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Insights on the Working Principles of Secondary Electrospray Ionization High-Resolution Mass Spectrometry for Quantitative Analysis of Aerosol Chemical Composition

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Abstract

Real-time mass spectrometry (MS) has attracted increasing interest in environmental analysis due to its advantages in high time resolution, minimization of sampling artifact, and avoidance of time-consuming sample pretreatment. Among real-time MS methods, secondary electrospray ionization MS (SESI-MS) is showing great promise for the detection of organic compounds in atmospheric particulate matter. In this study, we demonstrated the working principles of secondary nanoelectrospray ionization (Sec-nESI) for real-time measurement of laboratory-generated organic aerosols using L-tartaric acid (TA) as a model compound. Factors affecting the detection of TA particles using a homemade Sec-nESI source coupled with a high-resolution mass spectrometer are systematically investigated. Temperature of ion transport capillary (ITC) was found to be the key factor in determining the ion signal intensity, which shows an increase of intensity by a factor of 100 from ITC temperature of 100–300 °C and could be attributed to more efficient desolvation and ionization. The characteristic fragment ion at *m*/*z* 72.99 was selected for quantitative analysis of TA at normalized collision energy of 50%, the optimal value applied during MS/MS analysis. Detection limit of 0.14 µg/m³ and a linear range of 0.2–2.97 µg/m³ are achieved. Satisfactory correlations between ion signal intensity and particle surface area (R^2 =0.969) and mass concentration (R^2 =0.967) were obtained. Although an equally good correlation was observed between signal intensity and particle surface area, the good correlation between signal intensity and particle mass concentration indicates that high solubility of TA ensures efficient dissolution of TA in the primary ESI droplets for further ionization.

Keywords Particulate organic matter \cdot Secondary nanoelectrospray ionization \cdot High-resolution mass spectrometry \cdot Normalized collision energy \cdot Ion transport capillary

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1 Introduction

Recent developments in real-time chemical characterization of aerosol particles have garnered attention for their high time resolutions (seconds to minutes) (Gallimore and Kalberer 2013; Li et al. 2015, 2017b; Brüggemann et al. 2015). Furthermore, compared to off-line methods, these real-time techniques can minimize artifacts due to condensation and evaporation, and avoid time-consuming sample pretreatment (Brüggemann et al. 2015). Advantages such as fast data acquisition and high sensitivity have made mass spectrometry (MS) among the most popular techniques for real-time aerosol chemical characterization (Pratt and Prather 2012; Li et al. 2015, 2017b). Aerosol mass spectrometers (AMS), which sample and analyze bulk aerosol particles, and, single-particle aerosol mass spectrometers (SPAMS), which sample and analyze individual particles, are the most widely used real-time mass spectrometric methods for aerosol analysis (Lee et al. 2013; Li et al. 2014, 2017b; Laskin et al. 2018; Passig et al. 2020). However, the extensive fragmentation of organic components by high-energy ionization confounds molecular identification in aerosol measurements despite the high sensitivity demonstrated by these instruments. Other widely used real-time techniques, such as online particle collection followed by ion chromatography (IC), are semi-continuous (typical time resolutions of tens of minutes). Moreover, IC-based methods are designed for water-soluble components (e.g., inorganic ions and very small carboxylic acids) and thus are not suitable for many hydrophobic organic compounds present in atmospheric particles (Li et al. 2017b).

Soft ionization MS, which does not cause extensive fragmentation, has proven to be a promising alternative for the molecular characterization of organic aerosols (Cooks et al. 2006; Huang et al. 2010; Gallimore and Kalberer 2013; Brüggemann et al. 2015). Several novel ionization techniques in recently developed ambient MS methods do not occur under high-vacuum conditions, which reduces potential artifacts caused by the evaporation of semi-volatile organics in particle phase. Hoffmann et al. (Hoffmann et al. 1998) studied particulate organics thermally vaporized within an atmospheric pressure chemical ionization (APCI) source and ionized by O_2^- ions produced via corona discharge. Direct analysis in real-time mass spectrometry (DART-MS) ionizes analytes using glow discharge plasma and has been successfully applied to the analysis of surface components on nanoparticles generated from a variety of atmospherically relevant organic compounds (Nah et al. 2013; Gross 2014).

