio PS, Nitsche MA, et al. Treatment of major depression with transcranial direct current stimulation. *Bipolar Disord.* 2006;8:203–204.
10. Fregni F, Boggio PS, Santos MC, et al. Noninvasive cortical stimulation with transcranial direct current stimulation in Parkinson's disease. *Mov Disord.* 2006;21:1693–1702.
11. Brunoni AR, Moffa AH, Sampaio-Junior B, et al. Trial of electrical direct-current therapy versus escitalopram for depression. *N Engl J Med.* 2017;376:2523–2533.
12. Antal A, Alekseiuk I, Bikson M, et al. Low intensity transcranial electric stimulation: safety, ethical, legal regulatory and application guidelines. *Clin Neurophysiol.* 2017;128:1774–1809.
13. Nitsche MA, Cohen LG, Wassermann EM, et al. Transcranial direct current stimulation: state of the art 2008. *Brain Stimul.* 2008;1:206–223.
14. Lefaucheur J-P, Antal A, Ayache SS, et al. Evidence-based guidelines on the therapeutic use of transcranial direct current stimulation (tDCS). *Clin Neurophysiol Off J Int Fed Clin Neurophysiol.* 2017;128:56–92.

15. Bikson M, Grossman P, Thomas C, et al. Safety of transcranial direct current stimulation: evidence based update 2016. *Brain Stimul.* 2016;9:641–661.
16. Fregni F, Boggio PS, Lima MC, et al. A sham-controlled, phase II trial of transcranial direct current stimulation for the treatment of central pain in traumatic spinal cord injury. *Pain.* 2006;122:197–209.
17. Chhatbar PY, Sawers JR, Feng W. The proof is in the pudding: does tDCS actually deliver DC stimulation? *Brain Stimulat.* 2016;9:625–626.
18. Salimpour Y, Wei Z, Duy PQ, et al. Does transcranial direct current stimulation actually deliver DC stimulation? *Brain Stimul.* 2016;9:623–624.
19. Turkeltaub PE, Benson J, Hamilton RH, et al. Left lateralizing transcranial direct current stimulation improves reading efficiency. *Brain Stimul.* 2012;5:201–207.
20. Datta A, Bansal V, Diaz J, et al. Gyri-precise head model of transcranial Director current stimulation: improved spatial focality using a ring electrode versus conventional rectangular pad. *Brain Stimul.* 2009;2:201–207. 207.e1.
21. Antal A, Polania R, Schmidt-Samoa C, et al. Transcranial direct current stimulation over the primary motor cortex during fMRI. *Neuroimage.* 2011;55:590–596.
22. Charvet LE, Dobbs B, Shaw MT, et al. Remotely supervised transcranial direct current stimulation for the treatment of fatigue in multiple sclerosis: results from a randomized, sham-controlled trial. *Mult Scler.* 2017;1:1352458517732842.
23. Leite J, Morales-Quezada L, Carvalho S, et al. Surface EEG-transcranial direct current stimulation (tDCS) closed-loop system. *Int J Neural Syst.* 2017;27:1750026.
24. Merrill DR, Bikson M, Jefferys JG. Electrical stimulation of excitable tissue: design of efficacious and safe protocols. *J Neurosci Methods.* 2005;141:171–198.
25. Dmochowski JP, Datta A, Bikson M, et al. Optimized multi-electrode stimulation increases focality and intensity at target. *J Neural Eng.* 2011;8:046011.
26. Reckow J, Rahman-Filipiak A, Garcia S, et al. Tolerability and blinding of 4x1 high-definition transcranial direct current stimulation (HD-tDCS) at two and three milliamps. *Brain Stimul.* 2018.
27. Chhatbar PY, Chen R, Deardorff R, et al. Safety and tolerability of transcranial direct current stimulation to stroke patients—a phase I current escalation study. *Brain Stimul.* 2017;10:553–559.
28. Bikson M, Paneri B, Mourdoukoutas A, et al. Limited output transcranial electrical stimulation (LOTES-2017): engineering principles, regulatory statutes, and industry standards for wellness, over-the-counter, or prescription devices with low risk. *Brain Stimulat.* 2018;11:134–157.
29. Jackson MP, Truong D, Brownlow ML, et al. Safety parameter considerations of anodal transcranial direct current stimulation in rats. *Brain Behav Immun.* 2017;64:152–161.
30. Garnett EO, Malyutina S, Datta A, et al. On the use of the terms anodal and cathodal in high-definition transcranial direct current stimulation: a technical note. *Neuromodulation.* 2015;18:705–713.
31. Woods AJ, Antal A, Bikson M, et al. A technical guide to tDCS, and related non-invasive brain stimulation tools. *Clin Neurophysiol.* 2016;127:1031–1048.
32. Bikson M, Datta A, Rahman A, et al. Electrode montages for tDCS and weak transcranial electrical stimulation: role of “return” electrode's position and size. *Clin Neurophysiol.* 2010;121:1976–1978.
33. Huang Y, Liu AA, Lafon B, et al. Measurements and models of electric fields in the in vivo human brain during transcranial electric stimulation. *Elife.* 2017;6:e18834.
34. Opitz A, Paulus W, Will S, et al. Determinants of the electric field during transcranial direct current stimulation. *Neuroimage.* 2015;109:140–150.

