

Fungicide resistance screening of a *Botrytis cinerea* population from Australian vineyards

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Background

Botrytis cinerea, which causes botrytis bunch rot, is one of the most important grapevine diseases. Fungicides are integral for the control of *B. cinerea*. Resistance to most single-site fungicides is geographically widespread. In this study, phenotypic and genotypic resistance screening was undertaken on a population of *B. cinerea* isolates.

Methods

504 isolates were collected between 2020 – 2024 from 53 vineyards across WA, SA, Vic, NSW and the ACT. Isolates were screened on discriminatory concentrations of fungicides from FRAC groups 9, 12 and 17. Resistance associated genes *Pos5* (group 9), *Mdl1* (group 9) and *Erg27* (group 17) were screened.

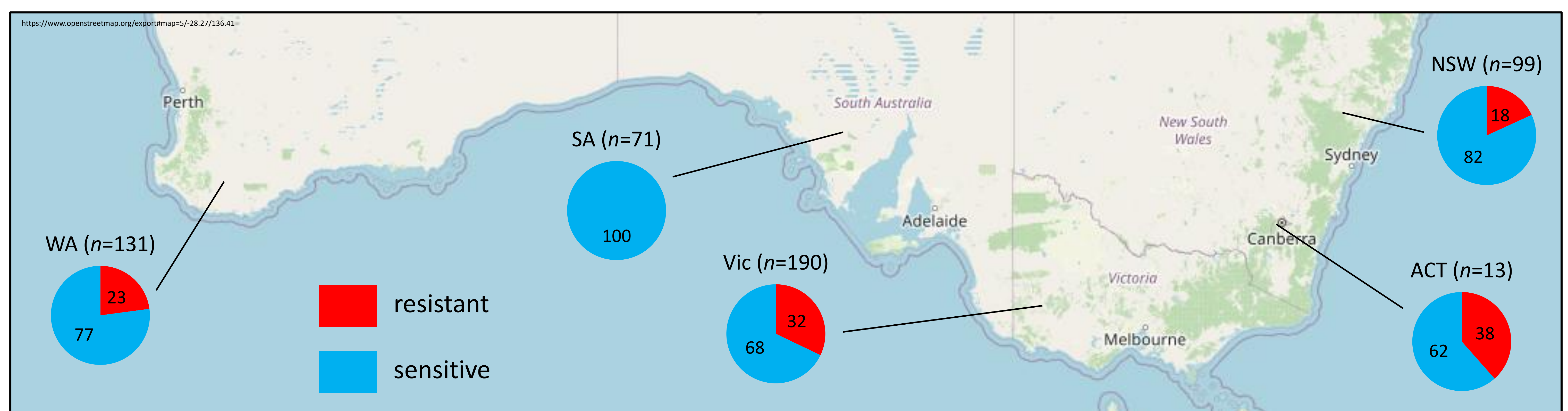


Fig. 1. Total resistance frequencies for WA, Vic, NSW and ACT. n indicates number of isolates tested. Red indicates resistance, blue indicates sensitive.

Results

Total resistance frequencies for WA, SA, Vic, NSW and ACT were 23, 0, 32, 18 and 38%, respectively (Fig. 1). Total (national) resistance frequencies per fungicide were 14.5, 9.7 and 8.7% for group 9, 12 and 17, respectively (Fig. 2). Most isolates were resistant to group 9 only, group 12 only or both group 9 and 17 simultaneously (Fig. 3a). In *Pos5* and *Mdl1*, the changes V273I, P293S, P319A or L142F/V and E407K or G408V were identified, respectively (Fig. 3b). In *Erg27*, the changes F412C/I/S/V were found (Fig. 3c). The predominant genotypes for group 9 and 17 resistance were L412F and F412S, respectively (Fig. 3b-c).