In contrast to APCI and DART, secondary/extractive electrospray ionization technique generates primary reagent ions by applying conventional ESI (Gallimore and Kalberer 2013; Li et al. 2017a). Doezema et al. (2012) demonstrated that EESI-MS can detect both gaseous and particulate products from α -pinene ozonolysis in real time. Later, Gallimore and Kalberer showed that EESI-MS can be used to measure tartaric acid (TA) particles with sizes of 70–200 nm and mass concentrations of 3–600 μ g/m³ (Gallimore et al. 2017). Moreover, EESI-MS systems using a high-resolution mass spectrometer (mass resolution of 100,000 at m/z 400) have demonstrated unparalleled advantages over ESI-MS in chamber studies in both time resolution and quantification (Doezema et al. 2012; Gallimore et al. 2017). On the other hand, the ionization process is more complicated for the particles than for gases. For example, a recent study on the EESI mechanism revealed that surface energy and dissolution kinetics can influence the ionization process, and thus, that EESI might be more sensitive to particle surface layers than it is to the bulk particle composition (Kumbhani et al. 2018).

Despite the aforementioned progress, it remains necessary to lean the working principles of SESI and optimize relevant SESI-MS parameters to further improve the sensitivity, selectivity, and ease of use. In this study, a secondary nanoelectrospray ionization (Sec-nESI) source, which can achieve higher sensitivity with little solution consumption, was constructed and coupled with a high-resolution Quadrupole Orbitrap Mass Spectrometer (HRMS), then tested in measurements of particulate organics in real time. TA, a multi-functional dicarboxylic acid (Figure S1) found in atmospheric particles and believed to be formed secondarily from the photochemical oxidation of aliphatic and aromatic hydrocarbons, was used as a test compound (Röhrl and Lammel 2002; Claeys et al. 2004; Fu et al. 2014; Brent et al. 2014). Due to its low vapor pressure $(2.4 \times 10^{-12} \text{ mmHg})$ and high hygroscopicity, TA tends to partition almost entirely to the particle phase (Hildemann and Saxena 1996; Röhrl and Lammel 2002; Treuel et al. 2007). Factors affecting the detection of TA particles using Sec-nESI-HRMS are systematically investigated and discussed below.

2 Materials and Methods

2.1 Particle Generation and Characterization

Particles were produced by nebulizing TA solutions of $3.3-667 \mu mol/L$ with a commercial aerosol generator (Model 9302, TSI Inc., Shoreview, MN, US) in the laboratory (Fig. 1); the aerosol generator was operated under a constant flow of high-purity nitrogen gas (N₂, 99.999%), and the output flow rate was 2 L/min. TA (99%, Sigma-Aldrich, St. Louis, MO) was used as received (Figure S1 and Table S1). A TA solution of 670 µmol/L was prepared

Fig. 1 Schematic for the experimental detection of TA particles using an SMPS; the photo inset shows the relevant instrumentation in the laboratory



in ultrapure water (typically resistivity 18.2 M Ω cm at 25 °C), followed by stepwise dilution to lower concentrations. The TA solution concentrations and output flow rates were optimized in previous experiments to generate stable and reproducible particles (Figure S2).

The size distribution of the generated polydisperse particle population was characterized by a scanning mobility particle sizer (SMPS, Model 3938, TSI Inc., Shoreview, MN, US) (Figure S3). The sample and DMA sheath flow rates were set to 0.2 and 2 L/min, respectively (Fig. 1). Before entering the SMPS, particles were dried to 30% RH by passing through a diffusion dryer.

2.2 The Sec-nESI-HRMS System

A custom-built Sec-nESI source was developed using a commercially available nESI source (Nanospray Flex Ion Source, Thermo Fisher Scientific) (Fig. 2). Ultrapure water was used as the nESI solvent to generate the primary spray via a spray capillary with a flow rate of 200 nL/min by a nano-pump (NCS-3500 nano-pump, Thermo Fisher Scientific). A sample stream containing particles was introduced directly into the stainless-steel Sec-nESI chamber through a Teflon tube (2 mm I.D.) at a constant flow rate. The geometry of the Sec-nESI chamber has been reported previously (Li et al. 2017a, 2018). MS analyses were performed with a Quadrupole Orbitrap Mass Spectrometer (Q Extractive, Thermo Fisher Scientific). Tandem MS measurements



Fig. 2 Schematic of TA particle experimental detection using Sec-nESI-HRMS; the inset photo shows the instrumentation depicted in the schematic

were performed using higher energy collisional dissociation (HCD), with high-purity N_2 as the collision gas. Since deprotonated TA ([M–H]⁻) was observed in our preliminary test, m/z 149.01 was used as the precursor ion. MS and MS/ MS analyses were executed in negative ion mode at a resolution of 70,000.