35. Noetscher GM, Yanamadala J, Makarov SN, et al. Comparison of cephalic and extracephalic montages for transcranial direct current stimulation—a numerical study. *IEEE Trans Biomed Eng.* 2014;61:2488–2498.
36. Kronberg G, Bikson M. Electrode assembly design for transcranial direct current stimulation: a FEM modeling study. In: *2012 Conf Proc IEEE Eng Med Biol Soc*; San Diego, CA, 28 Aug.–1 Sept. 2012; 2012:891–895.
37. Dundas JE, Thickbroom GW, Mastaglia FL. Perception of comfort during transcranial DC stimulation: effect of NaCl solution concentration applied to sponge electrodes. *Clin Neurophysiol.* 2007;118:1166–1170.
38. Minhas P, Datta A, Bikson M. Cutaneous perception during tDCS: role of electrode shape and sponge salinity. *Clin Neurophysiol.* 2011;122:637–638.
39. Turi Z, Ambrus GG, Ho K-A, et al. When size matters: large electrodes induce greater stimulation-related cutaneous discomfort than smaller electrodes at equivalent current density. *Brain Stimul.* 2014;7:460–467.
40. Teichmann M, Lesoil C, Godard J, et al. Direct current stimulation over the anterior temporal areas boosts semantic processing in primary progressive aphasia. *Ann Neurol.* 2016;80:693–707.
41. Parazzini M, Fiocchi S, Cancelli A, et al. A computational model of the electric field distribution due to regional personalized or nonpersonalized electrodes to select transcranial electric stimulation target. *IEEE Trans Biomed Eng.* 2017;64:184–195.
42. Richardson J, Datta A, Dmochowski J, et al. Feasibility of using high-definition transcranial direct current stimulation (HD-tDCS) to enhance treatment outcomes in persons with aphasia. *NeuroRehabilitation.* 2015;36:115–126.
43. Witte S, Klooster D, Dedoncker J, Duprat R, Remue J, Baeken C. Left prefrontal neuronavigated electrode localization in tDCS: 10–20 EEG system versus MRI-guided neuronavigation. *Psychiatry Res.* 2018.
44. Rich TL, Menk JS, Rudser KD, et al. Determining electrode placement for transcranial direct current stimulation: a comparison of EEG- versus TMS-guided methods. *Clin EEG Neurosci.* 2017;48:367–375.
45. Seibt O, Brunoni AR, Huang Y, et al. The pursuit of DLPCF: non-neuronavigated methods to target the left dorsolateral pre-frontal cortex with symmetric bicephalic transcranial direct current stimulation (tDCS). *Brain Stimulat.* 2015;8:590–602.
46. Dmochowski JP, Koessler L, Norcia AM, et al. Optimal use of EEG recordings to target active brain areas with transcranial electrical stimulation. *Neuroimage.* 2017;157:69–80.
47. Fernández-Corazza M, Turovets S, Luu P, et al. Transcranial electrical neuromodulation based on the reciprocity principle. *Neuroimaging Stimul.* 2016;7:1–19.
48. Wagner S, Lucka F, Vorwerk J, et al. Using reciprocity for relating the simulation of transcranial current stimulation to the EEG forward problem. *Neuroimage.* 2016;140:163–173.
49. Schestatsky P, Morales-Quezada L, Fregni F. Simultaneous EEG monitoring during transcranial direct current stimulation. *J Vis Exp.* 2013; 76:1–11.
50. Kasschau M, Sherman K, Haider L, et al. A protocol for the use of remotely-supervised transcranial direct current stimulation (tDCS) in multiple sclerosis (MS). *J Vis Exp.* 2015;e53542. DOI: 10.3791/53542.
51. DaSilva AF, Volz MS, Bikson M, et al. Electrode positioning and montage in transcranial direct current stimulation. *J Vis Exp.* 2011;51:1–11.
52. Minhas P, Bansal V, Patel J, et al. Electrodes for high-definition transcutaneous DC stimulation for applications in drug delivery and electrotherapy, including tDCS. *J Neurosci Methods.* 2010;190:188–197.
53. Villamar MF, Volz MS, Bikson M, et al. Technique and considerations in the use of 4×1 ring high-definition transcranial direct current stimulation (HD-tDCS). *J Vis Exp.* 2013;e50309.
54. Rawji V, Ciocca M, Zacharia A, et al. tDCS changes in motor excitability are specific to orientation of current flow. *Brain Stimul.* 2018; 11:289–298.