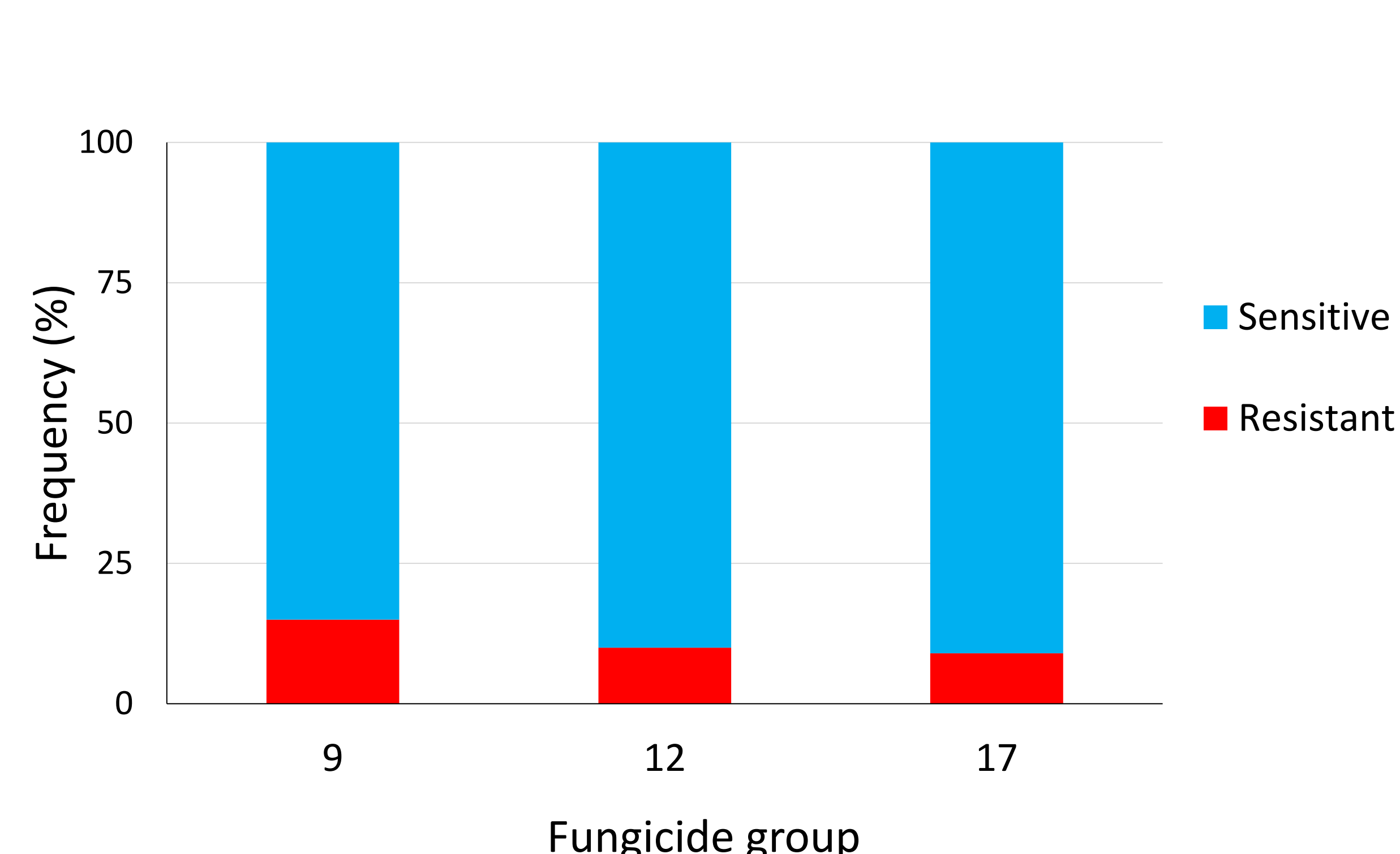


Fig. 2. Total resistance frequencies per fungicide group (N = 504 isolates)