3 Results and Discussion

3.1 TA Particle Characterization by SMPS and Sec-nESI-HRMS

Figure 3 shows the particle number-size distribution generated by nebulizing a TA solution of 67 µmol/L. The particle size range spanned 20-150 nm with a mode diameter of ~35 nm (Fig. 3a). The TA signal was seven times higher than the background under a particle sample flow rate of 2 L/min into the Sec-nESI chamber (Fig. 3b). The signal intensity remained constant over a 1-min sampling period (Fig. 3b) and generally the values of standard deviation and coefficient of variance of replicate measurements are 140,480.2558-248,788.8165 (*n* = 3) and 0.1239-0.2294(n=3), respectively. Similar results were achieved using particles from a 667 µmol/L TA solution (Figure S2d), for which good signal (at least one order of magnitude higher than the background) was observed even at lower sampling flow rates (e.g., 0.8 L/min; Figure S2d); this may be attributed to the higher particle number and mass concentrations generated (Figure S3). In contrast, TA was hardly detected using 3.3 µmol/L TA solution for particle generation (Figure S2b), as the number of particles in the chamber was not sufficient (Figure S3) to produce detectable amounts of TA ions. Thus, 67 µmol/L TA solutions were used in the following MS/MS investigations unless stated otherwise.

3.2 Factors Affecting the Performance of Sec-nESI

The Sec-nESI process involves production of primary ions and the delivery of aerosol particles (Fig. 2). Thus, factors In our experiments, the TA signal intensity was signifi-





including ESI voltage, ESI solution flow rate, and sample flow rate were investigated to optimize the Sec-nESI process. The ESI voltage, ESI solution flow rate, and sample flow rate parameters were initially set to the values typically used, i.e., -2.5 kV, 200 nL/min and 2 L/min, respectively.

Figure 4a and Figure S4 show a significant increase in signal intensity with an increase in ESI voltage from -2.0to -2.5 kV, indicating that satisfactory nESI process was achieved at -2.5 kV when ultrapure water was applied as the ESI solution. A higher ESI voltage might lead to more efficient formation of charged droplets and production of ionized molecules due to charge-transfer reactions, no matter which model of ESI process holds (Hermans et al. 2017). On the other hand, in the negative ion detection mode, nESI process becomes unstable and discharge may occur when ESI voltage is too high, for example explosion of droplets was observed at ESI voltage of -2.7 kV (Straub and Voyksner 1993). Thus, an ESI voltage of -2.5 kV was optimal in the range explored. A low ESI solution flow of 200 nL/min was found to produce the highest signal among the flow rates studied (Fig. 4b and Figure S4), probably due to smaller droplet formation that is easier for solvent evaporation. Therefore, the ESI solution flow rate of 200 nL/min was adopted as a sufficient ESI solution to generate primary ions. The use of ultrapure water can lead to higher ionization compared with the use of the mixtures of water and methanol (Martinez-Lozano Sinues et al. 2012; Lee and Zhu 2020).

The TA signal intensity increased with increasing sample flow rate. For example, the signal increased by a factor of seven when the sample flow rate was increased from 1.0 to 2.0 L/min (Fig. 4c and Figure S2). This increase in TA signal can be explained by the fact that more particles were delivered into the chamber at higher sample flow rates. The sample flow rate was set to 2 L/min to coordinate with the flow rate at the MS inlet interface (1.8 L/min).