55. Guler S, Dannhauer M, Erem B, et al. Optimization of focality and direction in dense electrode array transcranial direct current stimulation (tDCS). *J Neural Eng.* 2016;13:036020.
56. Ota B, Dutta A, Foerster Á, et al. Opportunities for guided multichannel non-invasive transcranial current stimulation in poststroke rehabilitation. *Front Neurol.* 2016;7:21.
57. Ruffini G, Fox MD, Rippolles O, et al. Optimization of multifocal transcranial current stimulation for weighted cortical pattern targeting from realistic modeling of electric fields. *Neuroimage.* 2014;89:216–225.
58. Kuo H-I, Bikson M, Datta A, et al. Comparing cortical plasticity induced by conventional and high-definition 4 × 1 ring tDCS: a neurophysiological study. *Brain Stimulat.* 2013;6:644–648.
59. Alam M, Truong DQ, Khadka N, et al. Spatial and polarity precision of concentric high-definition transcranial direct current stimulation (HD-tDCS). *Phys Med Biol.* 2016;61:4506.
60. Guo H, Zhang Z, Da S, et al. High-definition transcranial direct current stimulation (HD-tDCS) of left dorsolateral prefrontal cortex affects performance in Balloon Analogue Risk Task (BART). *Brain Behav.* 2018;8:e00884.
61. Cabibel V, Muthalib MM, Teo WP, et al. High-definition transcranial direct-current stimulation of the right M1 further facilitates left M1 excitability during crossed-facilitation. *J Neurophysiol.* 2018;119:1266–1272.
62. Chua EF, Ahmed R, Garcia SM. Effects of HD-tDCS on memory and metamemory for general knowledge questions that vary by difficulty. *Brain Stimul.* 2017;10:231–241.
63. Shen B, Yin Y, Wang J, et al. High-definition tDCS alters impulsivity in a baseline-dependent manner. *Neuroimage.* 2016;143:343–352.
64. Turski CA, Kessler-Jones A, Chow C, et al. Extended multiple-field high-definition transcranial direct current stimulation (HD-tDCS) is well tolerated and safe in healthy adults. *Restor Neurol Neurosci.* 2017;35:631–642.
65. Pixa NH, Steinberg F, Doppelmayr M. Effects of high-definition anodal transcranial direct current stimulation applied simultaneously to both primary motor cortices on bimanual sensorimotor performance. *Front Behav Neurosci.* 2017;11:130.
66. Pixa NH, Steinberg F, Doppelmayr M. High-definition transcranial direct current stimulation to both primary motor cortices improves unimanual and bimanual dexterity. *Neurosci Lett.* 2017;643:84–88.
67. Fischer DB, Fried PJ, Ruffini G, et al. Multifocal tDCS targeting the resting state motor network increases cortical excitability beyond traditional tDCS targeting unilateral motor cortex. *Neuroimage.* 2017; 157:34–44.
68. Peterchev AV, Wagner TA, Miranda PC, et al. Fundamentals of transcranial electric and magnetic stimulation dose: definition, selection, and reporting practices. *Brain Stimulat.* 2012;5:435–453.
69. Ho K-A, Taylor JL, Chew T, et al. The effect of transcranial direct current stimulation (tDCS) electrode size and current intensity on motor cortical excitability: evidence from single and repeated sessions. *Brain Stimul.* 2016;9:1–7.
70. Huang Y-Z, Lu M-K, Antal A, et al. Plasticity induced by non-invasive transcranial brain stimulation: a position paper. *Clin Neurophysiol.* 2017;128:2318–2329.
71. Brunoni AR, Nitsche MA, Bolognini N, et al. Clinical research with transcranial direct current stimulation (tDCS): challenges and future directions. *Brain Stimul.* 2012;5:175–195.
72. Labruna L, Jamil A, Fresnoza S, et al. Efficacy of anodal transcranial direct current stimulation is related to sensitivity to transcranial magnetic stimulation. *Brain Stimul.* 2016;9:8–15.
73. Datta A, Truong D, Minhas P, et al. Inter-individual variation during transcranial direct current stimulation and normalization of dose using MRI-derived computational models. *Front Psychiatry.* 2012;3:91.