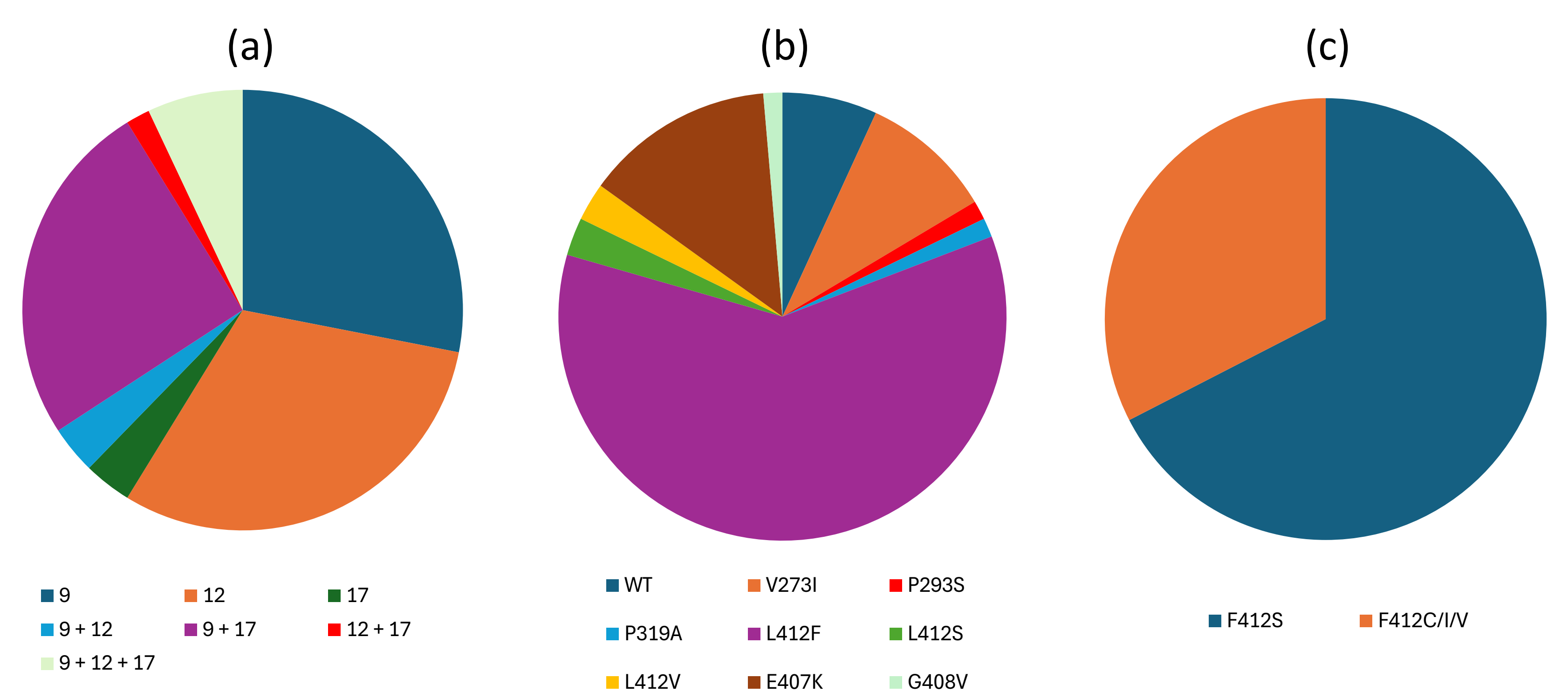


Fig. 3. Frequency of single and multi-resistance phenotypes in resistant isolates (n=115) (a). Frequency of group 9 (b) and group 17 (c) associated genotypes

Conclusions

These results provide valuable information on the resistance status of *B. cinerea* in Australian vineyards and could assist in improving resistance management strategies. Further monitoring of resistance to critical Botryticidal chemistry is essential to improving current resistance management strategies.