3.3 Effects of ITC Temperature

cantly enhanced by increased ion transport capillary (ITC)



Fig. 4 Effects of changes in **a** ESI voltage, **b** ESI solution flow rate, **c** sample flow rate and **d** ITC temperature on measured signal intensity; note that a base-10 logarithmic scale is used for the *y* axis. Mark-

temperature from 100 to 300 °C (Fig. 4d and Figure S5), while the background noise was greatly reduced (Figure S5–S7), both of which contributed to higher signal-tonoise ratios. From a mechanistic perspective, a higher ITC temperature can enhance the evaporation and desolvation processes, thereby enhancing the ion–molecule reactions and increasing the delivery efficiency of ions into the mass analyzer for detection and, respectively (Zherebker et al. 2016; Martinez-Lozano Sinues et al. 2012). As a result, 300 °C was found to be the optimal value within the temperature range studied.

To further explore the influence of ITC temperature on TA detection, deprotonated TA at m/z 149.01 obtained at ITC temperatures of 300 and 150 °C was fragmented in the HCD cell at a normalized collision energy (NCE) of 35%, the default NCE value of the instrument. The observed peaks at m/z 59.01 (C₂H₃O₂⁻), m/z 87.01 (C₃H₃O₃⁻), m/z 103.00 (C₃H₃O₄⁻), and m/z 131.00 (C₄H₃O₅⁻) (Fig. 5 and Figure S6) are consistent with results from Gallimore and Kalberer, who used a LTQ Velos mass spectrometer (Gallimore and Kalberer 2013). The fragment ion at m/z

ers represent the average values and error bars represent the standard deviations of three replicate measurements

72.99 (C₂HO₃⁻) was reported by Brent et al., in which an ESI source and triple quadrupole mass analyzer were used (Fu et al. 2014). The peaks at m/z 59.98 (CO₃⁻), m/z 61.99 (NO₃⁻), and m/z 76.99 (CHO₄⁻) (Fig. 5 and Figure S6) are attributed to the ambient background. In the MS/MS spectrum obtained at 300 °C (Fig. 5 and Figure S7), background ions such as m/z 59.98 (CO₃⁻), m/z 61.99 (NO₃⁻) and m/z 76.99 (CHO₄⁻) are virtually nonexistent (Table S2). A TA fragmentation pathway is proposed in the supporting information based on the observed fragment ions (Figure S8).

3.4 NCE Optimization for MS/MS Analysis

Fragmentation efficiency can be optimized for a specific precursor ion by systematically varying the NCE. MS/MS analysis was performed at 300 °C for the deprotonated TA ion at m/z 149.01 under NCE values ranging from 10 to 70% (the default NCE value of the instrument is 35%). Signal intensities were obtained and compared for fragment ion at m/z 59.01, m/z 72.99, m/z 87.01, m/z 103.00, and m/z 131.00 (Fig. 6a).



Fig. 6 At an ITC temperature of 300 °C, **a** variations in the intensities of individual TA fragment ions (generated from a 67 μ mol/L TA solution) under various NCE values, and **b** an EIC of *m*/*z* 72.99 signal under an NCE of 50%. Markers in Fig. 7a represent the average values of 50%.

ues and error bars represent the standard deviations of three replicate measurements. The smooth curve in Fig. 7b is obtained using Percentile Filter

As shown in Fig. 6a, the TA ion intensity remained relatively constant when NCE was $\leq 35\%$, and the resulting fragment ion mass spectra are similar to each other (i.e., the ion abundance decreases in the following order: m/z 87.01, m/z72.99, m/z 59.01, m/z 103.00, and m/z 131.00). The intensities of ions at m/z 87.01 and m/z 72.99 increased with increasing NCE, suggesting that these two fragment ions may be formed more easily under higher NCE (Figure S8). At NCE values of 40–50%, the intensity of the deprotonated TA ion dropped significantly, which may be ascribed to higher fragmentation efficiency. This assumption is supported by the significant increases in intensity for ions at m/z87.01 and m/z 72.99 and the slight intensity enhancement at m/z 59.01. The highest intensity was achieved for fragment ions at both m/z 87.01 and m/z 72.99 using an NCE of 50%. The precursor ion and fragment ion intensities decreased at NCE $\ge 60\%$, which may be caused by the inability of the ions to remain in the HCD cell. Therefore, due to its sensitivity, specificity, and reproducibility, the fragment ion at m/z 72.99 was selected to investigate the detection limit and linear range for TA particle detection (Fig. 6b).