74. Li LM, Uehara K, Hanakawa T. The contribution of interindividual factors to variability of response in transcranial direct current stimulation studies. *Front Cell Neurosci.* 2015;9:181.

75. Wiethoff S, Hamada M, Rothwell JC. Variability in response to transcranial direct current stimulation of the motor cortex. *Brain Stimul.* 2014;7:468–475.

76. Batsikadze G, Moliadze V, Paulus W, et al. Partially non-linear stimulation intensity-dependent effects of direct current stimulation on motor cortex excitability in humans. *J Physiol.* 2013;591:1987–2000.

77. Esmaeilpour Z, Marangolo P, Hampstead BM, et al. Incomplete evidence that increasing current intensity of tDCS boosts outcomes. *Brain Stimul.* 2018;11:310–321.

78. Giordano J, Bikson M, Kappenman ES, et al. Mechanisms and effects of transcranial direct current stimulation. *Dose Response.* 2017;15:59325816685467.

79. D'Ostilio K, Goetz SM, Hannah R, et al. Effect of coil orientation on strength-duration time constant and I-wave activation with controllable pulse parameter transcranial magnetic stimulation. *Clin Neurophysiol.* 2016;127:675–683.

80. Lujan JL, Chaturvedi A, McIntyre CC. Tracking the mechanisms of deep brain stimulation for neuropsychiatric disorders. *Front Biosci.* 2008;13:5892–5904.

81. Arle JE, Mei L, Carlson KW, et al. High-frequency stimulation of dorsal column axons: potential underlying mechanism of paresthesia-free neuropathic pain relief. *Neuromodulation.* 2016;19:385–397.

82. Bikson M, Name A, Rahman A. Origins of specificity during tDCS: anatomical, activity-selective, and input-bias mechanisms. *Front Hum Neurosci.* 2013;7:688.

83. Shah-Basak PP, Norise C, Garcia G, et al. Individualized treatment with transcranial direct current stimulation in patients with chronic non-fluent aphasia due to stroke. *Front Hum Neurosci.* 2015;9:201.

84. Gillick BT, Kirton A, Carmel JB, et al. Pediatric stroke and transcranial direct current stimulation: methods for rational individualized dose optimization. *Front Hum Neurosci.* 2014;8:739.

85. Bikson M, Rahman A, Datta A, et al. High-resolution modeling assisted design of customized and individualized transcranial direct current stimulation protocols. *Neuromodulation.* 2012;15:306–315.

86. Miranda PC, Lomarev M, Hallett M. Modeling the current distribution during transcranial direct current stimulation. *Clin Neurophysiol.* 2006;117:1623–1629.

87. Wagner T, Fregni F, Fecteau S, et al. Transcranial direct current stimulation: a computer-based human model study. *Neuroimage.* 2007;35:1113–1124.

88. Im CH, Jung HH, Choi JD, et al. Determination of optimal electrode positions for transcranial direct current stimulation (tDCS). *Phys Med Biol.* 2008;53:N219–N225.

89. Truong DQ, Hüber M, Xie X, et al. Clinician accessible tools for GUI computational models of transcranial electrical stimulation: BONSAI and SPHERES. *Brain Stimul.* 2014;7:521–524.

90. Antal A, Bikson M, Datta A, et al. Imaging artifacts induced by electrical stimulation during conventional fMRI of the brain. *Neuroimage.* 2014;85(Pt 3):1040–1047.