3.5 Detection Limit and Linear Range

Using the optimal parameters within the range explored, i.e., -2.5 kV ESI voltage, 200 nL/min ESI solution flow rate, 2.0 L/min sample flow rate, 300 °C ITC temperature, and 50% NCE value for the ion at *m*/*z* 72.99, we investigated the detection limit and linear range using TA particles in the mass concentration range of 0–2.97 µg/m³ (Figures S9 and S10). Figure 7a shows the relationship between the signal intensity at *m*/*z* 72.99 and the TA



Fig. 7 Signal intensity of fragment ion at m/z 72.99 against **a** particle number concentration (C_{Number}), **b** surface area concentration ($C_{\text{Surface area}}$), and **c** mass concentration (C_{Mass}) of aerosol samples generated from 0, 3.3, 6.7, 13, 33, 67, and 133 µmol/L TA solutions

under an ITC temperature of 300 $^\circ C.$ Markers represent the average values and error bars represent the standard deviations of three replicate measurements

particle number concentration measured by the SMPS. A non-linear relationship is observed ($R^2 = 0.905$, Fig. 7a inset), which may arise from the fact that the mode diameter and the width (or deviation) of the log-normal distribution of the polydisperse particles for different concentrations might change, and thus the number concentration does not reflect the TA ionized. Figure 7b shows the relationship between ion signal and TA particle surface area concentration, which is more linear ($R^2 = 0.969$, inset of Fig. 7b) than the relationship between ion signal and number concentration; this suggests that the ionization process may have involved a surface process. Figure 7c shows the relationship between ion signal and TA particle mass concentration calculated from the SMPS size distribution by assuming particle sphericity and a TA density of 1790 kg/ m³ (Table S1). In the 0.2–2.97 μ g/m³ mass concentration range (Fig. 7c), the coefficient of determination (R^2) is $0.967 \text{ (slope} = 2.96 \times 10^6 \text{, intercept} = 3.61 \times 10^5 \text{)}$. Notably, the ion signal intensity correlates very well with both particle surface area ($R^2 = 0.969$) and mass ($R^2 = 0.967$) (Fig. 7b and Fig. 7c). Although an equally good correlation was observed between signal intensity and particle surface area, the good correlation between signal intensity and particle mass concentration indicates that high solubility of TA ensures efficient dissolution of TA in the primary ESI droplets for further ionization. As a result, the current technique has a great potential in quantifying the organics in the entire submicron particle even if the particle is not homogeneous, which is common in atmospheric particles.

Using pure water background signal measurements, a detection limit of 0.14 μ g/m³ was estimated for TA particles. The LOD is calculated using a signal-to-noise ratio (S/N) of 3 according to LOD = $3 \times \sigma_{\text{blank}}$ /sensitivity, where σ_{blank} is the standard deviation of the background noise level measured at 0 ppb of gas sample (i.e., pure N₂). The detection limit obtained in this study is comparable to that reported previously (0.22 μ g/m³) (Gallimore and Kalberer 2013). With analytical merits including high time resolution, high sensitivity, high selectivity, and low detection limit, this new method is well suited for the chemical characterization and quantification of laboratory-generated organic aerosols and chamber-derived SOA.

4 Conclusions

We investigated the real-time measurement of laboratorygenerated TA particles using a custom-built Sec-nESI source coupled with a HRMS. Parameters involved in the Sec-nESI process, including ESI voltage, ESI solution flow rate, sample flow rate, and ITC temperature, were optimized. Using a specific fragment ion for qualitative and quantitative analysis and NCE for MS/MS analysis, TA can be quantified over a linear range of 0.2–2.97 µg/ m^3 with high sensitivity (detection limit = 0.14 $\mu g/m^3$). Among the parameters investigated, the ITC temperature had the most pronounced impact on ion signal intensity, which may be attributed to more efficient desolvation and ionization. In MS/MS measurements, different fragments showed quite different dependence on the applied NCE, implying that the NCE should also be tuned when exploring organic compounds with unknown structures. This work successfully demonstrates the highly sensitive detection of a multi-functional organic compound using sec-nESI-HRMS, demonstrating that secondary ionization using primary ions generated from either conventional ESI or nESI (as in this study) holds promise for the identification and quantification of organic particles in laboratory studies.

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