91. Datta A, Zhou X, Su Y, et al. Validation of finite element model of transcranial electrical stimulation using scalp potentials: implications for clinical dose. *J Neural Eng.* 2013;10:036018.

92. Opitz A, Falchier A, Yan C-G, et al. Spatiotemporal structure of intracranial electric fields induced by transcranial electric stimulation in humans and nonhuman primates. *Sci Rep.* 2016;6:srep31236. doi: 10.1038/srep31236.

93. Datta A, Krause MR, Pilly PK, et al. On comparing in vivo intracranial recordings in non-human primates to predictions of optimized transcranial electrical stimulation. *Conf Proc IEEE Eng Med Biol Soc.* 2016;2016:1774–1777.

94. Nitsche MA, Doemkes S, Karaköse T, et al. Shaping the effects of transcranial direct current stimulation of the human motor cortex. *J Neurophysiol.* 2007;97:3109–3117.

95. Bikson M, Brunoni AR, Charvet LE, et al. Rigor and reproducibility in research with transcranial electrical stimulation: an NIMH-sponsored workshop. *Brain Stimul.*

96. Halko MA, Datta A, Plow EB, et al. Neuroplastic changes following rehabilitative training correlate with regional electrical field induced with tDCS. *Neuroimage.* 2011;57:885–891.

97. Miranda PC. Physics of effects of transcranial brain stimulation. *Handb Clin Neurol.* 2013;116:353–366.

98. Rahman A, Lafon B, Bikson M. Multilevel computational models for predicting the cellular effects of noninvasive brain stimulation. *Prog Brain Res.* 2015;222:25–40.

99. Datta A, Elwassif M, Battaglia F, et al. Transcranial current stimulation focality using disc and ring electrode configurations: FEM analysis. *J Neural Eng.* 2008;5:163.

100. Ranck JB. Which elements are excited in electrical stimulation of mammalian central nervous system: a review. *Brain Res.* 1975;98:417–440.

101. Tranchina D, Nicholson C. A model for the polarization of neurons by extrinsically applied electric fields. *Biophys J.* 1986;50:1139–1156.

102. Rahman A, Reato D, Arloti M, et al. Cellular effects of acute direct current stimulation: somatic and synaptic terminal effects. *J Physiol.* 2013;591:2563–2578.

103. Radman T, Ramos RL, Brumberg JC, et al. Role of cortical cell type and morphology in subthreshold and suprathreshold uniform electric field stimulation in vitro. *Brain Stimul.* 2009;2:215–228. 228 e1–3.

104. Creutzfeldt OD, Fromm GH, Kapp H. Influence of transcortical D-C currents on cortical neuronal activity. *Exp Neurol.* 1962;5:436–452.

105. Radman T, Su Y, An JH, et al. Spike timing amplifies the effect of electric fields on neurons: implications for endogenous field effects. *J Neurosci.* 2007;27:3030–3036.

106. Chan CY, Hounsgaard J, Nicholson C. Effects of electric fields on transmembrane potential and excitability of turtle cerebellar Purkinje cells in vitro. *J Physiol.* 1988;402:751–771.

107. Bikson M, Inoue M, Akiyama H, et al. Effects of uniform extracellular DC electric fields on excitability in rat hippocampal slices in vitro. *J Physiol.* 2004;557(pt 1):175–190.

108. Chakraborty D, Truong DQ, Bikson M, et al. Neuromodulation of axon terminals. *Cereb Cortex.* 2017;1991:1–9.

109. Márquez-Ruiz J, Leal-Campanario R, Sánchez-Campusano R, et al. Transcranial direct-current stimulation modulates synaptic mechanisms involved in associative learning in behaving rabbits. *Proc Natl Acad Sci U S A.* 2012;109:6710–6715.

110. Kabakov AY, Muller PA, Pascual-Leone A, et al. Contribution of axonal orientation to pathway-dependent modulation of excitatory transmission by direct current stimulation in isolated rat hippocampus. *J Neurophysiol.* 2012;107:1881–1889.

111. Fritsch B, Reis J, Martinowich K, et al. Direct current stimulation promotes BDNF-dependent synaptic plasticity: potential implications for motor learning. *Neuron.* 2010;66:198–204.

112. Jackson MP, Rahman A, Lafon B, et al. Animal models of transcranial direct current stimulation: methods and mechanisms. *Clin Neurophysiol.* 2016;127:3425–3454.

113. Rahman A, Lafon B, Parra LC, et al. Direct current stimulation boosts synaptic gain and cooperativity in vitro. *J Physiol.* 2017;595:3535–3547.

114. Reato D, Rahman A, Bikson M, et al. Effects of weak transcranial alternating current stimulation on brain activity—a review of known mechanisms from animal studies. *Front Hum Neurosci.* 2013;7:687.

115. Krause MR, Zanos TP, Csorba BA, et al. Transcranial direct current stimulation facilitates associative learning and alters functional connectivity in the primate brain. *Curr Biol*. 2017;27:3086.e3–3096.e3.
116. Bindman LJ, Lippold OC, Redfean JW. Long-lasting changes in the level of the electrical activity of the cerebral cortex produced by polarizing currents. *Nature*. 1962;196:584–585.
117. Pelletier SJ, Cicchetti F. Cellular and molecular mechanisms of action of transcranial direct current stimulation: evidence from in vitro and in vivo models. *Int J Neuropsychopharmacol*. 2014;18.
118. Reato D, Bikson M, Parra LC. Lasting modulation of in vitro oscillatory activity with weak direct current stimulation. *J Neurophysiol*. 2015;113:1334–1341.
119. Schmidt SL, Iyengar AK, Foulser AA, et al. Endogenous cortical oscillations constrain neuromodulation by weak electric fields. *Brain Stimul*. 2014;7:878–889.
120. Polanía R, Nitsche MA, Paulus W. Modulating functional connectivity patterns and topological functional organization of the human brain with transcranial direct current stimulation. *Hum Brain Mapp*. 2011;32:1236–1249.
121. Filmer HL, Dux PE, Mattingley JB. Applications of transcranial direct current stimulation for understanding brain function. *Trends Neurosci*. 2014;37:742–753.
122. Vöröslakos M, Takeuchi Y, Brinyiczki K, et al. Direct effects of transcranial electric stimulation on brain circuits in rats and humans. *Nat Commun*. 2018;9:483.
123. Antal A, Varga ET, Kincses TZ, et al. Oscillatory brain activity and transcranial direct current stimulation in humans. *Neuroreport*. 2004;15:1307–1310.
124. Poreisz C, Boros K, Antal A, et al. Safety aspects of transcranial direct current stimulation concerning healthy subjects and patients. *Brain Res Bull*. 2007;72:208–214.
125. Nikolin S, Huggins C, Martin D, et al. Safety of repeated sessions of transcranial direct current stimulation: a systematic review. *Brain Stimul*. 2018;11:278–288.
126. Aparicio LVM, Guarienti F, Razza LB, et al. A systematic review on the acceptability and tolerability of transcranial direct current stimulation treatment in neuropsychiatry trials. *Brain Stimul*. 2016;9:671–681.
127. Fregni F, Nitsche MA, Loo CK, et al. Regulatory considerations for the clinical and research use of transcranial direct current stimulation (tDCS): review and recommendations from an expert panel. *Clin Res Regul Aff*. 2015;32:22–35.
128. Palm U, Kumpf U, Behler N, et al. Home use, remotely supervised, and remotely controlled transcranial direct current stimulation: a systematic review of the available evidence. *Neuromodulation*. 2018;21:323–333.
129. Charvet LE, Kasschau M, Datta A, et al. Remotely-supervised transcranial direct current stimulation (tDCS) for clinical trials: guidelines for technology and protocols. *Front Syst Neurosci*. 2015;9:26.
130. Shaw MT, Kasschau M, Dobbs B, et al. Remotely supervised transcranial direct current stimulation: an update on safety and tolerability. *J Vis Exp*. 2017.
131. Wexler A. The social context of “do-it-yourself” brain stimulation: neurohackers, biohackers, and lifehackers. *Front Hum Neurosci*. 2017;11:224.
132. Bikson M, Paneri B, Giordano J. The off-label use, utility and potential value of tDCS in the clinical care of particular neuropsychiatric conditions. *J Law Biosci*. 2016;3:642–646.
133. Wagner K, Maslen H, Oakley J, et al. Would you be willing to zap your child's brain? Public perspectives on parental responsibilities and the ethics of enhancing children with transcranial direct current stimulation. *AJOB Empir Bioeth*. 2018;9:29–38.